

Dose–response effects of equine chorionic gonadotrophin (eCG) and human chorionic gonadotrophin (hCG) on early embryonic development and viable pregnancy rate in rats

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The present study examined the dose–response effects of eCG treatment alone and in combination with various doses of hCG on early embryonic development *in vivo* and viable pregnancy rate in rats. Mated female Wistar rats were treated with eCG alone (0, 10, 20 or 40 iu), or with 20 iu eCG in combination with various doses of hCG (10, 20, 40 or 80 iu) administered 48 h later. The animals were killed on days 2, 3, 4, 5 or 14 of pregnancy and the numbers of embryos and fetuses recovered were scored. All rats treated with 0 or 10 iu eCG were pregnant. The pregnancy rate was reduced from 62.5% on day 2 to 25% on day 14 and from 31% on day 2 to 10% on day 14 in the groups treated with 20 and 40 iu eCG, respectively. The

reduction in pregnancy rate induced by 20 iu eCG was negated by the increasing doses of hCG used. A 100% pregnancy rate was noted on days 2 and 3 in the groups treated with doses of hCG between 10 and 80 iu and from day 2 to day 4 in the groups treated with doses of hCG between 20 and 80 iu. However, a higher viable pregnancy rate was observed only in the group treated with 10 iu hCG compared with the group treated with 20 iu eCG and 0 iu hCG. These results imply that hyperstimulation of rats with high doses of eCG compromises pregnancy rate and markedly reduces litter size and that the addition of hCG is required for complete ovulation, which results in higher embryo yield and a delay in early embryo demise.

Introduction

The low pregnancy rates in farm and laboratory animals, despite high oocyte yield after hyperstimulation with exogenous administration of eCG, could be the result of a high incidence of over-maturity (Moor *et al.*, 1985) and chromosomal aberrations in oocytes (Hansmann *et al.*, 1988; Badenas *et al.*, 1989; Yun *et al.*, 1989). It has also been suggested that the disrupted pattern of steroidogenesis after hyperstimulation might create an environment that is hostile to normal and sustained embryonic development, leading to failed pregnancy in immature rats (Miller and Armstrong, 1981). Therefore, it is possible that low pregnancy rates after hyperstimulation with gonadotrophins might be attributable to poor oocyte quality and a compromised microenvironment that is hostile to early embryonic development.

hCG is given routinely to induce ovulation, as a substitute for the LH surge; thus, poor pregnancy outcome could also be attributed to the dose of hCG used (Ertzeid and Storeng, 1992; Ertzeid *et al.*, 1993). The aim of the present study was to examine the dose–response effects of eCG treatment alone and in combination with various doses of hCG on early embryonic development *in vivo* and on viable pregnancy rate using a rat model.

Materials and Methods

Animals

Mature female Wistar rats (200–250 g body weight, aged 7–8 weeks) were housed in the Animal Holding Unit at a temperature of 28–30°C and kept under a 12 h light:12 h dark photoperiod. The animals had free access to water and food chow (Glen Forrest Stockfeeders, Glen Forrest, WA). Fertile Wistar male rats were used for mating.

Gonadotrophin administration

A total of 706 mature female rats was injected i.p. with 0, 10, 20 or 40 iu eCG (Sigma, St Louis, MO) in 0.5 ml normal saline between 12:00 and 12:30 h on the day of early dioestrus, followed by 0.5 ml saline 48–52 h later. In some animals injected initially with 20 iu eCG, instead of the saline injection 48–52 h later, separate groups were administered with various doses of hCG (10, 20, 40 or 80 iu; Pregnyl; Oragon, Holland). Each female rat was caged with a fertile male overnight for mating (day 0). Successful mating was confirmed by the presence of spermatozoa in vaginal smears taken on the next day, which was designated as day 1 of pregnancy. Only rats that had mated successfully were included in the study.

Embryo retrieval

Groups of pregnant rats were killed (by excessive inhalation of carbon dioxide) between 09:30 and 10:00 h

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on days 2, 3, 4 or 5 of pregnancy. The oviducts and uterine horns were excised and flushed with T6 medium (Ng, 1986) to retrieve embryos, which were examined and scored under a phase-contrast microscope. Embryos with regular blastomeres, intact zonae pellucidae and even granulation of the cytoplasm were classified as embryos with normal morphology, whereas embryos with irregular blastomeres, or those that were degenerating or fragmented were classified as embryos with abnormal morphology.

In other groups, pregnancy was allowed to progress up to day 14, at which time the animals were killed (by excessive inhalation of carbon dioxide) to check the viability of the pregnancy. The total number and number of normal, abnormal and resorbed fetuses (a black spot with no visible fetus) were counted. Only the average total number and mean number of normal embryos and fetuses were used for the statistical analyses.

Statistical analysis

SPSS statistical software was used to analyse the experimental data. The mean total and average numbers of normal embryos and fetuses retrieved from the various treatment groups were analysed using Kruskal–Wallis or Mann–Whitney tests. The chi-squared test was used to analyse the pregnancy rates in different treatment groups. Differences were considered to be statistically significant when the P value was ≤ 0.05 .

Results

Pregnancy rates

For the analyses, a positive pregnancy was defined as the presence of at least one embryo or fetus. In the control group and the group treated with 10 iu eCG, all rats (100%) were pregnant on days 2–5 and day 14 of pregnancy (Fig. 1a). However, in the groups treated with 20 and 40 iu eCG, the pregnancy rate was compromised as early as day 2 and the pregnancy rates were 62.5% and 31%, respectively. These rates were significantly reduced (chi-squared, $P < 0.05$) as pregnancy progressed, and reached a minimum of 25% and 10% on day 14 in the groups treated with 20 and 40 iu eCG, respectively (Fig. 1a).

Various doses of hCG were administered to rats hyperstimulated with 20 iu eCG. The reduced pregnancy rates on days 2–4 in the group treated with 20 iu eCG + 0 iu hCG were restored to 100% by doses of hCG between 20 and 80 iu (chi-squared, $P < 0.05$; Fig. 1b). In the group treated with 10 iu hCG, the restoration of pregnancy rate to 100% was noted on days 2 and 3 (Fig. 1b). However, by day 14, the pregnancy rates in all groups treated with hCG were not different from those in the group treated with 0 iu hCG.

Number and quality of embryos

The administration of 10 iu eCG significantly increased the average total number of embryos compared with that in

the control group on days 2–5 of pregnancy, and the number of embryos with normal morphology on days 3–5 (Table 1). However, the increases in the number of embryos were not reflected in an increased number of viable fetuses on day 14 of pregnancy.

As with pregnancy rates, the average total number of embryos and number of embryos with normal morphology in the groups treated with 20 and 40 iu eCG were markedly reduced from day 2 to day 14, particularly in the group treated with 40 iu eCG (Table 1). Embryo demise occurred at an early stage, that is, by day 2, in the group treated with 40 iu eCG (Table 1).

Significantly larger numbers of embryos with normal morphology were observed in the groups given doses of hCG between 10 and 80 iu (Table 2). The numbers of embryos in all groups treated with hCG were significantly larger than those in the group treated with 0 iu hCG. Increases in the total number of embryos and number of embryos with normal morphology in the groups treated with hCG ranged from two- to ninefold. However, by day 5 and day 14, the total number and mean number of normal embryos and fetuses in the groups treated with hCG had decreased significantly and were not significantly different from the corresponding values in the group treated with 0 iu hCG.

Discussion

The present study was undertaken to ascertain the effects of eCG administered to stimulate multiple folliculogenesis on early embryonic development and viable pregnancy rate in rats. The aims were: (i) to determine the dose–response effects of eCG on early embryonic development; (ii) to determine whether there is an optimal dose for oocyte quality and embryonic development; and (iii) to determine whether additional doses of hCG are required for the complete ovulation of the expanded cohort of preovulatory follicles and improvement in embryonic development and viability of pregnancy.

The largest number of embryos was retrieved from the group of rats treated with 10 iu eCG, which was about 56% higher than the number retrieved from the control group. Furthermore, increasing the dose of eCG to ≥ 20 iu did not result in a higher pregnancy rate and yield of embryos and fetuses from day 2 to day 14 of pregnancy. A high degree of embryo loss was observed from day 2 to day 5 in the eCG-treated group: the percentage of embryos with normal morphology at day 2 that developed into viable fetuses was 77% and 23%, respectively, in the groups treated with 10 and 20 iu eCG. The reduction in the number of viable fetuses in the group treated with 10 iu eCG (mildly stimulated) implies that embryos with normal morphology did not necessarily have the potential to develop normally and lead to sustained pregnancy. Although many studies have shown that embryo loss is attributed to high oestrogen concentrations after hyperstimulation (Walton and Armstrong, 1981), measurement of oestrogen concentrations

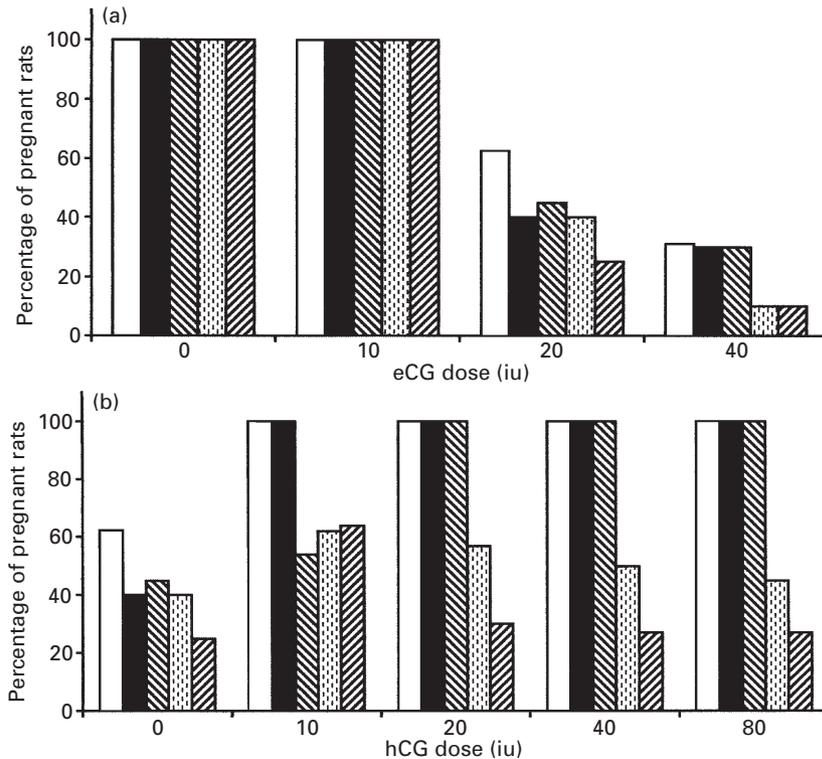


Fig. 1. Effects of (a) eCG alone (0–40 iu) and (b) 20 iu eCG in combination with various doses of hCG (0, 10, 20, 40 and 80 iu) on pregnancy rates in rats on days 2 (□), 3 (■), 4 (▨), 5 (▩) and 14 (▮) of pregnancy. Percentage of pregnant rats = number of pregnant rats/total number of rats in the treatment group. (a) Pregnancy rates in the groups treated with 20 and 40 iu eCG were significantly reduced from day 2 to day 14 of pregnancy (*chi-squared, $P < 0.05$). (b) Within groups treated with 0, 10, 20, 40 and 80 iu hCG, pregnancy rates were significantly reduced from day 2 to day 14 (chi-squared, $P < 0.05$). Among groups treated with doses of hCG between 0 and 80 iu, pregnancy rates were significantly different on days 2–5 of pregnancy (chi-squared, $P < 0.05$), but there were no differences on day 14 of pregnancy.

at day 1 showed that the concentrations in the group treated with 10 iu eCG ($101 \pm 6.6 \text{ pg ml}^{-1}$) were not different from those in the control group ($98 \pm 3.3 \text{ pg ml}^{-1}$) (C. F. Tain and H. H. V. Goh, unpublished), indicating that, in the present study, embryo loss may be due, at least in part, to compromised oocyte or embryo quality.

It is evident that the embryonic development was compromised in rats hyperstimulated with doses of eCG ≥ 20 iu. In the group treated with 40 iu eCG, an average of only 1.5 embryos was retrieved on day 2, which may be the result of ovulation failure, as administration of high doses of eCG suppresses endogenous LH secretion (Greenwald and Roy, 1994). However, other problems may be associated with the use of high doses of eCG. One possibility is that the long half-life and high LH-like activities of eCG may be detrimental to oocytes and early preimplantation embryos (Walton and Armstrong, 1981). The high LH-like activity of

eCG may cause over-maturation of the oocytes before they are ovulated. In other studies, embryo demise has been attributed to impaired oocyte quality (Moor *et al.*, 1985), genetically abnormal oocytes (Hansmann *et al.*, 1988; Badenas *et al.*, 1989; Yun *et al.*, 1989) and a hostile hormonal milieu in the reproductive tract induced by hyperstimulation (Miller and Armstrong, 1981).

The results from the present study indicate that the optimal dose for stimulating mature rats is about 10 iu eCG and that eCG doses of ≥ 20 iu (given alone) impair early embryonic development, which eventually leads to low viable pregnancy rates. Furthermore, it was shown that embryo demise in hyperstimulated animals occurred early in pregnancy, that is, at the preimplantation stage. It is possible that embryo demise could be the result of a hostile environment created by the hyperstimulation (Walton *et al.*, 1983) and this possibility is the subject of ongoing research

Table 1. Effects of administration of eCG on pregnancy outcome, the mean total number of embryos and the number of normal embryos or fetuses (mean \pm SEM) retrieved at days 2, 3, 4, 5 and 14 of pregnancy in rats

Day of pregnancy		Dose of eCG (iu)			
		0	10	20	40
2	<i>n</i>	19/19	10/10	10/16	4/13
	Embryos*	11.6 \pm 0.4 ^a	19.5 \pm 4.1	14.4 \pm 3.9	1.5 \pm 0.8 ^b
	Normal embryos [†]	10.4 \pm 0.5	15.1 \pm 3.4	9.9 \pm 2.8	0.7 \pm 0.3 ^b
3	<i>n</i>	18/18	10/10	6/15	3/10
	Embryos	11.3 \pm 0.5 ^a	16.4 \pm 2.1	9.0 \pm 3.6	6.6 \pm 3.8 ^a
	Normal embryos	9.7 \pm 0.5	14.5 \pm 2.0 ^c	8.0 \pm 3.4	5.4 \pm 3.6 ^d
4	<i>n</i>	18/18	10/10	5/11	3/10
	Embryos	11.2 \pm 1.1	17.7 \pm 1.8 ^c	3.5 \pm 1.3 ^d	2.9 \pm 2.0 ^d
	Normal embryos	10.1 \pm 1.0	15.2 \pm 1.9 ^c	2.8 \pm 1.1 ^d	2.4 \pm 1.7 ^d
5	<i>n</i>	19/19	11/11	5/12	1/10
	Embryos	10.3 \pm 2.3 ^a	15.5 \pm 1.7	7.8 \pm 2.5	0.9 \pm 0.9 ^b
	Normal embryos	9.7 \pm 0.6	14.0 \pm 1.4 ^c	5.4 \pm 1.7	0.7 \pm 0.7 ^b
14	<i>n</i>	20/20	10/10	3/12	1/12
	Embryos	12.6 \pm 0.4	14.6 \pm 1.8	4.1 \pm 2.0 ^d	1.0 \pm 1.0 ^d
	Normal embryos	11.5 \pm 0.5	11.6 \pm 1.3	2.3 \pm 1.2 ^{de}	0.6 \pm 0.6 ^b

n: number of pregnant rats/total number of rats.

*Embryos: mean total number of embryos; [†]normal embryos: mean number of embryos with normal morphology.

^aSignificantly lower than corresponding values in the group treated with 10 iu eCG.

^bSignificantly lower than corresponding values in all other groups.

^cSignificantly higher than corresponding values in all other groups.

^dSignificantly lower than corresponding values in the groups treated with 0 and 10 iu eCG.

^eSignificantly lower than corresponding values on day 2 in the group treated with 20 iu eCG.

Values are statistically significant when $P \leq 0.05$.

Table 2. Effects of administration of 20 iu eCG and various doses of hCG on pregnancy outcome, the mean total number of embryos and the number of embryos with normal morphology (mean \pm SEM) retrieved at days 2, 3, 4, 5 and 14 of pregnancy in rats

Day of pregnancy		Dose of hCG (iu)				
		0	10	20	40	80
2	<i>n</i>	10/16	10/10	10/10	10/10	10/10
	Embryos*	14.4 \pm 3.9 ^e	34.9 \pm 7.6 ^a	28.8 \pm 3.8 ^b	34.2 \pm 6.3 ^b	35.6 \pm 7.6 ^b
	Normal embryos [†]	9.9 \pm 2.8 ^e	27.3 \pm 4.9 ^a	26.2 \pm 3.4 ^b	29.2 \pm 4.6 ^b	26.6 \pm 6.6 ^b
3	<i>n</i>	6/15	10/10	10/10	11/11	10/10
	Embryos	9.0 \pm 3.6 ^c	28.6 \pm 5.0 ^a	22.9 \pm 4.1 ^b	36.5 \pm 6.8 ^b	42.1 \pm 6.6 ^b
	Normal embryos	8.0 \pm 3.4 ^e	26.6 \pm 4.7 ^a	20.2 \pm 3.9 ^b	28.2 \pm 5.0 ^b	33.9 \pm 5.7 ^b
4	<i>n</i>	5/11	8/14	11/11	10/10	11/11
	Embryos	3.5 \pm 1.3 ^f	10.9 \pm 3.6 ^d	18.4 \pm 3.7 ^b	30.1 \pm 4.4 ^b	31.7 \pm 3.2 ^b
	Normal embryos	2.8 \pm 1.1 ^f	9.0 \pm 3.1 ^f	17.4 \pm 3.6 ^b	25.2 \pm 4.5 ^b	25.0 \pm 4.1 ^b
5	<i>n</i>	5/12	9/14	8/14	6/12	5/11
	Embryos	7.8 \pm 2.5	6.2 \pm 2.0	4.8 \pm 1.5	6.3 \pm 2.3	3.9 \pm 1.6
	Normal embryos	5.4 \pm 1.7	5.5 \pm 1.7	4.6 \pm 1.4	6.1 \pm 2.2	3.9 \pm 1.6
14	<i>n</i>	3/12	7/11	3/10	3/11	3/11
	Embryos	4.1 \pm 2.0	10.8 \pm 2.8	7.1 \pm 3.4	8.7 \pm 3.4	6.1 \pm 3.2
	Normal embryos	2.3 \pm 1.2	7.9 \pm 2.0	3.7 \pm 2.5	3.5 \pm 2.2	3.2 \pm 2.1

n: number of pregnant rats/total number of rats.

*Embryos: mean total number of embryos; [†]normal embryos: mean number of embryos with normal morphology.

^aSignificantly higher than the value at days 4, 5 and 14 within the same hCG group.

^bSignificantly higher than the value at days 5 and 14 within the same hCG group.

^cSignificantly lower than corresponding values in the groups treated with 10, 40 and 80 iu hCG.

^dSignificantly lower than corresponding values in the groups treated with 40 and 80 iu hCG.

^eSignificantly lower than corresponding values in all other groups.

^fSignificantly lower than corresponding values in the groups treated with 20, 40 and 80 iu hCG.

Values are statistically significant when $P \leq 0.05$.

in our centre, which includes transferring two-cell embryos from hyperstimulated rats into non-stimulated pseudo-pregnant rats.

This is the first study to show that administration of hCG (10–80 iu) to rats hyperstimulated with eCG can increase embryo yield and improve pregnancy rates, which is in contrast to the studies by Ertzeid and Storeng (1992) and Ertzeid *et al.* (1993). The various doses of hCG administered after injection of 20 iu eCG induced between two and five times more embryos on days 2 and 3 than the corresponding values in the group treated with 20 iu eCG alone. This finding implies that, in the rats treated with 20 iu eCG alone, the endogenous LH surge was probably inadequate to induce ovulation. Therefore, the administration of exogenous hCG can trigger the maturation and ovulation of the expanded cohort of follicles. However, there were no dose-related increases in the number of embryos retrieved with increasing doses of hCG.

These findings imply that additional hCG not only resulted in more complete ovulation of the expanded cohort of preovulatory follicles, but that the hCG resulted in an increase in the quality of the oocytes and subsequent embryos such that more rats were able to sustain pregnancy beyond the implantation stage. However, the positive effect of administration of 10 iu hCG was not enhanced when the dose was increased. This finding implies that the addition of hCG was unable to reverse the embryo demise caused by high doses of eCG administered earlier, even though the cohort of early embryos was much larger.

The results of the present study indicate that rats hyperstimulated with a high dose of eCG have compromised pregnancy rates and markedly reduced litter sizes. Furthermore, in hyperstimulated cases, the addition of high doses of hCG might be required for complete ovulation as well as for improving the maturation of the expanded cohort of preovulatory follicles. There also appears to be a critical threshold value for the additional hCG used in rats.

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