Non-invasive assessment of oestrous cycles and evaluation of reproductive seasonality in the female wild black rhinoceros (Diceros bicornis minor)

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Female wild black rhinoceroses in Zimbabwe were monitored non-invasively using faecal progesterone metabolite analysis and observation of reproductive behaviour. A postpartum period of reproductive inactivity of at least 4 months, followed by a period of 4–7 months of oestrous cyclicity, was detected in six multiparous females. Three-quarters of the oestrous cycles (n = 21) had a total duration (mean ± SEM) of 26.8 ± 1 days. Other types of cycle were characterized either by an extended luteal phase, lasting on average twice as long as the normal cycle, or by an extended follicular phase. These extended cycles may have resulted from early embryo loss and heat stress. Female rhinoceroses did not conceive before 8 months after giving birth and some females (n = 2) most likely aborted after 3.0–3.5 months of gestation. The detected period of cyclic oestrus occurred between May and March in females (n = 9), and there was a 3 month extended interoestrous interval in nulliparous females during the period of decreasing daylengths that can be presumed to be the period of poorest fertility for the black rhinoceros under tropical latitudes. In contrast, the period of optimum fertility in the Southern hemisphere coincided with the late spring and early summer, and corresponded to the early rainy season. As a result, a higher incidence of births was detected in the late rainy season, providing the lactating female with the most suitable environment in terms of nutritional requirements.

Introduction

The Rhinocerotidae appeared during the early Eocene, some 50 million years ago, and represent one of the most ecologically diverse and widespread families of mega-herbivores (Owen-Smith, 1988). Five rhinoceros species survive today, all of which are endangered, and comprise a total of <15 000 animals (Foosse and Van Strien, 1997; Emslie and Brooks, 1999). The population of black rhinoceroses was estimated to be only 2600 in 1997 (Emslie and Brooks, 1999).

As a result of the demand for rhinoceros horn, the remaining rhinoceros populations are exposed to a permanent threat of poaching. These populations are small and isolated, making them also vulnerable to stochastic environmental, demographic or genetic factors (Bride et al., 1996). Remaining populations must present optimum breeding output, both in the wild and in captivity, which is far from being achieved (Lindemann, 1982; Kock and Garnier, 1993; Brett, 1998; Gölténboth, 1999; Garnier, 2001).

The first obstacle in attaining optimum reproductive performances in black rhinoceroses is the difficulty of assessing reproductive success in an animal with such a long generation period, solitary habits and which lives in dense habitats at low densities (Ritchie, 1963; Goddard, 1967; Shenkel and Shenkel-Hulliger, 1969; Joubert and Eloff, 1971; Adcock, 1994). As a result of this difficulty, simple reproductive parameters are generally based on a limited number of observations and can give information only retrospectively. Measurements neither reflect individual variations nor indicate a potential decline in fertility. Such information is also difficult to interpret because so little is known of reproductive features of the rhinoceros.

The information available on oestrous cycles and seasonality in free-ranging black rhinoceroses is rare and anecdotal. One study established that interoestrous intervals ranged from 26 to 46 days (Hitchins and Anderson, 1983). Oestrous behaviour in the wild has been described by Brett et al. (1989), Goddard (1966), Hitchins and Anderson (1983) and Shenkel and Shenkel-Hulliger (1969) and the period from the first day of attendance by the bull to the time of copulation is reported to be 6–7 days and the duration of receptivity only 1–3 days. Some peaks in parturition have been identified in southern Africa at different times of the year (Joubert and Eloff, 1971; Hall-Martín and Penzhorn, 1977; Hitchins and Anderson, 1983). In east Africa, little or no evidence of reproductive seasonality has been found (Ritchie, 1963; Goddard, 1967; Shenkel and Shenkel-Hulliger, 1969).

In captivity, individual interoestrous intervals of rhinoceroses ranged from 17 to 83 days (Lindemann, 1982). Hindle et al. (1992) used an enzymeimmunoassay for
measuring urinary 20α-dihydroprogesterone to establish that an average cycle duration of 21 days was represented by a 16–17 day luteal phase with a 3–4 day follicular phase in two captive females. Radcliffe et al. (2001) observed an interovulatory period of 26 days during two cycles by combining ultrasound examination and faecal progesterone metabolite analysis.

The collection of faecal samples represents a practical approach for monitoring reproduction under free-ranging conditions, and the analysis of faecal steroid metabolites has been used to assess breeding activity in an increasing number of species, especially in captivity (for a review, see Schwarzenberger et al., 1996a). In the wild, the use of faecal steroid analysis has been limited to populations or social groups in a very limited number of species (see Monfort, in press).

In rhinoceros species, faecal progestagen analysis has permitted the monitoring of gestation and diagnosis of pregnancy in both wild and captive black and white rhinoceroses (Schwarzenberger et al., 1993, in press; Berkeley et al., 1997; Radcliffe et al., 1997; Garnier et al., 1998a; Patton et al., 1999), and in captive Indian rhinoceroses (Schwarzenberger et al., 2000). The analysis of faecal progestagen has also allowed the assessment of luteal activity in captivity in white (Patton et al., 1999), Indian (Schwarzenberger et al., 2000) and Sumatran rhinoceroses (Heistermann et al., 1998).

The main objective of the present study was to characterize the reproductive cycles and patterns of seasonality in black rhinoceroses in their natural environment. In particular, the study aimed to: (i) monitor oestrus cycles during the intergestation period in wild females using faecal progestagen analysis; and (ii) assess the influence of seasonality on reproductive activity.

### Materials and Methods

#### Study site

The study was conducted in Zimbabwe in two different sites: the Save Valley Conservancy (SVC; 20°E, 31°S), which covers 3387 km² in the south-east of the country, and the smaller Imire game ranch (18°E, 31°S), which covers 300 km² in the Midlands area. The climate in Zimbabwe consists essentially of a rainy season lasting 5 months (November until April), and the average rainfall is lower in the south (300–500 mm) than in the Midlands (600–800 mm). The dry season can be divided into a cold period (May to mid-August) and a hot period (mid-August to October). Temperatures reach peak values in October.

#### Animals and data collection

The study animals in the SVC were located in the southern section of the area, where a resident sub-population originated from 20 black rhinoceroses translocated there in 1986–1988. This sub-population consisted of 43 animals in March 1999; it had a mean adult sex ratio of 1.2 males:1 female and a mean proportion of adults of 56% between 1994 and 1999, during which time there was a recruitment rate of 7.2% (Garnier, 2001).

Individual reproductive monitoring was undertaken in six wild females, including four founder animals (Pukwani, Bulawayo, Netsai and Sirica) and two first generation animals (Jete and Sara) after they gave birth. The two first generation animals were 6 and 10 years old, respectively, when parturition was recorded during the study, and the founder animals were estimated to be between 25 and 37 years old at that time. The female Pukwani belonged to a different community of black rhinoceroses compared with

### Table 1. Types and periods of data collection in each study area in the present study of female black rhinoceroses

<table>
<thead>
<tr>
<th>Location</th>
<th>Study animals</th>
<th>Type of data collected and date collected</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Animals</td>
<td>Year of birth*</td>
</tr>
<tr>
<td></td>
<td>Other females (n = 8)</td>
<td></td>
</tr>
<tr>
<td>Imire</td>
<td>Amber, Mvu, D]</td>
<td>1987</td>
</tr>
</tbody>
</table>

Dates indicate periods during which each type of data was collected.

*The age of the first four animals from the Save Valley Conservancy had been determined during previous immobilization procedures by evaluating the toothwear pattern, whereas the dates of birth of the other study animals were recorded.

† Faecal samples were collected every 2–3 days during the dry season and every 4–5 days during the rainy season.

‡ Reproductive behaviour was recorded at the same frequency as for the collection of faecal samples.

§ Birth dates were recorded by weekly sightings.
Returned into oestrus within 1 week of removal of the calf (Garnier, 2001); therefore, only the first parturition was recorded in each female, as artificial weaning of the calf might have interfered with naturally occurring periods of reproductive inactivity.

Both wild and semi-wild situations are characterized by a natural breeding system (Bride et al., 1996) and both wild and semi-wild females are hereafter referred to as ‘wild’.

One captive black rhinoceros female (Rosie) kept at London Zoo was also studied. She had been hand-reared and was estimated to be 8 years old during the study. She was paired with a male in an outdoor paddock during periods of sexual receptivity, when the male showed increased interest towards her, and when climatic conditions (temperature and humidity) were favourable. Both hormonal and behavioural data were collected every day for 9 months (Table 1). This female did not produce a calf and was subsequently transferred to another institution.

**Hormonal analysis**

Faecal samples were collected by breaking apart a fresh faecal pellet and placing an amount equivalent to a handful into polythene bags using a wooden stick. A sample was considered to be fresh when the superficial layer of faecal pellets was still wet and no insect contamination had occurred (Garnier et al., 1998b). Samples were placed in a thermos flask that had been stored at 4°C until they could be stored in an electrical cool box in the car, which occurred between 30 min and 4 h after the sample collection. Faecal samples were transferred to permanent freezing facilities and stored at -18°C after returning to the research base. Samples were thawed and dried at 63°C for 18 h in an oven (Labotec, Johannesburg). Eighteen hours was initially estimated to be sufficient to remove the moisture content from faecal samples, on the basis of visual and tactile examination of samples. A subsequent trial involving the regular weighing of samples throughout the drying process showed that the weight of samples became stable after 12 h of drying. Samples were brought back to London every year to be analysed in batches.

Faecal progestagen analysis followed a method described and validated in wild black rhinoceros (Garnier et al., 1998a). Briefly, steroids were extracted by combining 0.1 g dry faeces with 0.2 g aluminium oxide, 0.6 ml methanol and 0.5 ml distilled water. Faecal progestagens were then measured with an enzyme immunoassay that used an anti-20α-dihydroprogesterone (20α-OHP) antibody which had been validated for urinary steroid analysis during oestrous cycles in African rhinoceroses (Hindle et al., 1992) and for faecal steroid analysis during pregnancy in wild black rhinoceroses (Garnier et al., 1998a). The enzyme immunoassays were performed in microtitre plates coated with an anti-IgG antibody, using a double antibody technique. The antibody was raised in rabbits immunized against 20α-OHP conjugated to BSA through carboxymethylxime (donated by M. J. Peddie, Department of Physics and Pharmacy, University of Southampton). The
enzyme label was biotin conjugated to 20α-OHP (donated by E. Möstl, Institut für Biochemie, Veterinärmedizinische Universität, Vienna).

Data manipulation and statistical analysis

Oestrous cycles. The initial observation of intergestation profiles showed differences in background concentrations of progesterone in each profile. For this reason, each intergestation period was divided into distinct periods, I1–I4. The definition of each period corresponded to the part of the profile that presented similar background concentrations, although the distinction between one period and the next was determined subjectively. Mean background concentrations for each period were calculated subsequently by removing progressively higher and lower values from the original data set until the coefficient of skewness was no longer significant and then by calculating the mean of the remaining concentrations (Brinkley, 1991). Differences between mean background concentrations were tested by one-way ANOVA.

A cyclic pattern was identified on the basis of endocrine criteria, when two consecutive values below mean background concentrations were followed by a sustained increase in faecal progestagen concentrations above mean background concentrations, before decreasing again to a nadir of at least two consecutive values below mean background concentrations. An increase of concentrations above mean background concentration was considered to be sustained if two of three consecutive values were above mean background concentration.

The duration of the cycle was defined as the time interval between the first day of successive periods of sustained increase in faecal progestagen concentrations as defined above. Cycle analysis applied only to parts of the hormonal profile that had no sampling interval > 6 days, as a longer sampling interval could result in the lack of identification of the period of low progesterone concentrations. This restriction is based on the results obtained (see below) and on previous reports that periods of reproductive activity in wild black rhinoceros females lasted 6–7 days (Hitchins and Anderson, 1983). As samples were not collected every day, the number of days between the end of the period of high concentrations and the subsequent period of low concentrations was divided equally between the two periods. The same principle applied between the end of a period of low concentrations and a subsequent period of high concentrations.

Subsequently, cycles were categorized into different types. Cycles that were the most represented and the total durations of which best fitted a normal distribution were found to have a total duration \( \approx 40 \) days and were categorized as type I cycles. Type II cycles had a total duration of >40 days and were further classified into type IIA cycles, with a period of low concentrations \( \leq 15 \) days, and type IIB cycles, with a period of low concentrations >15 days.
Comparisons between cycle durations were made using the Mann–Whitney U test.

Intervals between interactions with males were calculated by measuring the interval between the first days of periods of observed interactions, considering only intervals that were > 10 days, as periods of interactions with the same male could last as long as 12 days.

*Seasonality.* Seasonality was assessed by: (i) aligning the periods of ovarian cyclic activity and gestation with the time of the year; (ii) assessing the relative proportion of reproductive cycles of types I and II for each month; and (iii) assessing the monthly distribution of births. The beginning of periods of cyclic activity was determined using the results of faecal steroid analysis and behavioural observations for the six females studied in the SVC, and through observations of behavioural oestrus in the three females studied in the Imire game ranch. On the basis of the findings of the present study, cycles determined through behavioural observations at Imire were also categorized into two types: duration of type I cycles was \(\leq 40\) days and duration of type II cycles was > 40 days. The monthly assessment of type of reproductive cycle used dates related to the onset of the luteal phase for cycles determined by faecal steroid analysis and dates related to the first signs of oestrous behaviour for cycles determined through behavioural observations.

The proportions of births in the dry and rainy seasons were compared using a chi-squared test with Yates’ continuity correction.

**Results**

**Validation of the enzymeimmunoassays for progestagen measurement during oestrous cycles**

A cyclic pattern of variations of faecal 20α-progestagens was obtained in the captive female monitored for 9 months.
Table 2. Mean background concentrations of faecal 20α-progestagens from parturition up to concentrations of > 2000 ng g⁻¹, which is indicative of pregnancy in female wild black rhinoceroses.

<table>
<thead>
<tr>
<th>Female ID</th>
<th>Duration – days of profile (dates)</th>
<th>I1 Mean (± se) faecal 20α-progestagens (ng g⁻¹)</th>
<th>I2 Mean (± se) faecal 20α-progestagens (ng g⁻¹)</th>
<th>I3 Mean (± se) faecal 20α-progestagens (ng g⁻¹)</th>
<th>I4 Mean (± se) faecal 20α-progestagens (ng g⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bulawayo</td>
<td>1–112 (15/5/96–4/9/96)</td>
<td>987 ± 49</td>
<td>487 ± 26</td>
<td>698 ± 33*</td>
<td></td>
</tr>
<tr>
<td>Pukwani</td>
<td>1–164 (18/1/97–2/7/97)</td>
<td>915 ± 64</td>
<td>682 ± 59</td>
<td>259 ± 24</td>
<td>749 ± 50*</td>
</tr>
<tr>
<td>Netsai</td>
<td>1–159 (6/2/97–9/7/97)</td>
<td>1189 ± 82</td>
<td>458 ± 12</td>
<td>734 ± 24*</td>
<td></td>
</tr>
<tr>
<td>Jete</td>
<td>1–282 (2/10/96–10/7/97)</td>
<td>820 ± 31</td>
<td>470 ± 22</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sirica</td>
<td>1–213 (1/12/96–2/7/97)</td>
<td>720 ± 37</td>
<td>494 ± 14</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sara</td>
<td>1–35 (26/5/98–30/6/98)</td>
<td>1375 ± 117</td>
<td>37 ± 81</td>
<td>575 ± 37</td>
<td></td>
</tr>
</tbody>
</table>

Data from Garnier et al., 1998a.

Different periods (I1, I2, I3 and I4) were considered on each hormonal profile corresponding to differences in background observed on the profile. The definition of each period corresponded to the part of the profile that presented similar background concentrations, although the distinction between one period to the next was determined subjectively. Mean background concentrations for each period were calculated subsequently by removing progressively higher and lower values from the original data set until the coefficient of skewness was no longer significant and then by calculating the mean of the remaining concentrations.

*Periods of early gestation.
The hormone profile showed cyclic fluctuations of faecal progesterone metabolite concentrations. Periods of high concentrations of faecal 20α-progestagens, lasting an average of 15 ± 1.5 days (range 9–26 days, n = 11), were followed by periods of low concentrations, which lasted an average of 11.6 ± 2.1 days (range 5–29 days, n = 11). Reproductive activity lasted up to 4 days in this female and occurred either just before or during periods of low concentrations of faecal 20α-progestagens. Concentrations of faecal 20α-progestagens increased significantly between day 5 and day 12 after the first signs of reproductive activity were observed. The only time when behavioural observations of oestrus and hormonal variations were not related was represented by the absence of oestrous behaviour at about day 160, although a marked decline in faecal 20α-progestagens occurred.

**Individual hormone profiles**

Three wild females showed no interaction with an adult male during the first 4.0–4.5 months after parturition (Fig. 2a–c), whereas two females only resumed regular interactions with males 7–8 months after parturition (Fig. 2d,f). In the female whose calf was killed by lions at 3 weeks of age (Fig. 2e), the resumption of interactions with males occurred within 2 days after the calf died. In all females, the postpartum period was characterized by a higher mean background concentration of faecal 20α-progestagens (700–1400 ng g⁻¹) compared with subsequent intergestational periods during which it was about 500 ng g⁻¹ (Table 2). During this postpartum period of behavioural reproductive inactivity, faecal progesterone metabolite concentrations were fluctuating (as shown in Fig. 2a), but statistical analysis did not reveal any clear cyclic pattern.

Individual hormonal profiles in six wild females showed between two and eight cyclic variations of faecal 20α-progestagen concentrations in each female, occurring between 4 and 15 months after parturition (Fig. 2). The last cyclic variation detected on some profiles (Fig. 2a–c) occurred before the sustained increase in faecal 20α-progestagens to concentrations > 2000 ng g⁻¹ and simultaneously with a significant increase in mean background concentration (P < 0.01), indicating that these females were already in a phase of early pregnancy (Table 2). However, in two females, the sustained increase of faecal 20α-progestagens to concentrations > 2000 ng g⁻¹ was followed by an abrupt decline in concentrations between day 70 and day 100 after presumed conception (Fig. 2d,e). Such a marked and rapid decrease in concentrations, associated with the resumption of regular interactions with males and of cyclic variations in faecal 20α-progestagen concentrations, indicates that there is a very strong probability that these two females aborted.

Each female interacted with between one and three adult males, and with up to two sub-adults. One male always dominated the consortship for a female and consequently will be called the dominant male for that female. Interactions with dominant males represented between 38 and 100% of interactions with both adult and sub-adult males. In all cases, it was the dominant male that was observed to mate the female and to fight with other males. Most observations of consortship with the dominant male were coincidental with periods of low concentrations of faecal 20α-progestagens, whereas periods of consortship with other males, including sub-adults, could occur during periods of high concentrations. Significant increases in faecal 20α-progestagens were detected within 6 days after the observation of mating, although in one instance it was detected 1 day before the observation of mating activity. One female mated after presumed conception (Fig. 2e).

There were great individual differences in the number of male interactions recorded for each female and some females continued interacting with their previous male calves. Such consortship represented up to 25% of all direct observations recorded. As a result, associations of up to five animals were seen, including the breeding male, the cow–calf unit and her two previous calves.

**Durations and types of cycle**

The different types of cycle established from individual cycles (n = 27) obtained in the six females studied are presented (Table 3). Type I cycles were most frequent and were characterized by a total duration of 26.8 ± 1 days (mean ± se) and by periods of high and low concentrations of faecal 20α-progestagens of 18 ± 1.1 days and 9 ± 0.5 days, respectively. Type IIa cycles were detected during the first half of the hormonal profiles of two study animals and were preceded by the observation of mating activity (Fig. 2b,f). However, the fluctuations of 20α-progestagen concentrations that were observed during early pregnancy in three animals (Fig. 2a–c) had the same characteristics as type IIa cycles (total duration of ≥ 40 days and period of low 20α-progestagen concentrations < 15 days) and all were associated with a significantly longer period of high 20α-progestagen concentrations compared with type I cycles (P < 0.001).

Type IIb cycles presented a significantly longer period of low 20α-progestagen concentrations compared with type I cycles (P < 0.001). Such cycles were identified once in four of the six females studied. Of these cycles, two cycles were associated with the observation of a fight between males during the extended period of low 20α-progestagen concentrations; one cycle had a period of low 20α-progestagen concentrations surrounded by two periods of consortship with males; and one cycle was not associated with any interactions with males.

Type I cycles represented 75% (n = 21) of all cycles, whereas type IIa cycles represented at least 7% (n = 2) and type IIb at least 13% (n = 4) of all cycles (one type IIa cycle could not be classified definitively as type IIa or IIb).

**Periods of reproductive activity**

The periods of post partum, oestrous cyclicity and gestation recorded in the six multiparous females studied...
Table 3. Characteristics of the different types of oestrous cycle in six female wild black rhinoceroses (Diceros bicornis minor)

<table>
<thead>
<tr>
<th>Type of cycle</th>
<th>Type Ia</th>
<th>Type IIa&lt;sup&gt;b&lt;/sup&gt;</th>
<th>Type IIb&lt;sup&gt;c&lt;/sup&gt;</th>
</tr>
</thead>
<tbody>
<tr>
<td>(n = 21)</td>
<td>(n = 2)</td>
<td>(n = 4)</td>
<td></td>
</tr>
<tr>
<td>Total duration of oestrous cycle (days)</td>
<td>26.8 ± 1.0</td>
<td>46.7</td>
<td>53.0 ± 6.6</td>
</tr>
<tr>
<td></td>
<td>(19–36)</td>
<td>(45–48)</td>
<td>(43–72.5)</td>
</tr>
<tr>
<td>Duration of maintenance of high concentrations</td>
<td>18.0 ± 1.1</td>
<td>35.7</td>
<td>23.1 ± 4.6</td>
</tr>
<tr>
<td>of faecal progestagen (days)</td>
<td>(10–30)</td>
<td>(34–37)</td>
<td>(10–31)</td>
</tr>
<tr>
<td>Duration of maintenance of low concentrations</td>
<td>9.0 ± 0.5</td>
<td>11</td>
<td>30.3 ± 6.3</td>
</tr>
<tr>
<td>of faecal progestagen (days)</td>
<td>(6–13)</td>
<td>(8–14)</td>
<td>(19–45)</td>
</tr>
</tbody>
</table>

Values are mean ± SEM. Values in parentheses indicate range.

<sup>a</sup>Total duration of the cycle < 40 days.

<sup>b</sup>Total duration of the cycle ≥ 40 days and period of low faecal progestagen concentrations ≤ 15 days.

<sup>c</sup>Total duration of the cycle ≥ 40 days and period of low faecal progestagen concentrations > 15 days.

Fig. 3. Annual reproductive cycles in female wild black rhinoceroses (Diceros bicornis minor), including multiparous and nulliparous females. Asterisks indicate nulliparous females. The first six females in the list were studied in 1996–1999, whereas the last three females were studied in 1994–1997. Dotted lines (····) indicate periods of gestation or periods during which faecal 20α-progestagen concentrations increased to > 2000 ng g⁻¹. Dashed lines (---) indicate a postpartum period without detected oestrous behaviour in multiparous females or an inter-oestrous interval of > 60 days in nulliparous females. Complete lines indicate periods of oestrous cyclicity. Values in boxes indicate the duration (months) of each period.
above were aligned by months (Fig. 3). Periods of oestrous cyclicity started in May–June in four females and lasted between 4 and 7 months, except for the female whose calf died, for whom oestrous cyclicity lasted 1 month. In females whose calves stayed alive, all conceptions (n = 6) determined through faecal steroid analysis occurred in November–February, corresponding to the early rainy season and late spring–early summer, and half of these were detected in November. As a result, all females (except the two females that presumably aborted) gave birth in March–May, that is, during the late and after the rainy season.

In nulliparous females, periods of reproductive cyclicity lasted more than 1 year in two females (Amber, Mvu) and more than 2 years in one female (DJ) (Fig. 3). An extended interoestrous interval of about 2.5 months was observed in all females in October–December 1994. Other extended interoestrous intervals of 3.0–3.5 months were detected every year in April–June and were extremely synchronized among females (Amber, Mvu, DJ), beginning between 19 and 24 March and ending between 25 June and 12 July. These females conceived between the end of November and January. Two of these females did not interact with any male until about 7 months after parturition, after which time their calves were removed to be hand-reared. These females conceived 9 days and 2 months, respectively, after removal of their calves.

Distribution of cycle durations

The monthly distribution of cycle durations in multi-

parous females (Fig. 4a) shows that type I and type II cycles were recorded between May and February.

In nulliparous females (Fig. 4b), type I cycles occurred every month from June to February, but predominated between November and January. Type II cycles prevailed in February–March and in October.

Distribution of births

The monthly distribution of births (n = 21) recorded during the study (Fig. 5) shows that the proportion of births recorded in December–May (71%) was significantly higher than in June–November (chi-squared = 3.85, P = 0.03). In fact, more births (62%) occurred between February and May, coinciding with the late rainy season, reflecting peak conception between November and February, that is, during the early rainy season. However, four births were also recorded in mid-winter in July and August, two of which were observed in nulliparous females.

Discussion

The individual monitoring of six wild females in Zimbabwe showed that they did not conceive before 8 months after parturition when their calves survived and that they had very limited or no contact with males for 4–7 months after parturition. This is the first report of a postpartum period of reproductive inactivity in black rhinoceroses, although postpartum anoestrus has been reported in captive white and Indian rhinoceroses (of 3–4 months duration for Indian rhinoceroses; Schwarzenberger et al., 1999; Rietschel, 2000).

The hormone profiles obtained in females that could be monitored frequently during this period of reproductive inactivity showed fluctuating faecal progesterone metabolite concentrations and a high background concentration. Although there is no evidence that these progesterone metabolites were of ovarian origin, this finding may indicate that ovaries start to develop follicles shortly after parturition, as results from previous studies showed that follicular maturation resumed within 2–4 weeks after birth in some captive
black rhinoceros females (Hindle, 1991; Berkeley et al., 1997). However, the death or removal of a black rhinoceros calf shortened the period of reduced reproductive activity, indicating that expression of reproductive behaviour is inhibited by the suckling stimulus. Therefore, the post-partum period in the black rhinoceros is similar to that of many mammals, in which ovulation and oestrus are delayed by the suckling stimulus (Cupps, 1991; Youngquist, 1997). The mechanisms by which the suckling stimulus inhibits oestrous behaviour could not be determined during the present study. However, the higher mean background of faecal progestagen concentrations may reflect the luteinization of follicles that develop during this phase, as high background concentrations of faecal progestagens have been reported to be indicative of persistent luteal activity in captive white rhinoceroses (Schwarzenberger et al., 1998).

Periods of oestrous cyclicity lasted between 4 and 7 months and were characterized by a great variability in cycle duration. Three-quarters of all cycles were classified as type I and presented a mean total duration of 26.8 ± 1 days, which is in agreement with other reports of cycle durations in this species (Lindemann, 1982; Hitchins and Anderson, 1983; Godfrey et al., 1991; Hindle et al., 1992; Schwarzenberger et al., 1993; Berkeley et al., 1997; Radcliffe et al., 2001). Most observations of reproductive activity (for example, a fight between males, periods of prolonged interactions with the dominant male and mating) occurred during periods of low faecal progestosterone metabolite concentration and were followed by a significant increase in faecal progesterone metabolite concentration. Therefore, in type I cycles, periods of low faecal progestosterone metabolite concentration are associated with the follicular phase of an ovarian cycle, whereas periods of high concentrations are associated with the luteal phase.

A lag of variable duration (5–12 days) was observed between the observation of reproductive activity and a significant increase in concentrations of faecal progestosterone metabolites. Part of this lag may be attributable to the delay coinciding with the entero-hepatic circulation and faecal excretion of steroids, estimated to be 2–3 days in rhinocerotids (Hindle and Hodges, 1990; Heistermann et al., 1998), as well as to the delay of 2–3 days observed between onset of oestrus and ovulation in black rhinoceroses (Radcliffe et al., 2001). In addition, another interval probably occurs between ovulation and the subsequent increase in faecal progestagens, as has been described in white rhinoceroses (Radcliffe et al., 1997; Schwarzenberger et al., 1999).

One of the unexpected findings of the present study was the natural occurrence of extended cycles classified as type II, which were approximately twice as long as type I cycles. The observation of these cycles was unlikely to have been caused by a failure to detect a change in concentrations as the sampling frequency was very high during the period involved (every 2–3 days). Type II cycles were characterized by either a prolonged luteal phase or by an extended follicular phase.

A plausible explanation for these prolonged luteal phases is embryo loss. This pathology has been identified in three captive white rhinoceros females, and the corresponding luteal phases, identified by faecal pregnancy analysis, lasted about twice the average cycle duration (Radcliffe et al., 1997; Patton et al., 1999). These reports, combined with the fact that a history of mating followed by anoestrus without pregnancy in mares is indicative of early embryo loss (Youngquist, 1997), indicate that such pathology may be associated with type Ila cycles. The fact that extended luteal phases were detected during the early gestation period in the present study reinforces this hypothesis. In contrast, type Ila cycles are unlikely to be associated with persistent luteal activity, a condition that occurs frequently in captive white rhinoceroses but which results in different hormone patterns (Schwarzenberger et al., 1998).

Type Iib cycles were characterized by an extended follicular phase and were anovulatory. The fact that most of these cycles occurred in October, which is the hottest time of the year in Zimbabwe and the time during which nulliparous females exhibit an extended interoestrous interval, indicates that these cycles may be caused by heat stress. This hypothesis is corroborated by the observation that free-ranging white rhinoceros did not exhibit oestrous behaviour when dry conditions prevailed (Owen-Smith, 1988).

Heat stress is known to affect follicular growth, to decrease the duration of oestrous behaviour and to disrupt pregnancy in domestic cattle, in which conception rates decrease markedly during the warmest months of the year (Youngquist, 1997).

Periods of interactions with the dominant male lasted up to 12 days, which is about twice as long as the duration of consortship reported by Hitchins and Anderson (1983). This difference may be attributed to the high frequency of observations that were undertaken during the present study. Fighting between the dominant male and another male occurred during such periods of consortship, between 11 and 18 days before a significant increase in faecal progestagen concentrations. This finding indicates that the dominant male tries to prevent other males, including the receptive female’s previous calves after they have become sub-adult, from accessing her (sub-adults continue interacting with their mother before reaching sexual maturity; Garnier et al., 2001). However, it is not known whether the dominant male exhibits this behaviour of mate guarding on his home range, as has been reported in white rhinoceroses in the wild. Moreover, the potential for male and female to overlap home ranges and for dominant male to mate-guarding does not seem to prevent other males from accessing the receptive female within her home range. The observations that two adult females with overlapping home ranges also had the same dominant male and also had overlapping home ranges with that male indicate that a dominant male may become territorial over their common home range when a female becomes receptive (J. N. Garnier, unpublished).

During the gestation period, reproductive activity...
occurred after presumed conception in a wild female, and this activity may contribute towards an explanation of gestation durations of under 14 months noted in black rhinoceroses, which usually have a duration of gestation of approximately 15 months (Hall-Martin and Penzhorn, 1977; Lindemann, 1982; Hitchins and Anderson, 1983; Garnier et al., 1998a; Garnier, 2001). This finding emphasizes the difficulty of diagnosing gestation from behavioural observations only. Such post-conception activity closely resembles that of domestic mares and is probably associated with a temporary surge in oestrogens, identified in a captive black rhinoceros female by Berkeley et al. (1997).

In two females, the marked and rapid decrease in concentrations 2.5–3.5 months after presumed conception associated with the resumption of regular interactions with males is indicative of abortion. This hypothesis is reinforced by the absence of delivery of a calf within 16 months of presumed conception in these two females and by the observation that profiles of faecal progestagen excretion obtained with the same assay as used in pregnant wild black rhinoceros females did not show a decrease in concentrations after the second month of gestation (Garnier et al., 1998a). Abortion has been reported in some captive black rhinoceroses and Sumatran females, but mostly in isolated cases (Hodges and Green, 1989; Schwarzenberger et al., 1993, 1996b; Berkeley et al., 1997; Roth et al., 2001). Because the onset of placental steroid production occurs between months 2 and 4 of gestation (Schwarzenberger et al., 1993; Garnier et al., 1998a), this period of pregnancy may represent a vulnerable phase in black rhinoceros females.

The natural incidence of abortion in wild black rhinoceros populations may be higher than previously thought, especially since the two females that aborted during the study belonged to a population that presented good average reproductive performances. This finding emphasizes the importance of using non-invasive reproductive assessment for monitoring individual breeding output in free-ranging endangered wildlife species. In particular, females that have entered post-reproductive life need to be detected. Shenkel and Shenkel-Hulliger (1969) suggested that the reproductive life of wild black rhinoceroses lasts until 30–35 years of age, which is corroborated by observations in the present study that three females estimated to be about 35 years old were still producing calves, although one of these aborted.

The analysis of the timing of reproductive cycles and gestation periods in black rhinoceros females in Zimbabwe indicated that more successful fertilizations took place between November and February, during the summer and rainy season. Despite the limited number of births recorded, the results of the present study support findings from other studies on black and white rhinoceros populations in southern Africa, as well on Indian rhinoceros populations, which identified a calving peak during the wettest months of the year (Joubert and Eloff, 1971; Hall-Martin and Penzhorn, 1977; Laurie 1982; Hitchins and Anderson, 1983; Hall-Martin, 1986; Owen-Smith, 1988). Such timing of births ensures that temperature and food resources are most suitable for the optimum growth and survival of the young (the rainy season is considered an important factor of influence on reproductive activity in tropical zones; Flowerdew, 1987). Peak conception occurs during the rainy season in a variety of African mammals, including Perissodactyla such as plains zebra and Grevy's zebra (Owen-Smith, 1988; Estes, 1992).

Three nulliparous female rhinoceroses had an interoestrous interval of 3 months in April–June, the onset of which was highly synchronized. The beginning of this anoestrous period coincided with the autumal equinox in the southern hemisphere (21 March) and its termination corresponded to the winter solstice (21 June). The precise timing of these patterns from one year to another, combined with their synchronization among animals, indicates the occurrence of a seasonal period of reduced reproductive activity during the period of decreasing daylengths in nulliparous females. It could not be determined whether multiparous females also exhibited this seasonal anoestrus, as they were either pregnant or in a phase of postpartum anoestrus. Nevertheless photoperiodicity probably contributes to the regulation of the breeding season in black rhinoceros females. This hypothesis is corroborated by results obtained from captive black rhinoceroses in the northern hemisphere, where all nulliparous females studied presented extended interoestrous intervals of 2–3 months between November and February, whereas multiparous females presented a significantly higher proportion of conceptions during the summer months, except in June (Garnier, 2001). In addition, Radcliffe et al. (2001) reported the formation of large anovulatory follicles during the winter months in a captive black rhinoceros female monitored by ultrasound examination.

Therefore, the seasonal pattern of reproductive activity identified in black rhinoceros females in Zimbabwe may be similar to that occurring in equids, which present a winter anoestrus and a natural breeding season during the spring and summer months (Youngquist, 1997). However, interactions among various environmental factors, such as rainfall, periodicity, temperature and parity, are probably complex. For example, the breeding seasons of pubertal zebu cattle heifers are determined by photoperiodic constraints, whereas by the fifth calving, rainfall has become the factor of overriding influence (Cupps, 1991).

In conclusion, although the number of animals in the present study was limited, the results indicate that black rhinoceros females in the wild exhibit a lactational anoestrus of at least 4 months followed by a period of oestrous cyclicity characterized by a great variability in cycle duration. Photoperiodicity, among other environmental factors such as rainfall and temperature, probably contributes to the regulation of reproductive patterns. This study emphasizes the importance of monitoring naturally occurring patterns of reproductive activity in situ to help in providing a template for breeding programmes of this endangered species.
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References


Berkeley EV, Kirkpatrick JF, Schaffer NE, Bryant WM and Threlfall WR (1997) Serum and faecal steroid analysis of ovulation, pregnancy and parturition in the black rhinoceros (Diceros bicornis) Zoo Biology 16 121–132


Hall-Martin AJ (1986) Recruitment in a small black rhinoceros population Pachyderm 6 7–8

Hall-Martin AJ and Penzhorn BL (1977) Behaviour and recruitment of translocated black rhinoceros Koedoe 20 147–162


Joubert E and Elloff FC (1971) Notes on the ecology and behaviour of the black rhinoceros (Diceros bicornis) in South West Africa Madogga 5 3–5


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