Disruption of imprinting in cloned mouse fetuses from embryonic stem cells

H. Ogawa1, Y. Ono2, N. Shimozawa3, Y. Sotomaru3, Y. Katsuzawa2, H. Hiura1, M. Ito3 and T. Kono1∗

1Department of BioScience, Tokyo University of Agriculture, 1-1-1 Sakuragaoka, Setagaya-ku, Tokyo 156-8502, Japan; 2Department of Animal Science, Tokyo University of Agriculture, 1737 Funako, Atsugi-shi, Kanagawa 243-0034, Japan; and 3Central Institute for Experimental Animals, 1430 Nogawa, Miyamae-ku, Kawasaki-shi, Kanagawa 216-0001, Japan

Cloned mice typically display abnormal development, such as overgrowth of fetuses and placentae. Quantitative expression analysis of eight imprinted genes (H19, Igf2, Igf2r, Air, Peg1/Mest, Peg3, Nuronatin (Nnat) and Ndn) and an alternate transcript of Igf2 (P0) in embryonic stem cloned fetuses and placentae at days 9.5, 12.5 and 17.5 after mating was carried out by real time PCR to investigate whether epigenetic modification of imprinted genes is responsible for overgrowth of the fetus and placental hypertrophy. In addition, the methylation pattern through the bisulphite sequencing method in differentially methylated regions of H19 and Igf2r was examined in day 9.5 fetuses and placentae. The results showed clearly that the expression of H19 gene decreased in cloned fetuses at days 12.5 and 17.5 after mating and in placentae at day 17.5 after mating, and Igf2 was also repressed in fetuses at days 9.5 and 12.5 after mating and in placentae at day 17.5 after mating. In contrast, the transcription of P0, which is a placental-specific transcript variant of Igf2, increased at more than four times the control in cloned placentae at day 12.5 after mating. Day 9.5 fetuses that have developed normally revealed only hypermethylated alleles in the H19 differently methylated region (DMR), and both hyper- and hypomethylated alleles in the Igf2r DMR2. These results show that inappropriate reprogramming in some imprinted genes affects the development of cloned embryos, and that aberrant P0 Igf2 transcription in particular may cause the overgrowth of cloned fetuses and placentae.

Introduction

Nuclear transfer technology has shown that transfer of somatic cells to enucleated unfertilized oocytes can result in successful development of cloned individuals in sheep (Wilmut et al., 1997), cattle (Kato et al., 1998), pigs (Onishi et al., 2000), goats (Baguisi et al., 1999), cats (Shin et al., 2002), rabbits (Chesne et al., 2002) and mice (Wakayama et al., 1998). However, the efficiency of somatic cell nuclear transfer in all species is still extremely low regardless of the origin of the cells. Studies so far have shown that cloned individuals often suffer from a wide range of severe malformations, such as overgrowth of the fetus and placenta (Ono et al., 2001a,b; Shimozawa et al., 2002a) and deficient immune systems (Renard et al., 1999; Wells et al., 1999; Ogonuki et al., 2002).

Embryonic stem (ES) cells that exhibit high pluripotency and that can differentiate into all tissues and organs of the fetus, but contribute only poorly to extraembryonic tissues in chimaeric mice, have been used as donor cells for nuclear transfer in attempts to improve cloning efficiency. Indeed, cloning from ES cells is more efficient than cloning from somatic cells (Jaenisch et al., 2002), but ES cloned mice are accompanied by the typical malformations seen in somatic cloned mice, such as placental hypertrophy and overgrowth of the fetus (Wakayama and Yanagimachi, 1999; Eggan et al., 2001; Ono et al., 2001a). It has been reported that placental hypertrophy is accompanied by abnormal formation of spongiotrophoblasts and the labyrinthine layers (Shimozawa et al., 2002a). These abnormal formations may interfere with utero–placental circulation and cause death before and after birth in cloned mice.

It has been argued that it is the irregular expression of imprinted and non-imprinted genes in cloned mice that leads to the abnormal development of the fetus and placenta (Humphryes et al., 2001; Rideout et al., 2001; Inoue et al., 2002). In these studies, aberrant expression in cloned tissues was particularly evident for four imprinted genes, H19, Igf2, Peg1/Mest and Meg1/Grb10 (Humphryes et al., 2001; Inoue et al., 2002). Proper epigenetic modifications, such as DNA methylation, by which imprinted gene expression is generally repressed,
is necessary for the development of fertilized eggs into individuals. Otherwise, inappropriate reprogramming of epigenetic modifications after nuclear transfer may cause malformations in cloned embryos. The methylation status of DNA cytosine residues has been analysed (Humpherys et al., 2001; Kang et al., 2001a–c; Xue et al., 2002). Recent studies showed that inappropriate reprogramming of epigenetic modifications is evident, which may in turn affect mechanisms underlying the low cloning efficiency and the abnormal development (Rideout et al., 2001). For example, in cloned calves, both X chromosomes were active with a differently methylated region (DMR) in which the Xist gene was unmethylated (Xue et al., 2002). Expression of the H19 gene was depleted in an ES cloned mouse in which the DMR of the gene was hypermethylated (Humpherys et al., 2001). In contrast, parent specific expression of imprinted genes was maintained in somatic cloned mice (Inoue et al., 2002). However, to date, direct evidence that aberrant expressions of imprinted genes cause abnormal development in cloned embryos is lacking.

Shimozawa et al. (2002a) obtained an ES cell line that resulted in overgrowth of the fetus and severe placental hypertrophy. It was concluded that further insight into these typical occurrences in ES cloned embryos might be achieved by using this line to determine the expression of imprinted genes and the DNA methylation status of the DMR region during development after implantation. Therefore, the aim of the present study was to examine the expression of eight imprinted genes: H19, Igf2, Igf2r, Air, Peg1/Mest, Peg3, Nuronatin (Nnat) and Ndn, and an alternate transcript of Igf2 (P0) in ES cloned fetuses at days 9.5, 12.5 and 17.5 after mating by quantitative expression analysis using real-time PCR. Moreover, methylation of CpG sites in DMRs of H19 and Igf2r genes was analysed, as these two genes are closely related to these typical occurrences in ES cloned embryos.

Materials and Methods

Preparation of donor cells

TT2 ES cells, which were derived from B6C6F1 male mice and had been targeted at the oviduct-specific glycoprotein gene, were used as donor cells. ES cells were cultured in knockout-Dulbecco’s modified Eagle’s medium (Gibco BRL, Grand Island, NY) with 15% knockout serum replacement (Gibco BRL), 10% fetal bovine serum (Gibco BRL, Grand Island, NY) with 15% hypoxanthine, aminopterin, thymidine (HAT) medium (Gibco BRL, Grand Island, NY) and 1% (v/v) non-essential amino acid solution (Gibco BRL). Cells were cultured in knockout-Dulbecco’s modified Eagle’s medium (Gibco BRL, Grand Island, NY) with 1% (v/v) non-essential amino acid solution (Gibco BRL) and 15% fetal bovine serum (Gibco BRL) and 1% (v/v) non-essential amino acid solution (Gibco BRL) with 15% hypoxanthine, aminopterin, thymidine (HAT) medium (Gibco BRL, Grand Island, NY) for 3 days on a feeder layer derived from fibroblasts of day 15 fetuses. ES cells were synchronized at metaphase with 0.5 μg nocodazole ml⁻¹ (Sigma Chemical Co., St Louis, MO) for 2 h. Metaphase-arrested cells were selected and used as donors for oocyte reconstruction.

Embryo manipulation

The superovulated oocytes from B6C6F1 female mice were collected 14 h after hCG administration. After enucleation of the metaphase II chromosomes, an ES cell arrested at metaphase was introduced into the perivitelline space of the enucleated oocyte with inactivated Sendai virus (HVJ), which induces fusion within 20 min. Reconstructed embryos were cultured in CZB (Chatot et al., 1990) for 2 h and activated artificially with 10 mmol strontium l⁻¹ for 6 h. These reconstructed embryos were cultured in CZB for 4 days. Control embryos were produced by pronuclear transfer, in which C57BL/6N oocytes were fertilized in vitro with spermatozoa from CBA/N males and the resultant pronuclei were transferred into enucleated B6C6F1 eggs. Blastocysts derived from in vitro culture were transferred to the uterine horns of ICR female mice on day 2.5 of pseudopregnancy. Four to six blastocysts were transferred to exclude the effect of litter size on fetuses and placentae (Shimozawa et al., 2002a). At days 9.5, 12.5, 17.5 and 19.5 after mating, pregnant mice were killed by cervical dislocation and the fetuses were collected.

Quantitative expression analysis of imprinted genes

Total RNA was isolated from fetuses at days 9.5, 12.5 and 17.5 after mating and from placentae at days 12.5 and 17.5 after mating with SV Total RNA Isolation System (Promega, Madison, WI). Total RNA was reverse transcribed to cDNA using SUPERSCRIPT II (Gibco BRL). cDNA synthesized from either 0.01 or 0.1 μg total RNA was applied to real-time PCR for quantitative expression analysis of β-actin, which is an internal control gene, eight imprinted genes (Igf2, H19, Igf2r, Air, Peg1/Mest, Peg3, Nuronatin (Nnat) and Ndn, and an alternate transcript of Igf2 (P0) in ES cloned fetuses at days 9.5, 12.5 and 17.5 after mating by quantitative expression analysis using real-time PCR. Moreover, methylation of CpG sites in DMRs of H19 and Igf2r genes was analysed, as these two genes are closely related to fetal development.

The superovulated oocytes from B6C6F1 female mice were collected 14 h after hCG administration. After enucleation of the metaphase II chromosomes, an ES cell arrested at metaphase was introduced into the perivitelline space of the enucleated oocyte with inactivated Sendai virus (HVJ), which induces fusion within 20 min. Reconstructed embryos were cultured in CZB (Chatot et al., 1990) for 2 h and activated artificially with 10 mmol strontium l⁻¹ for 6 h. These reconstructed embryos were cultured in CZB for 4 days. Control embryos were produced by pronuclear transfer, in which C57BL/6N oocytes were fertilized in vitro with spermatozoa from CBA/N males and the resultant pronuclei were transferred into enucleated B6C6F1 eggs. Blastocysts derived from in vitro culture were transferred to the uterine horns of ICR female mice on day 2.5 of pseudopregnancy. Four to six blastocysts were transferred to exclude the effect of litter size on fetuses and placentae (Shimozawa et al., 2002a). At days 9.5, 12.5, 17.5 and 19.5 after mating, pregnant mice were killed by cervical dislocation and the fetuses were collected.

Quantitative expression analysis of imprinted genes

Total RNA was isolated from fetuses at days 9.5, 12.5 and 17.5 after mating and from placentae at days 12.5 and 17.5 after mating with SV Total RNA Isolation System (Promega, Madison, WI). Total RNA was reverse transcribed to cDNA using SUPERSCRIPT II (Gibco BRL). cDNA synthesized from either 0.01 or 0.1 μg total RNA was applied to real-time PCR for quantitative expression analysis of β-actin, which is an internal control gene, eight imprinted genes (Igf2, H19, Igf2r, Air, Peg1/Mest, Peg3, Nuronatin (Nnat) and Ndn, and an alternate transcript of Igf2 (P0) in ES cloned fetuses at days 9.5, 12.5 and 17.5 after mating by quantitative expression analysis using real-time PCR. Moreover, methylation of CpG sites in DMRs of H19 and Igf2r genes was analysed, as these two genes are closely related to fetal development.

Materials and Methods

Preparation of donor cells

TT2 ES cells, which were derived from B6C6F1 male mice and had been targeted at the oviduct-specific glycoprotein gene, were used as donor cells. ES cells were cultured in knockout-Dulbecco’s modified Eagle’s medium (Gibco BRL, Grand Island, NY) with 15% knockout serum replacement (Gibco BRL), 10% fetal bovine serum (Gibco BRL) and 1% (v/v) non-essential amino acid solution (Gibco BRL) and 55 μmol 2-mercaptoethanol l⁻¹ (Wako Pure Chemical Industries, Osaka) for 3 days on a feeder layer derived from fibroblasts of day 15 fetuses. ES cells were synchronized at metaphase with 0.5 μg nocodazole ml⁻¹ (Sigma Chemical Co., St Louis, MO) for 2 h. Metaphase-arrested cells were selected and used as donors for oocyte reconstruction.

Embryo manipulation

The superovulated oocytes from B6C6F1 female mice were collected 14 h after hCG administration. After enucleation of the metaphase II chromosomes, an ES cell arrested at metaphase was introduced into the perivitelline space of the enucleated oocyte with inactivated Sendai virus (HVJ), which induces fusion within 20 min. Reconstructed embryos were cultured in CZB (Chatot et al., 1990) for 2 h and activated artificially with 10 mmol strontium l⁻¹ for 6 h. These reconstructed embryos were cultured in CZB for 4 days. Control embryos were produced by pronuclear transfer, in which C57BL/6N oocytes were fertilized in vitro with spermatozoa from CBA/N males and the resultant pronuclei were transferred into enucleated B6C6F1 eggs. Blastocysts derived from in vitro culture were transferred to the uterine horns of ICR female mice on day 2.5 of pseudopregnancy. Four to six blastocysts were transferred to exclude the effect of litter size on fetuses and placentae (Shimozawa et al., 2002a). At days 9.5, 12.5, 17.5 and 19.5 after mating, pregnant mice were killed by cervical dislocation and the fetuses were collected.

Quantitative expression analysis of imprinted genes

Total RNA was isolated from fetuses at days 9.5, 12.5 and 17.5 after mating and from placentae at days 12.5 and 17.5 after mating with SV Total RNA Isolation System (Promega, Madison, WI). Total RNA was reverse transcribed to cDNA using SUPERSCRIPT II (Gibco BRL). cDNA synthesized from either 0.01 or 0.1 μg total RNA was applied to real-time PCR for quantitative expression analysis of β-actin, which is an internal control gene, eight imprinted genes (Igf2, H19, Igf2r, Air, Peg1/Mest, Peg3, Nuronatin (Nnat) and Ndn, and an alternate transcript of Igf2 (P0) in ES cloned fetuses at days 9.5, 12.5 and 17.5 after mating by quantitative expression analysis using real-time PCR. Moreover, methylation of CpG sites in DMRs of H19 and Igf2r genes was analysed, as these two genes are closely related to fetal development.
produces multiple transcripts from alternative promoters resulting in sense and anti-sense transcripts (Moore et al., 1997). Primers for first-round PCR were selected to avoid amplification of anti-sense transcripts. The following primers were used: 5'-CTTTGGAGGGGCTGCTAATA-3' at P0 and 5'-CGACCCCGGCGACGCAGG-3' at E6. PCR amplification was carried out with a HotStarTaq DNA Polymerase (QIAGEN, Hilden). The PCR was performed by an initial denaturation at 95°C for 15 min and 65 cycles of 95°C for 1 min, 65°C for 1 min and 72°C for 2 min. A final period of extension was carried out for 12 min. The PCR products were used for second-round PCR using the LightCycler System. Expression of each gene was evaluated on the basis of the expression of β-actin for individual samples, and represented a relative percentage of the expression of controls at day 9.5 after mating.

Bisulphite analysis

Fetuses and extraembryonic tissues at day 9.5 after mating were incubated in 163 μl lysis solution (6 mol guanidine hydrochloride 1-1 (140 μl), 7.5 mol ammonium acetate 1-1 (10 μl), 20% (w/v) sarkosyl (10 μl) and 10 mg proteinase K ml-1 (3 μl)) at 55–60°C for 2 h. The isolated DNA was treated with sodium bisulphite using a CpGenome DNA Modification kit (INTERGEN, Purchase, NY). The bisulphite-modified DNA was amplified by PCR. The methylated status of H19 DMR at P0 and 5-4 (140 μl) was 0.1 ± 0.01 g (n = 5), which was about 70% of the final size at term. In contrast, the ES cloned placentae showed excessive growth between day 12.5 and day 17.5 after mating, reaching 0.54 ± 0.11 g (n = 5), almost fourfold the mass of the controls (Fig. 1a,b,d).

Expression of imprinted genes in ES cloned fetuses and placentae

The expression of imprinted genes in the fetuses at days 9.5, 12.5 and 17.5 after mating and the placenta at days 12.5 and 17.5 after mating was evaluated quantitatively by real-time PCR and represented as relative levels to the level of the expression of controls at day 9.5 after mating (Fig. 2a). The genes were classified into four groups on the basis of their normal expression during development (H19 and Ndn); and (iv) gene expression increases with development (Igf2, Igf2r and Peg3, were expressed at significantly lower amounts except Igf2r, whereas the other genes
DMR methylation pattern in H19 and Igf2r

Monoallelic expression of imprinted genes is regulated by the methylated status of CpG sites in the DMR of each gene. The methylation status of DMR regions of the H19 and Igf2r genes, which are imprinted and expressed solely from the maternal allele, in five cloned fetuses and the extraembryonic tissues at day 9.5 after mating were assessed (Fig. 3). In clone 21, the fetus was not turned, with no heartbeat and developmental retardation; in clone 8, the fetus was not turned, with heartbeat and developmental retardation; in clone 13, the fetus was turned, with no heartbeat and developmental retardation; and in clones 23 and 24, the fetuses were turned, with heartbeat and normal phenotype.

In control fetuses and the extraembryonic tissues, both hyper- and hypomethylated alleles in the DMR of the H19 gene were detected, which could be interpreted as paternal and maternal alleles, respectively (Fig. 4a). The methylation status was divided into two patterns in the cloned fetuses and the extraembryonic tissues. Both hyper- and hypomethylated alleles of the DMR were detected in clones 8 and 21, which were retarded and not turned at recovery. In contrast, only the hypermethylated allele of the DMR was detected in clones 13, 23 and 24, which were turned at recovery. The methylation status of the DMR in the extraembryonic tissues was similar to those of the fetus. The donor ES cells (TT2 line) were analysed to compare the methylation status with that of cloned fetuses. The data showed that the ES cells contain both hyper- and hypomethylated alleles with three predominantly methylated (3375, 3462 and 3580) and unmethylated (3302, 3384 and 3563) CpG sites, respectively.

Methylation analysis of the Igf2r gene detected both hyper- and hypomethylated alleles of DMR2 in controls and also in cloned fetuses 13, 23 and 24, although the hypermethylated allele was dominant in clone 13 (Fig. 4b). In contrast, cloned fetuses 8 and 21 contained only unmethylated alleles except in the one DNA clone in fetus 8. In the extraembryonic tissues, both hyper-and hypomethylated alleles were detected except in the one case, clone 21, in which the DMR2 was completely unmethylated. In the ES cells, both hyper- and hypomethylated alleles of the DMR2 were detected except for one CpG site (site no. 1002), which was completely methylated.

Discussion

Studies to date have determined that ES and somatic cloned animals display overgrowth of the fetus and placenta (Wakayama and Yanagimachi, 1999; Eggan et al., 2001; Ono et al., 2001a). The present study produced ES cloned fetuses using the ES cell line that is known to result in overgrowth of the fetus and placenta when used in cloning (Shimozawa et al., 2002a). The expression of
imprinted genes was examined at early (day 9.5 after mating), mid- (day 12.5 after mating) and late (day 17.5 after mating) gestation to clarify such developmental abnormalities. As expected, these cloned fetuses were 30% heavier than controls at day 17.5 and day 19.5 after mating. Furthermore, the associated placentae were remarkably increased between day 12.5 and day 17.5 after mating and were four times heavier than those of controls. This finding is unusual as the mass of placentae at day 12.5 after mating was up to 80% of the final size at day 17.5 after mating in controls.

Eight imprinted genes were analysed quantitatively to understand mechanisms underlying the overgrowth of fetuses and placentae. The results showed that the H19 gene was completely repressed through development in cloned fetuses and the placentae. Mouse H19 and

---

**Fig. 2.** Quantitative analysis of imprinted gene expression from (a) fetuses and (b) placentae of control (□) and embryonic stem cloned mice (■) by real-time PCR. The mean expression was calculated as a percentage of the expression of controls at day 9.5 after mating. Standard errors of means are indicated by bars. Asterisks denote significant difference from the control (*P < 0.05 and **P < 0.01).
Igf2 genes, which are located on the distal portion of chromosome 7 (Caspary et al., 1998), exhibit reciprocal expression by the parental allele. When one of these genes is transcribed, another gene is silenced. Humphreys et al. (2001) reported that the H19 gene expression was repressed in the ES cloned pups in which the DMR was hypermethylated and expression of the Igf2 gene was increased. Unexpectedly, the depleted expression of H19 was not associated with an increase of Igf2 expression in cloned fetuses at day 9.5 and day 12.5 after mating. Thus, transcription of the Igf2 gene in the ES cloned fetuses at day 9.5 and day 12.5 after mating may be downregulated. Further, the enhancer competition model between the Igf2 and H19 genes, in which these two genes compete for the shared enhancer elements located downstream of H19, may be disrupted. Furthermore, the Igf2r gene, which is maternally expressed and interferes with the mitogenic effect of Igf2, was expressed at half the amount of the controls at day 12.5 after mating. The expression of Air, the anti-sense mRNA of Igf2r, which could affect the function of the Igf2r gene (Lyle et al., 2000), in clones did not significantly differ from that in controls at days 9.5 and 12.5 after mating; however, Air was still expressed by day 17.5 after mating in clones, after it had disappeared from controls. These results indicate that lower expression of the Igf2r gene with expression of the Air at day 17.5 after mating in clones can induce overgrowth of the fetus in the late stage of gestation.

Gene expression in the placenta was also analysed to obtain further insight into the overgrowth phenomena. The expression of the H19 and Igf2 genes was reduced at day 17.5 after mating in cloned placenta. This expression is not inconsistent with the present theory for the regulation of the two genes. Expression of the Igf2r gene in the placenta in clones appeared the same as in controls. From these results, it is difficult to deduce a clear reason for the overgrowth in the clones. The expression of the P0 domain of Igf2, which contains the coding exons 4–6, and expresses specifically from the placenta (Moore et al., 1997) with predominant expression in the labyrinthine trophoblast cells, was examined (Constancia et al., 2002). The absence of the P0 transcript in a gene deletion experiment resulted in deficiency in growth of the placenta and reduction in permeability for nutrients and led to 69% of normal birth weight (Constancia et al., 2002). In the present study, the P0 transcript in the placenta of clones was expressed at four times the amount expressed in controls at day 12.5 after mating, indicating that the overexpression of the Igf2 P0 transcript is responsible for the placental overgrowth. This overexpression may lead to fetal overgrowth. Furthermore, considering the placental hypertrophy of the cloned fetuses, the P0 transcript from the placenta could be concerned in the overgrowth of the fetus.

Although the mechanism for aberrant expression of some imprinted genes in the cloned fetus is unclear, several factors including histone acetylation and DNA methylation may be involved (Bird and Wolffe, 1999; Reik et al., 2001). Studies so far have shown that DNA methylation status differs between fertilized and cloned embryos in cattle (Kang et al., 2001a). The present study analysed DNA methylation of the H19 DMR and Igf2r DMR2 in the fetuses at day 9.5 after mating, and found that methylation status differed depending on phenotype.
Fig. 4. (a) Methylation patterns in differently methylated region (DMR) of the mouse H19 gene (GenBank accession number AF049091); analysed region is from 3248 to 3615 of the gene containing 20 CpG sites. A filled and open circle corresponds to methylated CpG and unmethylated CpG sites, respectively. (b) Methylation patterns in DMR2 of the Igf2r gene (GenBank accession number; L06446); analysed region is from 964 to 1413 of the gene containing 30 CpG sites. \( n \): number of DNA clones.
In the apparently normal-looking fetuses, H19 DMR was completely methylated (that is, not normal), but in the retarded fetuses, both methylated and hypomethylated DNA clones were observed (as expected in the normal situation). This finding supports the results of the present study on gene expression analysis and leads to the idea that the hypermethylated H19 DMR causes complete repression of the gene throughout development. On the contrary, in the Igf2r DMR2, all DNA clones except for one were apparently unmethylated in the developmentally retarded fetuses, indicating that this unmethylated status may be the result of overexpression of Igf2r, and may be one of the reasons for abnormal development in cloned embryos.

The methylation status of cytosine residue is not stable in specific genes when cells are cultured in vitro (Doherty et al., 2000). Sasaki et al. (1995) showed that preimplantation mouse embryos cultured in vitro led to biallelic H19 expression in the extraembryonic tissues. The ES cells used in the present study as donor nuclei for nuclear transfer maintained the methylated and unmethylated alleles of H19 DMR; however, three each of the 20 CpG sites examined, three were apparently hypermethylated and three were apparently hypomethylated. Comparing the methylation status of controls with that of cloned fetuses shows that methylation of CpG sites changes some time after embryo reconstruction. The expression of imprinted genes and methylated status were also different between ES cell lines and among subclones (Humphreys et al., 2001). The expression of imprinted genes was affected not only by the culture condition of donor cells, but also by nuclear transfer and subsequent embryo culture (Doherty et al., 2000; Khosla et al., 2001; Young et al., 2001). However, how and when DNA methylation status of donor cells is modified in the cloned embryos is still unclear. The findings from the present study indicate that aberrant expression of the imprinted genes is correlated with altered methylation status, which perhaps influences the developmental ability of ES cloned embryos. It may be valuable to determine the superior epigenetic modifications for donor cells and to establish a procedure for selecting and sorting them to produce cloned animals efficiently.

This work was supported by grants from the Ministry of Education, Science, Culture and Sports of Japan, the Ministry of Agriculture of Japan, the Japanese Society for Promotion of Science and Mecrogen Inc. (Korea).

References

Baguati A, Behboodi E, Melican DT et al. (1999) Production of goats by somatic cell nuclear transfer Nature Biotechnology 17 456–461


Reik W, Dean W and Walter J (2001) Epigenetic reprogramming in mammalian development Science 293 1089–1093

Disruption of imprinting in embryonic stem clones


Wells DN, Misica PM and Tervit HR (1999) Production of cloned calves following nuclear transfer with cultured adult mural granulosa cells Biology of Reproduction 60 996–1005


Received 10 April 2003.
First decision 21 May 2003.
Revised manuscript received 23 June 2003.
Accepted 4 July 2003.