

Steroidogenic responses of pig corpora lutea to insulin-like growth factor I (IGF-I) throughout the oestrous cycle

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This study was designed to investigate the roles of insulin-like growth factor I (IGF-I), IGF-type I receptor (IGF-IR) and IGF-binding proteins (IGFBPs) in regulating progesterone secretion by pig corpora lutea during the oestrous cycle, and the signal transduction pathways involved in mediating the steroidogenic actions of IGF-I. Corpora lutea were collected on days 4, 7, 10, 13 and 15 or 16 of the oestrous cycle, enzyme dissociated and the luteal cells were cultured for 24 h in Medium 199 with IGF-I (0–100 ng ml⁻¹), long R₃-IGF-I (0–100 ng ml⁻¹), anti-IGF-I (Sm 1.2B; 0–10 µg ml⁻¹), anti-IGF-IR (αIR3; 0–2 µg ml⁻¹), or IGF-I signal transduction pathway inhibitors (phosphatidylinositol (PI)-3-kinase: 100 nmol Wortmannin l⁻¹ and 10 µmol LY 294002 l⁻¹; MAP kinase: 50 µmol PD 98059 l⁻¹) to investigate their effects on IGF-I (100 ng ml⁻¹) stimulated progesterone secretion. Pig luteal

cells displayed dose-dependent responses to IGF-I and long R₃-IGF-I on days 4 and 7 of the oestrous cycle, but not on days 10–16. There was no difference in the ED₅₀ or V_{max} (maximal response) values between IGF-I and long R₃-IGF-I. Neither anti-IGF-I nor anti-IGF-IR had significant effects on progesterone secretion, at any dose or day. Wortmannin and LY 294002 blocked IGF-I stimulated progesterone secretion, but PD 98059 was without effect. Finally, IGF-I (6 µg) infused into the ovary on day 7 *in vivo* significantly increased progesterone secretion within 45 min of infusion. The conclusions of this study are: (1) IGF-I has steroidogenic actions only on 'young' (days 4–7) pig corpora lutea; (2) endogenous IGF-I and IGFBP are insufficient to modulate progesterone secretion *in vitro*; and (3) the steroidogenic actions of IGF-I are mediated via PI-3-kinase.

Introduction

In most species, corpora lutea can continue to develop for approximately 5 days without pituitary gonadotrophic support before premature luteolysis occurs (Niswender *et al.*, 2000). The pig, however, is unique in that its corpora lutea can continue to develop for 12–14 days without the support of pituitary gonadotrophins (Anderson and Melampy, 1968). Although the exact mechanisms by which this occurs are not well understood, it is hypothesized that a local autocrine or paracrine control pathway involving insulin-like growth factor I (IGF-I) plays an important role in the regulation of corpora lutea development and function during this period (Gadsby *et al.*, 1996; Nicholson *et al.*, 1999; Ge *et al.*, 2000a).

Although the importance of IGF-I to ovarian follicular development has been well established (Hammond *et al.*, 1993; Monget and Bondy, 2000; Guidice, 2001), the role of IGF-I in corpora lutea is less clearly defined. IGF-I mRNA expression has been shown in the corpora lutea of several species including rats (Parmer *et al.*, 1991), humans (Hernandez *et al.*, 1992), sheep (Perks

et al., 1995; Juengel *et al.*, 1997), cows (Einspanier *et al.*, 1990; Perks *et al.*, 1999) and pigs (Hammond *et al.*, 1993; Gadsby *et al.*, 1996). IGF-I has been identified in bovine corpora lutea throughout the oestrous cycle by immunocytochemistry (Amselgruber *et al.*, 1994) and is secreted by ovine luteal cells in culture (Wathes *et al.*, 1995). Studies in pig corpora lutea have shown that the steady-state expression of IGF-I mRNA is greater on days 4–10 compared with days 12–16 of the oestrous cycle, and that IGF-I mRNA is expressed to a greater extent in small compared with large luteal cells (Gadsby *et al.*, 1996). Recently, the expression of IGF-I in early luteal phase pig corpora lutea has been confirmed by demonstrating that luteal concentrations of IGF-I are highest on day 4, compared with the later stages (days 7–16; Z. Ge, W. E. Nicholson, V. Hedgpeth and J. E. Gadsby, unpublished).

The actions of IGF-I are dependent upon a functional type I IGF receptor (IGF-IR; Valentinis and Baserga, 2001). IGF-IR has been detected in the corpora lutea of rats (Parmer *et al.*, 1991; Talavera and Menon, 1991), humans (Hernandez *et al.*, 1992), sheep (Perks *et al.*, 1995) and cows (Sauerwein *et al.*, 1992; Perks *et al.*, 1999). Studies of pig corpora lutea have demonstrated that IGF-IR mRNA and protein are expressed and that IGF-IR

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(protein) concentrations are highest on days 4–10, compared with days 13–16 of the oestrous cycle (Ge *et al.*, 2000a). In addition, IGF-IR mRNA and protein are localized predominantly to large (compared with small) luteal cells (Ge *et al.*, 2000a). IGF-I stimulates luteal progesterone secretion *in vitro* in several species (rats, Parmer *et al.*, 1991; rabbits, Constantino *et al.*, 1991; humans, Apa *et al.*, 1996; sheep, Khan-Dawood *et al.*, 1994; and cows, McArdle and Holtorf, 1989; Sauerwein *et al.*, 1992; Brown and Braden, 2001) including pigs (Huang *et al.*, 1992; Yuan and Lucy, 1996). Furthermore, IGF-I has been shown to stimulate luteal progesterone secretion *in vivo* in cows (Sauerwein *et al.*, 1992).

The IGF-binding proteins (IGFBPs) are thought to play an important role in modulating the biological actions of the IGFs (Baxter, 2000; Grimberg and Cohen, 2000). IGFBP-2 and -3 are expressed in the corpora lutea of several species (rats, Erickson *et al.*, 1993; primates, Fraser *et al.*, 1998; humans, Fraser *et al.*, 2000; and cows, Brown and Braden, 2001). Expression of mRNA has been demonstrated for IGFBPs-2, -3, -4 and -5 in pig corpora lutea (Gadsby *et al.*, 1996), and IGFBP-3 mRNA is expressed predominantly in small luteal cells (Gadsby *et al.*, 1996; Ge *et al.*, 2000b). In addition, the IGFBP-3 protein content of pig corpora lutea is highest on days 4–7, compared with days 10–16 (Z. Ge, W. E. Nicholson, V. Hedgpeth and J. E. Gadsby, unpublished). The role that IGFBPs play in the corpora lutea is not well understood, although as detailed in the discussion, the overwhelming view of the published literature indicates that IGFBPs (in particular IGFBP-3) play an inhibitory role in regulating the action of IGF-I.

In summary, luteal expression of IGF-I, IGF-IR and IGFBP in pigs is greatest during the early luteal phase (days 4–10), supporting the hypothesis that IGF-I plays a critical role in corpora lutea development and function (that is, progesterone secretion) during the period (12–14 days) of autonomy from pituitary gonadotrophins. This hypothesis is explored further in the present study by investigating the steroidogenic responses of pig corpora lutea to IGF-I, and the functional roles of IGF-IR and IGFBPs throughout the oestrous cycle, as outlined in the following objectives: (1) to examine the function of the luteal IGF-IR at different stages of the oestrous cycle; (2) to explore the role of endogenous IGF-I; (3) to examine the role of endogenous IGFBPs in modulating luteal IGF-I responses; and (4) to explore the signal transduction pathway mediating the steroidogenic actions of IGF-I.

Materials and Methods

Materials

Medium 199 (M199 with Hank's salts and L-glutamine) was purchased from Gibco-Invitrogen Corp. (Grand

Island, NY). Hank's Balanced Salt Solution (HBSS; without calcium and magnesium), Hepes, sodium bicarbonate, hydrocortisone, DNase (type I from bovine pancreas), hyaluronidase (type II from ovine testes), BSA (fraction V), human low density lipoprotein (LDL), penicillin–streptomycin solution, dimethyl sulphoxide (DMSO), Wortmannin, LY 294002 and PD 98059 were obtained from Sigma-Aldrich Chemical Company (St Louis, MO). Gentamycin sulphate (biotech research grade) and sodium citrate were obtained from FisherBiotech (Fair Lawn, NJ). Collagenase (type IV) was purchased from Worthington Biochemical Corp. (Lakewood, NJ). The Coat-a-Count[®] progesterone radioimmunoassay kits were purchased from Diagnostic Products Corp. (Los Angeles, CA). Long R₃-IGF-I was obtained from Diagnostic Systems Laboratories (Webster, TX) and IGF-I was purchased from R & D Systems (Minneapolis, MN). Anti-human IGF-I (Sm 1.2B clone) was obtained from Upstate Biotechnology (Lake Placid, NY) and anti-IGF-IR (α IR3 clone) was purchased from Oncogene Research Products (Cambridge, MA).

Lipoprotein preparation

The LDL fraction from pig plasma was obtained by differential ultracentrifugation, using blood obtained from barrows as described by Richards *et al.* (1994).

Animals and collection of ovaries

The experimental protocol involving animal care and use was performed with the approval of the North Carolina State University (NCSU) Institutional Animal Care and Use Committee.

Naturally cycling pubertal gilts (*Sus scrofa*; Newsham \times Hampshire \times Duroc hybrid; from the NCSU Swine Education and Research Facility) were checked once per day for oestrous behaviour with a mature boar. The first day on which oestrus was observed was considered to be day 0 of the oestrous cycle. On days 4, 7, 10, 13 and 15 or 16 ($n = 5$ animal per day for all stages except days 15 and 16, for which $n = 4$) of the oestrous cycle, animals were ovariectomized (Gadsby *et al.*, 1990). Ovaries or corpora lutea were obtained from animals on day 6 or 7 of the oestrous cycle for the small or large luteal cell cultures and signal-transduction pathway studies.

Dissociation of corpora lutea and cell separation

Ovaries collected at surgery were immediately placed in sterile Hank's Medium (HBSS containing 20 mmol Hepes l⁻¹, 4 mmol sodium bicarbonate l⁻¹, 100 000 iu penicillin l⁻¹, 100 mg streptomycin l⁻¹, 50 mg gentamicin l⁻¹, and 2 g BSA l⁻¹) and transported to the laboratory on ice. All subsequent procedures, except where noted otherwise, were performed on ice. Corpora

lutea were dissected from the ovaries, weighed and then diced into small pieces with scalpel blades. The dissected tissue was placed in an Erlenmeyer flask with 40 ml of Hank's medium and incubated for 30 min at 37°C. Enzymatic dissociation of corpora lutea was then performed as described by Gadsby *et al.* (1990). The total number of large and small luteal cells was counted using a haemocytometer and the viabilities of small and large cells were monitored by Trypan blue exclusion.

Small and large luteal cells were separated using centrifugal elutriation (Gadsby *et al.*, 1990; Gadsby and Earnest, 1994; Ge *et al.*, 2000a), and were cultured separately.

Culture of luteal cells

The luteal cell culture procedure was performed as described by Richards *et al.* (1994). Briefly, the mixed, small or large luteal cell preparations were diluted in M199 containing 22 mmol Hepes l⁻¹, 4 mmol sodium bicarbonate l⁻¹, 100 000 iu penicillin l⁻¹, 100 mg streptomycin l⁻¹, 50 mg gentamicin l⁻¹, 1 g BSA l⁻¹, 110 nmol hydrocortisone l⁻¹ and 50 mg (cholesterol) pig LDL l⁻¹ (M199). Costar 24-well tissue-culture treated polystyrene plates (Corning Inc., Corning, NY) were seeded with 30 000 large (large cell preparations), 150 000 small (small cell preparations) or 30 000 large plus about 100 000–250 000 small luteal cells (mixed cell preparations; on the basis of a fixed large cell count and a variable number of small cells depending on stage of the oestrous cycle). Various concentrations of the following treatments were added to each well (in 1 ml): IGF-I (0, 0.1, 1, 10, 100 ng ml⁻¹; *n* = 4 per dose per animal), long R₃-IGF-I (0, 0.1, 1, 10, 100 ng ml⁻¹; *n* = 4), anti-IGF-I (Sm 1.2B; 0, 0.625, 1.25, 2.5, 5, 10 µg ml⁻¹; *n* = 4) and anti-IGF-IR (αIR3; 0, 0.1, 0.2, 0.5, 1, 2 µg ml⁻¹; *n* = 4). The doses selected for anti-IGF-IR were as suggested by Rohlik *et al.* (1987). In addition, a preliminary experiment showed that anti-IGF-IR (using the doses given above) dose-dependently inhibited the stimulatory effects of 10 ng IGF-I ml⁻¹, confirming the efficacy of this antibody with pig luteal cells. A range of concentrations was used for anti-IGF-I, encompassing the dose (equivalent to 3.6 µg ml⁻¹) found to be effective at blocking the action of IGF-I on pig granulosa cells (Mondschein *et al.*, 1989). Each treatment was diluted in the M199 culture medium described above. Plates were centrifuged at 250 g for 5 min at 4°C to facilitate attachment of cells to the bottom of the well. The plates were then incubated in a humidified incubator at 37°C with 5% CO₂ in air for 20–24 h. After culture, 24-well plates were again gently centrifuged and the spent media collected for progesterone determination by radioimmunoassay. Cells were removed from the culture wells by trypsinization for monitoring the number and viability of cells in

some experiments. This study confirmed the findings of Richards *et al.* (1994) that there are no significant changes in the number or viability of luteal cells over the course of the 20–24 h culture period (data not shown).

Inhibitor studies

The signal transduction pathways mediating the steroidogenic actions of IGF-I were explored using the luteal cell culture system described above. These studies used 50 000 (large cell count) mixed luteal cells per well and human LDLs. Preliminary studies found that human LDL (same dose as for pig LDL; 50 mg cholesterol l⁻¹) was a suitable substitute (for pig LDL) cholesterol source in these cultures (data not shown). These studies utilized the phosphatidylinositol (PI)-3-kinase inhibitors, Wortmannin and LY 294002, and the MAP-kinase inhibitor, PD 98059, at the following doses: 100 nmol Wortmannin l⁻¹; 10 µmol LY 294002 l⁻¹; 50 µmol PD 98059 l⁻¹ (Westfall *et al.*, 2000; Chen *et al.*, 2001). Inhibitors were dissolved in DMSO to generate stock solutions and added at the beginning of culture in ≤0.1% (v/v) culture medium. In preliminary experiments 0.1% DMSO had no effect on control or IGF-I-treated cultures.

Ovarian infusion studies, in vivo

Animals were anaesthetized and maintained at a surgical plane of anaesthesia (Gadsby *et al.*, 1990) on day 7 of the oestrous cycle. The ovaries were exposed via a mid-line ventral laparotomy and a 23-gauge butterfly catheter (3 or 4 inch needle, 6 inch catheter; Abbott Laboratories, Chicago, IL), for blood collection, was inserted into the vasculature located in the ovarian hilus (Oxenreider *et al.*, 1965); a second butterfly catheter for IGF-I or vehicle infusion was inserted adjacent to the first catheter. Both catheters were secured in place with super-glue (Quick Gel; Duro, Hartford, CT) and Dermabond (Ethicon/Johnson and Johnson, Somerville, NJ), and flushed with 3.5% sodium citrate. Blood samples were collected at approximately 5–10 min intervals for approximately 30 min before infusion. At time 0, IGF-I (6 µg per ovary; *n* = 3 animals) or vehicle (*n* = 2; sodium citrate) was infused into the ovary in 0.25 ml vehicle over 1–2 min. Blood sampling from the ovarian catheter was continued at 5–15 min intervals for 90–120 min. Blood samples were also collected at regular intervals (as described for ovarian samples) from the ear vein, before and after IGF-I or vehicle infusion. Blood samples were allowed to clot at room temperature, stored overnight at 4°C and then centrifuged (600 g for 20 min) to collect serum. Serum samples were stored at –20°C before assay for progesterone using radioimmunoassay.

Table 1. Dose–response characteristics of steroidogenic responses of pig luteal cells to insulin-like growth factor I (IGF-I) and long R_3 -IGF-I

Day of cycle	IGF-I		Long R_3 -IGF-I	
	ED ₅₀ (ng ml ⁻¹)	V _{max} (% of control)	ED ₅₀ (ng ml ⁻¹)	V _{max} (% of control)
4	6.6 ± 1.6 (5)	135 ± 2.8 (5)	7.2 ± 2.1 (4)	132 ± 7.5 (4)
7	4.1 ± 2.0 (5)	138 ± 4.4 (5)	3.6 ± 2.8 (3)	140 ± 2.8 (3)

Data are expressed as mean ± SEM. Numbers in parenthesis = *n*.

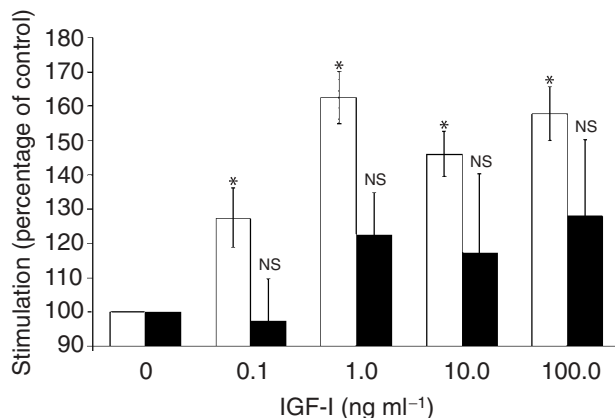


Fig. 1. Effects of insulin-like growth factor I (IGF-I) on progesterone secretion by large (□) and small (■) pig luteal cells. Data are presented as the stimulation of progesterone accumulation into the culture medium, expressed as a percentage, relative to controls (that is, no IGF-I = 100%). Data are presented as mean ± SEM for three separate experiments (luteal cells from three individual pigs at day 7). *Significant increase above controls ($P < 0.05$; ANOVA); NS, not significant.

Progesterone radioimmunoassay

Media were collected from luteal cell cultures, centrifuged (600 **g** for 20 min at 4°C) and immediately stored at -20°C. Progesterone concentrations (serum and media) were measured using the Coat-a-Count[®] radioimmunoassay kits as described and validated (serum, Gadsby *et al.*, 1991; media, Richards *et al.*, 1994).

Statistical analysis

Data were normalized within each culture (animal) for controls, expressed as a percentage of the control value (= 100%) and averaged across replicate for each treatment. Data were then averaged for day of oestrous cycle (for each treatment) and examined for treatment effects across days of the oestrous cycle using ANOVA and Fisher's *post hoc* test (Statview, Abacus Concepts, CA), with $P < 0.05$ as the significance level.

Results

In vitro studies

Responses of isolated large and small luteal cells to IGF-I. When large and small luteal cells (day 7), isolated and separated by elutriation, were cultured separately with 0–100 ng IGF-I ml⁻¹, only large cell preparations demonstrated a significant ($P < 0.05$) dose-dependent increase in progesterone secretion (Fig. 1), with a maximum stimulation of about 160% at 1 ng ml⁻¹. There was no significant stimulation observed with small luteal cells, although numerically they displayed a slight but variable response. All subsequent studies used mixed luteal cell preparations and a fixed number of large luteal cells (that is, 30 000 cells per well for most studies), as large luteal cells appeared to be the primary target cell for the steroidogenic actions of IGF-I.

IGF-I and long R_3 -IGF-I dose–response. Dose-dependent increases in progesterone secretion in response to IGF-I and long R_3 -IGF-I were observed only on days 4 and 7 (Fig. 2). From the dose–response curves constructed from the data for days 4 and 7, estimates of ED₅₀ value (that is, dose giving 50% of maximal response) and V_{max} (maximal response) were obtained (Table 1). These data indicate that steroidogenic responses of mixed luteal cells on days 4 and 7 to IGF-I and long R_3 -IGF-I were similar in terms of ED₅₀ (3–7 ng ml⁻¹) and V_{max} (132–140%) values.

Effects of IGF-I and long R_3 -IGF-I on different days of the oestrous cycle. The effects of IGF-I and long R_3 -IGF-I on progesterone secretion by pig luteal cells on days 4 and 7 are shown (Fig. 2a,b). IGF-I elicited significant ($P < 0.05$) dose-dependent increases in progesterone secretion at 1, 10 and 100 ng ml⁻¹ (in five out of five animals), on both days 4 and 7, giving maximal responses of 135% and 142% (above control; at 100 ng ml⁻¹), respectively. Long R_3 -IGF-I was stimulatory (significantly, in four out of five animals on day 4, and three out of five animals on day 7; $P < 0.05$) to steroidogenesis only at 10 and 100 ng ml⁻¹, although the maximal responses (128%, day 4; 140%, day 7) were not significantly different from that seen with IGF-I. During days 10–16, no significant stimulatory steroidogenic responses to 0.1–100 ng ml⁻¹ of either IGF-I or long R_3 -IGF-I were observed on days 10, 13, 15 or 16 (data not shown).

Effects of anti-IGF-I and anti-IGF-receptor sera on different days of the oestrous cycle. The effects of monoclonal antibodies to IGF-I (Sm 1.2B) and to IGF-IR (α IR3) were examined to investigate the roles of endogenous IGF-I in maintaining basal progesterone secretion by pig luteal cells in culture at different stages of the oestrous cycle. Neither of these antibodies had consistent or significant effects on progesterone secretion

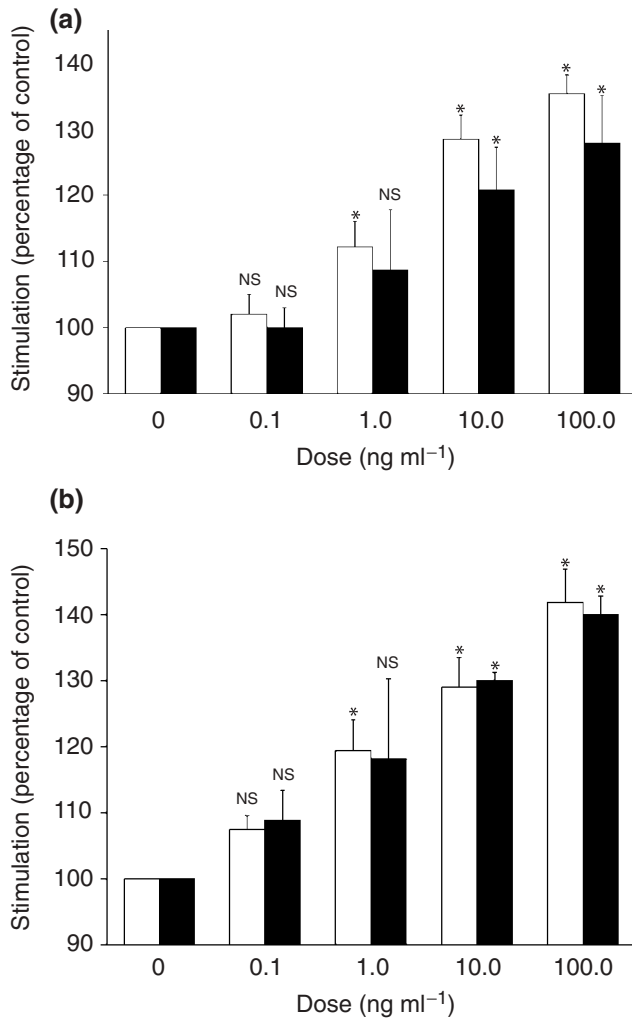


Fig. 2. Effects of insulin-like growth factor I (IGF-I; □) and long R₃-IGF-I (■) on progesterone secretion by mixed pig luteal cells obtained on (a) day 4 and (b) day 7 of the oestrous cycle. Data are presented as the stimulation of progesterone accumulation into the culture medium, expressed as a percentage, relative to controls (that is, no IGF-I = 100%). Data are presented as mean \pm SEM for five animals (days 4 and 7; IGF-I), and for four (day 4) and three (day 7) animals, respectively (long R₃-IGF-I). *Significant increase above controls ($P < 0.05$; ANOVA); NS, not significant.

at any dose or stage of the oestrous cycle (data not shown).

Effects of PI-3-kinase and MAP-kinase inhibitors on IGF-I stimulated steroidogenesis. The effects of PI-3-kinase and MAP-kinase inhibitors on progesterone secretion are shown (Fig. 3). These data indicate that Wortmannin ($111 \pm 1.5\%$) and LY 294002 ($116 \pm 1.3\%$) both significantly ($P < 0.05$ versus IGF-I alone) inhibit the stimulatory actions of IGF-I ($130 \pm 4.1\%$) on pig luteal cell steroidogenesis. However, PD 98059 ($127 \pm 2.7\%$) was without effect and was not able to significantly

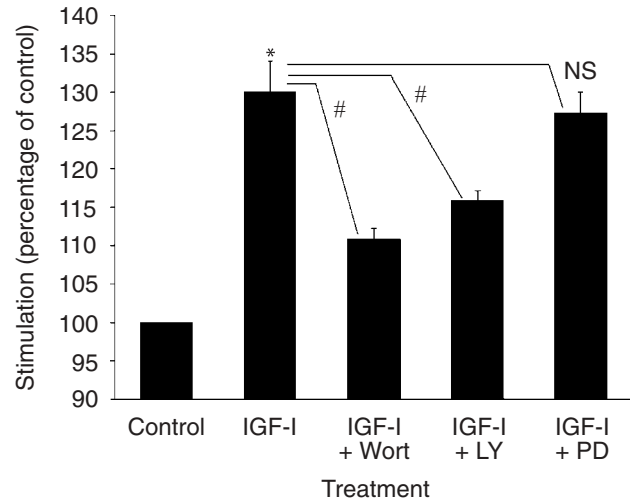


Fig. 3. Effects of inhibitors (100 nmol Wortmannin l^{-1} (Wort); $10 \mu\text{mol}$ LY 294002 l^{-1} (LY); $50 \mu\text{mol}$ PD 98059 l^{-1} (PD)) on insulin-like growth factor I (IGF-I; 100 ng ml^{-1}) stimulated progesterone secretion by mixed pig luteal cells obtained on day 6 or day 7 of the oestrous cycle. Data are presented as the stimulation of progesterone accumulation into the culture medium, expressed as a percentage, relative to their appropriate controls (that is, no-inhibitor, Wort, LY or PD, without IGF-I = 100%). Data are presented as mean \pm SEM ($n = 4$ replicates per treatment) for a representative experiment (of four or five separate experiments). *Significant increase above controls ($P < 0.05$; ANOVA); #significant inhibition of the IGF-I response ($P < 0.05$; ANOVA; decrease relative to IGF-I treatment); NS, not significant.

(versus IGF-I without inhibitor) inhibit the luteal cell response to IGF-I.

In vivo studies

Effects of IGF-I infusion in vivo. IGF-I (or vehicle, control) was infused (for 1–2 min) and the progesterone concentrations examined in ovarian and ear vein blood samples collected for 2 h in experiments designed to examine the steroidogenic responses to IGF-I *in vivo*. The data indicated that vehicle infusion (control treatment) had no significant effect on progesterone secretion (ovarian blood, Fig. 4; ear vein blood, data not shown). However, it was noted that at 45–120 min, serum progesterone concentrations were significantly reduced compared with pre-infusion concentrations in ovarian blood (Fig. 4). Infusion of IGF-I induced a significant increase (130% ; $P < 0.05$ versus pre-infusion concentrations) in progesterone concentrations in ovarian blood during the first 45 min after infusion (Fig. 4). Subsequently, progesterone concentrations returned to pre-infusion concentrations at 45–120 min (Fig. 4). Although progesterone concentrations were increased slightly during the 0–45 min after IGF-I infusion as measured in ear vein samples, this effect was significant (116%) only in one animal (data not shown).

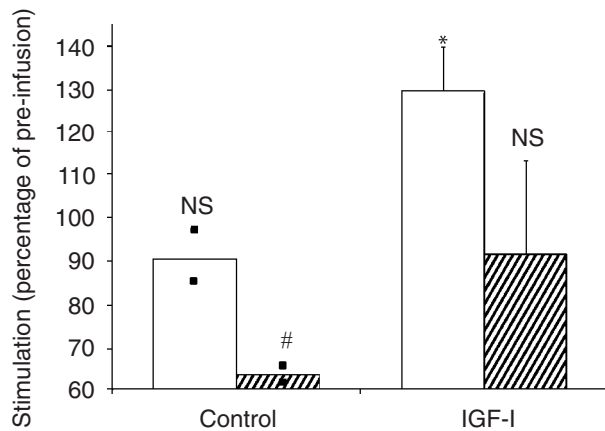


Fig. 4. Steroidogenic response of pig corpora lutea to insulin-like growth factor I (IGF-I) *in vivo*. IGF-I (6 µg per ovary) or vehicle (control) was infused into the ovarian vasculature and progesterone concentrations were measured in ovarian blood samples collected every 5–15 min from about 30 min before infusion (–30 to 0 min = pre-infusion), up to 120 min after infusion. Data are presented as the stimulation of progesterone secretion, expressed as a percentage, relative to pre-infusion (that is, –30 to 0 min = 100%) as measured at 0–45 min (□) and 45–120 min (▨) after infusion. Data are presented as the mean of two animals (vehicle, control) and the mean ± SEM of three animals (IGF-I), examined on day 7 of the oestrous cycle. The values for each of the individual control animals are indicated by (■). *Significant increase above pre-infusion values ($P < 0.05$; ANOVA); #significant decrease below pre-infusion values ($P < 0.05$; ANOVA); NS, not significant.

Progesterone concentrations in ear vein blood samples were not significantly changed at 45–120 min after infusion of IGF-I compared with pre-infusion values (data not shown).

Discussion

This study was designed to assess the function of IGF-IR and the roles of endogenous IGF-I and IGF-BPs on the steroidogenic activity of pig corpora lutea. In addition, the signal transduction pathway involved in mediating the steroidogenic actions of IGF-I on pig luteal cells was examined.

The function of the luteal IGF-IR at different stages of the oestrous cycle was examined by monitoring the capacity of IGF-I agonists to increase progesterone production by luteal cells in culture. As IGF-IR concentrations (protein) in pig corpora lutea are highest at the beginning of the oestrous cycle (days 4–10) and decline thereafter (Ge *et al.*, 2000a), it was anticipated that IGF-I would be more stimulatory to luteal cells on days 4–10 than on subsequent days. Indeed, IGF-I elicited dose-dependent increases in progesterone secretion by luteal cells on days 4 and 7, but not on days 10–16, partially confirming the expectation that response to IGF-I *in vitro* would correlate with

luteal IGF-IR concentrations. However, the lack of a significant response on day 10 was unexpected, especially as IGF-IR concentrations remain increased on this day (Ge *et al.*, 2000a). Potential explanations for the lack of IGF-I response on day 10 are that the IGF-IR was not functioning optimally or that there were deficiencies in one or more components of the IGF-I–IGF-IR signal transduction pathway leading to increased progesterone secretion. Future studies will be directed towards examining these and other possible mechanisms underlying the decline in luteal sensitivity to IGF-I on days 10–16.

Results from the present study confirm reports showing that IGF-I can stimulate luteal cell steroidogenesis in various species including rats (Parmer *et al.*, 1991), rabbits (Constantino *et al.*, 1991), humans (Apa *et al.*, 1996), cows (McArdle and Holtorf, 1989; Sauerwein *et al.*, 1992) and pigs (Huang *et al.*, 1992; Yuan and Lucy, 1996). However, the present study, which examined IGF-I responsiveness of luteal cells collected at different stages of the oestrous cycle in pigs, successfully (on days 4, 7, 13, 15 or 16) correlated functional (steroidogenic) activity of the luteal IGF-I receptor with IGF-IR concentrations. To the authors' knowledge, this has not been reported previously in pigs or any other species. Thus, these findings represent an important advance in the understanding of the role played by IGF-I in corpora lutea development and function.

In addition, the present study shows that large luteal cells respond to IGF-I with increased progesterone secretion in a dose-dependent manner, whereas steroidogenesis by small luteal cells is not significantly increased by IGF-I. This confirms the findings of Yuan and Lucy (1996) and is also consistent with the observations that IGF-IR mRNA and protein are preferentially expressed in large luteal cells in pigs (Ge *et al.*, 2000a). Thus, these data strongly indicate that IGF-I is an important regulator of steroidogenesis in large luteal cells in pigs, particularly during the early stages (days 4 and 7) of the luteal phase. However, as expression of IGF-IR mRNA and protein in small luteal cells is low (Ge *et al.*, 2000a), and small luteal cells show a tendency for increased steroidogenesis (the present study), these cells cannot be excluded as potential targets for the actions of IGF-I.

Finally, the present study, using an ovarian infusion model system, provides confirmation *in vivo* of the steroidogenic actions of IGF-I on pig corpora lutea. In addition, the acute (within 45 min) nature of the steroidogenic response to IGF-I has been demonstrated. The data are in agreement with studies carried out in bovine corpora lutea using *in vivo* microdialysis, in which the authors also demonstrated an acute (about 30 min) increase in progesterone secretion (Sauerwein *et al.*, 1992). Preliminary studies examined the time course of the action of IGF-I on pig luteal cells *in vitro* and found that IGF-I has significant stimulatory effects on

progesterone secretion as early as 4 h after administration (Z. Ge, V. Hedgpeth and J. E. Gadsby, unpublished), further demonstrating the acute nature of the steroidogenic response to IGF-I.

The second objective of this study was to explore the role of endogenous IGF-I and IGF-IR activity by the use of blocking antibodies in cell cultures. Thus, the study attempted to block endogenous IGF-I via the addition of a monoclonal antibody to IGF-I (Sm 1.2B), which has been shown to block the bioactivity of IGF-I in pig granulosa cell cultures (Mondschein *et al.*, 1989). In addition, an antibody known to block the type-I IGF receptor was used (α IR3; Rohlik *et al.*, 1987), thus preventing the IGF-I activation pathway. No significant decrease in progesterone secretion by luteal cells cultured with either the IGF-I or IGF-IR antibody, at any dose, or on any day of the oestrous cycle was shown. One possible explanation for these observations is that the endogenous amounts of IGF-I in the luteal cell cultures were insufficient to exert a stimulatory response on steroidogenesis. On the basis of data obtained from other studies (Nicholson *et al.*, 1999; Z. Ge, W. E. Nicholson, V. Hedgpeth and J. E. Gadsby, unpublished) in which the IGF-I protein content of pig corpora lutea was measured (by radioimmunoassay), it is estimated that IGF-I in the luteal cell cultures described here would be about 55 and about 37 pg per well (or per ml) on days 4 and 7, respectively; amounts probably too low to activate IGF-IR (ED_{50} value 4–7 ng ml⁻¹). It is not known how much IGF-I is secreted by pig luteal cells during 20–24 h of culture, but the present study strongly indicates that the concentrations of endogenous plus secreted IGF-I are too low to drive basal steroidogenesis in this cell culture system.

The role of endogenous IGFBPs in the luteal IGF-I response mechanism was examined. The presence of endogenous IGFBPs within pig corpora lutea has been documented by demonstrating the expression of mRNAs for IGFBPs 2–5 (Gadsby *et al.*, 1996), and by showing that the amounts of luteal IGFBP-3 protein in particular are highest on days 4–7 of the oestrous cycle (Ge *et al.*, 1998; Z. Ge, W. E. Nicholson, V. Hedgpeth and J. E. Gadsby, unpublished). Although it is clear that IGFBPs are important in regulating the bioavailability of IGF-I to its receptors, it is still controversial as to whether such actions facilitate or inhibit the action of IGF-I on its receptor (Baxter, 2000; Grimberg and Cohen, 2000). Thus, although there is some evidence indicating that IGFBPs can facilitate the actions of IGF-I on target cells, most of the data on IGFBPs indicates that they have inhibitory effects on the actions of IGF-I (Baxter, 2000; Grimberg and Cohen, 2000). For example, with regard to IGFBP-3, maximal IGFBP-3 mRNA expression was seen in corpora lutea undergoing luteolysis in rats (Erickson *et al.*, 1993). In addition, PGF_{2 α} stimulated, whereas PGE₂ inhibited, IGFBP-3 (protein) production by luteinizing pig granulosa cells in culture (Grimes *et al.*, 1993). Furthermore, the present study has shown

that the IGFBP-3 protein content was increased in pig corpora lutea after PGF_{2 α} treatment *in vivo* (Nicholson *et al.*, 1999). Finally, some recent data in bovine corpora lutea further indicate the inhibitory roles of IGFBP-2 and -3 on IGF-I binding to luteal (IGF-IR) receptors (Brown and Braden, 2001). Taken together, these results indicate that IGFBPs (in particular IGFBP-3) play an inhibitory role in regulating the action of IGF-I in the corpora lutea. The present study sought to examine the role of endogenous IGFBPs in the luteal IGF response mechanism by using the IGF-I analogue, long R₃-IGF-I, which binds to IGFBPs with lower affinity than does IGF-I (Baxter, 2000). As long R₃-IGF-I binds to IGF-IR, but minimally to IGFBPs (compared with IGF-I), a greater steroidogenic response to this analogue was expected compared with IGF-I. The results show that progesterone secretion by luteal cells is increased by both long R₃-IGF-I and IGF-I to the same degree, indicating that endogenous IGFBPs are not acting as inhibitors of IGF-I actions on pig luteal cells in this cell culture system. Nevertheless, in some related *in vitro* studies it has been shown that exogenous IGFBP-3 does inhibit the steroidogenic responses of pig luteal cells to exogenous IGF-I, and that an antibody to IGFBP-3 given alone is stimulatory to luteal progesterone secretion (Z. Ge, W. E. Nicholson, V. Hedgpeth and J. E. Gadsby, unpublished). Taken together, these observations indicate that IGFBP-3 can have inhibitory actions on the interactions of IGF-I with its receptors in pig corpora lutea, although the role that the increased IGFBP-3 on days 4–7 (Ge *et al.*, 1998; Z. Ge, W. E. Nicholson, V. Hedgpeth and J. E. Gadsby, unpublished) may play in modulating the action of IGF-I in developing corpora lutea in pigs is unclear.

The fourth objective of the present study was to explore the signal transduction pathway of the steroidogenic actions of IGF-I in pig corpora lutea. This study examined the effects of inhibitors of PI-3-kinase (Wortmannin and LY 294002) and MAP-kinase (PD 98059) pathways in pig luteal cells, on IGF-I stimulated progesterone secretion. The data presented show that Wortmannin and LY 294002 both significantly inhibit the stimulatory actions of IGF-I on pig luteal cell steroidogenesis, whereas PD 98059 was without effect, strongly indicating the involvement of the PI-3-kinase pathway in mediating the steroidogenic actions of IGF-I in pig corpora lutea. Although there are few, if any, reports of studies on the pathways of the steroidogenic actions of IGF-I in pig ovaries, there is evidence that PI-3-kinase and protein kinase B (or Akt) are involved in mediating the cell survival pathway in pig granulosa cells (Westfall *et al.*, 2000).

In summary, the present study has shown that IGF-I agonists stimulate luteal progesterone production *in vitro* on days 4 and 7, but not on days 10–16 of the oestrous cycle, demonstrating the functional competence of IGF-IR and the signal transduction pathway during the early luteal phase. This was confirmed by *in vivo*

studies conducted on day 7. In addition, this study has shown that large luteal cells are the primary target for the steroidogenic actions of IGF-I. The data also indicate that endogenous IGF-I is probably too low to activate steroidogenesis, and that endogenous IGF-BPs do not appear to inhibit the action of IGF-I *in vitro*. Finally, this study demonstrated that IGF-I acts to promote steroidogenesis via the PI-3-kinase pathway. Future studies will be directed towards a more detailed examination of the IGF-I signal transduction pathways and mediators of increased luteal steroidogenic activity, to determine the mechanism by which luteal IGF-I sensitivity changes during the oestrous cycle in pigs.

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