Effects of parity and body condition at parturition on endocrine and reproductive parameters of the cow

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Abstract

The effect of parity (multiparous vs primiparous) and body condition score (BCS; < 3.0 or ≥ 3.0, lean vs fat) at parturition on metabolic and endocrine profiles from 1 month before to 2 months after parturition were studied in 42 Holstein cows grazing on improved pastures. BCS and milk production were determined every 2 weeks. Non-esterified fatty acids (NEFA), β-hydroxybutyrate (BHB), insulin, IGF-I, leptin, thyroxine (T4) and 3,3',5-tri-iodothyroinine (T3) were determined in plasma every 10 days. Progesterone was determined three times per week after parturition. Primiparous cows had a lower BCS during the early postpartum period and produced less milk than multiparous animals. Primiparous cows had higher NEFA concentrations and they presented more samples with BHB concentrations of > 1 mmol/l than multiparous cows. Multiparous cows had higher T3, T4 and IGF-I concentrations, while fat cows had higher leptin and IGF-I concentrations. All hormone concentrations were diminished in the first week postpartum. Primiparous cows and fat cows presented a steeper decay of IGF-I and leptin around parturition than multiparous cows and lean cows. While thyroid hormones and IGF-I showed increasing concentrations from approximately day 30, leptin concentrations remained low until the end of the experimental period. The initiation of ovarian cyclicity was delayed in primiparous cows and especially in primiparous lean cows, consistent with longer intervals from parturition to first service and to conception. The endocrine signals most likely to inform the reproductive axis regarding a negative energy balance were IGF-I and leptin.

Introduction

Genetic selection for milk production during the last decades has been associated with decreased reproductive efficiency (Lucy 2001). The resumption of ovarian cyclicity after parturition is closely related to the negative energy balance (EB) in this period; the time to the beginning of the recovery of the EB is positively correlated with the time to first ovulation (Butler et al. 1981). Butler & Smith (1989) found that cows that lost less than 0.5 units of body condition score (BCS) during the first 5 weeks postpartum had higher conception rates at the first service than cows that lost more than 0.5 BCS. The physiological pathways by which the hypothalamic–pituitary–ovarian axis is informed about the energetic status of the animal are complex, and involve several metabolites and hormones, such as the growth hormone (GH)–insulin-like growth factor-I (IGF-I) system, insulin, thyroid hormones and leptin.

It has been proposed that the effect of negative EB on the resumption of ovulation may be mediated by the secretion of IGF-I (Spicer et al. 1990). Although GH concentrations are usually high in early lactating ruminants, the intrahepatic production of its mediator IGF-I is diminished (Chilliard 1999). Circulating concentrations of IGF-I in the peripartum period are good indicators of the capacity of energy-restricted cows to resume cyclicity after parturition (Roberts et al. 1997). Cows with ovulatory estrogen-active follicles have higher circulating IGF-I concentrations during the first 2 weeks than cows with anovulatory follicles (Beam & Butler 1997, 1998). Both insulin and IGF-I are known to stimulate in vitro steroidogenesis and proliferation of bovine thecal and granulosal cell cultures (Spicer et al. 1993, Spicer & Stewart 1996). Likewise, cows that ovulated within 35 days postpartum present higher IGF-I concentrations as well as higher glucose and insulin and lower non-esterified fatty acids.
(NEFA) and β-hydroxybutyrate (BHB) concentrations (Huszenicza et al. 2001).

Cows in postpartum negative EB have lower concentrations of thyroid hormones induced by altering central and peripheral mechanisms (Pethes et al. 1985, Capuco et al. 2001). A role of these hormones in regulating steroidogenesis has been reported (Spicer 2001), but data regarding their effect on ovarian function in vivo are limited and controversial (Huszenicza et al. 2002).

Leptin – secreted by the white adipose tissue – acts as an energy reserve signal for hypothalamic regions that control feeding behavior, metabolism and endocrine function so as to maintain energy homeostasis (Chilliard et al. 2001). In ruminants, as in other species, leptin concentrations vary with changes in body weight (BW) and percentage of body fat (Delavaud et al. 2002). Dairy cows often lose over 60% of their body fat during early lactation (Tamminga et al. 1997, Chilliard 1999), and it has been shown that the leptin concentration declines shortly before parturition (Kadokawa et al. 2000, Block et al. 2001, Liefers et al. 2003). There is less agreement on leptin concentrations after parturition: increasing (Kadokawa et al. 2000), unchanged (Huszenicza et al. 2001), diminished (Block et al. 2001, Holtenius et al. 2003) or a transient increase (Liefers et al. 2003) have been reported. The postpartum leptin reduction is likely to be due in part to the negative EB because plasma leptin remained high in cows not milked after parturition (Block et al. 2001).

There are only a few reports on the relation between leptin concentrations and resumption of cyclic ovarian activity during the postpartum period (Kadokawa et al. 2000, Huszenicza et al. 2001, Liefers et al. 2003). The interval from parturition to first ovulation correlated significantly with the interval from parturition to leptin nadir, but was not correlated with the prepartum, pre- and/or postovulatory leptin values (Kadokawa et al. 2000). Liefers et al. (2003) found that, although there was lack of relationship between leptin and postpartum luteal activity, higher leptin concentrations were associated with shorter intervals to first observed estrus.

Although the physiological mechanisms that the dairy cow undergoes to adapt to lactation requirements should be basically similar among different productive systems, the energy demands due to grazing may modify the important transformations that take place during this period. Moreover, dry matter (DM) intake in these production systems is usually lower than in confined systems and may be insufficient to sustain the high milk production that can be achieved with the genetic potential (Kolver & Muller 1998). Thus, the objective of this study was to characterize insulin, IGF-I, thyroid hormones and leptin concentrations of the transition cow along with BHB and NEFA profiles under grazing conditions. In addition, the effect of parity and BCS at parturition on endocrine and metabolite patterns and reproductive parameters were determined.

Materials and Methods

Animals

Holstein cows with two to five parturitions (multiparous cows, \( n = 21 \)) and cows without previous parturitions (primiparous cows, \( n = 21 \)) (average 305-day milk yield, 6000 and 4800 kg respectively) with normal parturitions during autumn (April) were selected from the herd of the experimental dairy farm of the Agronomy College (Paysandú, Uruguay). Animal experimentation was performed in compliance with regulations set by the Veterinary Faculty, University of Uruguay, Uruguay. Animals were grazing on improved pastures (mixture of grasses and legumes) and 3 weeks before parturition they were offered a diet that consisted of a mixture of 12 kg corn silage and 4 kg commercial concentrate (14% crude protein (CP), 1.7 MCal net energy of location (NEL/kg) which were given once a day, and bales of Setaria italica ad libitum. After parturition, cows had access to a daily strip of pasture (mixture of grasses and legumes), 15 kg (fresh basis) corn silage (33% DM, 6.8% CP) and 6 kg DM of a commercial concentrate (17% CP, 1.7 MCal net energy of lactation (NEL/kg). The pasture sward mass (1650 ± 230 kg DM) was estimated with a comparative yield method adapted from Haydock & Shaw (1975), and an allowance of 15–18 kg DM/cow per day was offered through weekly adjustments of the daily strip size. The cows had access to the grazing plot between the morning and the afternoon milking. The corn silage was fed in the afternoon (after milking), and the concentrate equally distributed during milking time (twice a day). Cows were milked twice a day and milk production was measured every 15 days. BCS was determined every 15 days by the same person from 2 months before until the third month after parturition using a scale from 1 (emaciated) to 5 (fat) according to Edmonson et al. (1989). Cows were scored to the nearest quarter point. At the same time, BW was determined. The BCS at parturition was determined by using the BCS closest to parturition. Animals were classified according to BCS at parturition in lean cows (< 3, \( n = 20 \)) or fat cows (≥ 3, \( n = 22 \)); BCS was not induced by dietary treatment. Estrus was checked twice a day, and animals were inseminated 12 h after heat detection (voluntary waiting period = 50 days). Reproduction was seasonal with a breeding period of 5 months (from May to September). Pregnancy diagnosis was performed by rectal palpation 45 days after artificial insemination. Blood samples were obtained three times a week, from around 30 days prepur- tum to 60 days postpartum. Blood was collected from the jugular vein in heparinized vials approximately 1 h after the morning milking (e.g. 1 h after the morning concent- trate), centrifuged and plasma was stored at −20°C until assayed. The reinitiation of ovarian cyclicity was determined by progesterone concentrations in plasma three times per week. Reproductive parameters measured were


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days to first ovulation, and intervals from parturition to first service and to conception.

**Metabolite and hormone determination**

BHB and NEFA were determined every 10 days from 1 month before to 2 months after parturition by a d-3-hydroxybutyrate kit and a NEFA kit (Randox Laboratories Ltd, Crumlin, Co. Antrim, UK) respectively in the laboratory of the Research Institute for Animal Husbandry and Nutrition, Herceghalom, Hungary. The intra-assay coefficients of variation (CV) for BHB and NEFA were ≤5.5% and ≤ 4.5% and the interassay CV values were ≤7.3% and ≤ 9.7% respectively.

Total plasmatic proteins, albumin, urea, cholesterol, calcium, magnesium, phosphorus and aspartate aminotransferase were determined by commercial kits from Weiner Laboratory (Rosario, Argentina) on a Vitalab Spectra 2 autoanalyzer (Vital Scientific NV, Dieren, The Netherlands). All samples were assayed in two assays at the laboratory Miguel C Rubino, Dirección Laboratorios Veterinarios, Pando, Uruguay. The intra-assay CV was ≤ 3.7% for all the parameters, and the interassay CV was ≤ 9.6%. The data for these metabolites, although used in some statistical analyses, are not shown.

Progesterone was determined three times per week in all plasma samples from parturition (day 0) until around 70 days postpartum (range 58–84 days). Concentrations of all other hormones were determined approximately every 10 days from 1 month before to 2 months after parturition. Progesterone was determined in the Biochemistry Laboratory, Faculty of Veterinary Medicine, Uruguay. All other hormone determinations were assayed in the Endocrine Laboratory of the Faculty of Veterinary Science, Budapest, Hungary.

Progesterone was determined with a commercial kit (Coat-a-count; DPC Diagnostic Products Co., Los Angeles, CA, USA). The intra- and interassay CV values were 6% and 11%. The sensitivity was 0.1 nmol/l.

Thyroxine (T4) and 3,3′,5-tri-iodothyronine (T3) were determined by 125I-Spec RIA coated tube kit (Institute of Isotopes Co. Ltd, Budapest, Hungary). The sensitivity was 0.5 nmol/l (T4) and 0.19 nmol/l (T3). The intra-assay CV values were 6.4–8.1% for T4 and 6.0–8.3% for T3. The interassay CV values were ≤5.8% and ≤ 6.5% respectively.

Insulin was determined by a 125I-insulin RIA CT kit (CIS Bio International Ltd, Gif-Sur-Yvette, France). The sensitivity of the assay was 1.08 μU/ml. The intra-assay CV ranked 5.5–8.4%, while the interassay CV was ≤ 8.8%.

IGF-I was determined in neutralized acid–ethanol extracts of plasma using a 125I-IGF RIA validated for bovine samples (Nikolić et al. 2001). Since the amino acid composition of bovine and human IGF-I is the same, human IGF-I was used as the working standard in the presence of human IGF-II (4 ng/tube). Human IGF-I (ICN Biomedicals Inc., Aurora, OH, USA) labeled with 125I was used as the tracer and polyclonal rabbit antibodies to human IGF-I (Biogenesis, Poole, Dorset, UK) as the reagent. The intra-assay CV values for duplicate samples were routinely from 3% to 6%. Interassay CV values were below 12%.

Plasma leptin concentration was quantified in accordance with our earlier report in ewes (Kulcsar et al. 2004). This assay system is a local version of the ovine-specific homologue 125I RIA of Delavaud et al. (2000) which was validated for bovine species (Delavaud et al. 2002). Under routine use, the sensitivity of this assay for bovine plasma is 0.03 nmol/l. The inter- and intra-assay CV values were 12.0, 5.5 and 6.0%, and 10, 4.5 and 5.2% for the low, medium and high quality control samples respectively.

**Statistical analyses**

Milk production, BCS, metabolites and hormonal concentrations were analyzed by a mixed procedure (Statistical Analysis System; SAS Institute Inc., Cary, NC, USA). The statistical model included the effects of parity (categories: primiparous = L1 or multiparous = L2), BCS at parturition (BCS <3 or ≥3), days including pre- and postpartum periods (linear and quadratic functions) and interactions. The covariance structure was autoregressive order 1 and data are presented in the Figures as least square means ± pooled standard error. To study reproductive parameters, a general linear model was used and the fixed effects were parity and BCS at parturition. Tukey–Kramer tests were conducted to analyze differences between groups. The reinitiation of ovarian cyclicity was defined as the day when progesterone increased from basal concentrations in two consecutive samples of >1.6 nmol/l or one sample of >3.2 nmol/l. If no detectable progesterone was observed until around day 70 postpartum, the last day of sampling was arbitrarily considered as the initiation of ovarian cyclicity for that cow. Correlation coefficients were calculated to study relationships between variables (correlation procedure of SAS). Factors affecting the reinitiation of ovarian cyclicity were evaluated by regression analysis using a backwards elimination procedure (initial inclusion of all independent variables and sequential elimination of the variables with $P > 0.10$ using the regression procedure of SAS) to determine those with $P$ values <0.10. The dependent variable was the reinitiation of ovarian cyclicity and the independent variables included were parity, BCS at parturition, BW, body condition, milk production, total protein, albumin, urea, NEFA, BHB, cholesterol, aspartate aminotransferase, calcium, phosphorus, magnesium, insulin, T3, T4, IGF-I and leptin. The last observation before the reinitiation of ovarian cyclicity in each cow was included in this study. Regression
analyses were performed to study the relationships between leptin and BCS, leptin and NEFA and leptin and IGF-I before and after parturition in cows with low and high BCS at parturition.

Results

Milk production

Primiparous cows produced less milk than multiparous cows during the experimental period (Tukey–Kramer, $P < 0.001$, Fig. 1). Milk production was affected by days postpartum (Table 1). Milk production was negatively correlated with body condition and with NEFA ($r = -0.35$, $n = 165$, $P < 0.05$ and $r = -0.24$, $n = 165$, $P < 0.05$ respectively).

BW and BCS

The loss of body condition is shown in Fig. 2. Lean cows (BCS at parturition $<3.0$) had a smaller BCS during the experimental period, while fat cows ($\geq 3.0$) tended to lose more BCS. BCS was affected by parity and days postpartum, with an interaction between both effects (Table 1). Primiparous cows had a steeper decline in BCS than multiparous cows but they recuperated faster (Table 2). BCS was negatively correlated with NEFA and BHB ($r = -0.35$, $n = 298$, $P < 0.05$ and $r = -0.17$, $n = 295$, $P < 0.05$ respectively).

Multiparous cows had a higher BW than primiparous cows (576 $\pm$ 7 vs 510 $\pm$ 7 kg, $P = 0.0001$). Parity and BCS at parturition affected BW changes during the experimental period. Statistical differences were found on the drop in BW (kg/day: $-1.7$, primiparous vs $-1.3$, multiparous) but not on its recuperation (Table 2). A strong correlation between BW and BCS was found for primiparous cows ($r = 0.7624$, $n = 146$, $P < 0.0001$) and multiparous cows ($r = 0.7416$, $n = 145$, $P < 0.0001$).

NEFA and BHB profiles

NEFA concentrations started to increase before parturition; in primiparous cows they reached peak concentrations at day 20 and in multiparous cows at day 14 and started to decrease thereafter (Fig. 3). The increase observed in NEFA concentrations was higher for primiparous cows and levels remained high for a longer period (Table 2). BHB concentrations were low at parturition, rose sharply to approximately day 10 and slowly decreased thereafter (Fig. 3). When BHB values of $>1$ mmol/l were considered (cows with subnormal levels according to Whitaker et al. (1999)) primiparous cows had more samples with these levels ($P < 0.05$). NEFA and BHB levels were highly correlated ($r = 0.53$, $n = 441$, $P < 0.001$).

Insulin

Insulin concentrations differed according to days postpartum: levels started to decrease before parturition, minimum concentrations were found around parturition but levels were fully recovered at day 30 postpartum (Fig. 4A). There was no effect of parity or BCS at parturition on insulin concentrations (Table 1).

Table 1 F-tests of fixed effects included in the model for measured parameters in cows under grazing conditions. Fixed effects are the effects of parity (P), BCS at parturition (BCS-C), days postpartum (linear (DPP) and quadratic functions (DPP2)) and interactions between them.

<table>
<thead>
<tr>
<th>Variable</th>
<th>$n$</th>
<th>$P$</th>
<th>BCS-C</th>
<th>DPP</th>
<th>DPP2</th>
<th>$P \times$ BCS-C</th>
<th>DPP $\times$ P</th>
<th>DPP $\times$ BCS-C</th>
<th>DPP2 $\times$ P</th>
<th>DPP2 $\times$ BCS-C</th>
</tr>
</thead>
<tbody>
<tr>
<td>BCS</td>
<td>328</td>
<td>0.07</td>
<td>***</td>
<td>***</td>
<td>***</td>
<td>0.11</td>
<td>***</td>
<td>0.11</td>
<td>0.08</td>
<td></td>
</tr>
<tr>
<td>BW</td>
<td>292</td>
<td>***</td>
<td></td>
<td></td>
<td>**</td>
<td>***</td>
<td>***</td>
<td>***</td>
<td>0.10</td>
<td></td>
</tr>
<tr>
<td>NEFA</td>
<td>446</td>
<td>0.11</td>
<td>***</td>
<td>***</td>
<td>***</td>
<td>***</td>
<td>***</td>
<td>***</td>
<td>***</td>
<td></td>
</tr>
<tr>
<td>BHB</td>
<td>441</td>
<td></td>
<td>***</td>
<td>***</td>
<td>***</td>
<td>0.07</td>
<td></td>
<td>0.08</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Insulin</td>
<td>446</td>
<td></td>
<td></td>
<td>***</td>
<td>***</td>
<td>0.09</td>
<td></td>
<td>***</td>
<td>***</td>
<td>0.09</td>
</tr>
<tr>
<td>T4</td>
<td>446</td>
<td>0.10</td>
<td>***</td>
<td>***</td>
<td>***</td>
<td>***</td>
<td></td>
<td>***</td>
<td>***</td>
<td>0.09</td>
</tr>
<tr>
<td>IGF-I</td>
<td>446</td>
<td>***</td>
<td></td>
<td>***</td>
<td>***</td>
<td>***</td>
<td>***</td>
<td>***</td>
<td>***</td>
<td>0.09</td>
</tr>
<tr>
<td>Leptin</td>
<td>446</td>
<td>0.13</td>
<td>***</td>
<td>***</td>
<td>***</td>
<td>0.07</td>
<td>0.07</td>
<td>***</td>
<td>***</td>
<td>0.09</td>
</tr>
</tbody>
</table>

* $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$. 

Figure 1 Milk production (means $\pm$ S.E.M.) of primiparous (lactation 1, L1, $n = 21$) and multiparous (lactation 2, L2, $n = 21$) cows under grazing conditions.
Thyroid hormones

Both hormones were affected by days postpartum (Table 1) but no other effect was found and parallel curves could be observed in both categories (Fig. 4B). Concentrations started to decrease before parturition and minimum levels were found soon after parturition. Thyroid hormones did not recover prepartum concentrations until the end of the study. Multiparous cows had higher T3 concentrations than primiparous cows (1.36 ± 0.03 vs 1.23 ± 0.03 nmol/l), and T4 concentrations tended to be different (43 ± 1.8 vs 39.4 ± 1.7 nmol/l, *P* < 0.1).

IGF-I

Effects of parity and BCS at parturition were found in IGF-I concentrations; primiparous cows and lean cows had lower concentrations of IGF-I (Fig. 5A and B). Concentrations of IGF-I started to decrease 20 days before parturition in all cows to reach half of the prepartum values after parturition. The curves of IGF-I concentration differed according to category (Table 2); primiparous cows had a steeper decrease than multiparous cows, remained

Table 2 Estimates of the functions in primiparous and multiparous cows. Only variables with significant differences according to parity are presented.

<table>
<thead>
<tr>
<th>Variable (unit)</th>
<th>Primiparous cows</th>
<th>Multiparous cows</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Intercept</td>
<td>DPP</td>
</tr>
<tr>
<td>BCS (scale 1–5)</td>
<td>2.61&lt;sup&gt;a&lt;/sup&gt;</td>
<td>-0.01861&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>BW (kg)</td>
<td>521&lt;sup&gt;a&lt;/sup&gt;</td>
<td>1.6933&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>NEFA (mmol/l)</td>
<td>0.42</td>
<td>0.00394&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>IGF-I (nmol/l)</td>
<td>3.81&lt;sup&gt;a&lt;/sup&gt;</td>
<td>-0.0394&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>Leptin (nmol/l)</td>
<td>0.40&lt;sup&gt;a&lt;/sup&gt;</td>
<td>-0.00312&lt;sup&gt;x&lt;/sup&gt;</td>
</tr>
</tbody>
</table>

Values with different superscripts within estimates (intercept, DPP and DPP<sup>2</sup>) differ according to parity (*P* < 0.05). x vs y differ (*P* = 0.066).
and (B) L1-T3 (L2, n = 21) cows under grazing conditions. (A) L1-Ins (■), L2-Ins (□) and (B) L1-T3 (■), L2-T3 (□), L1-T4 (●), L2-T4 (○).

Leptin

Leptin levels decreased sharply before parturition and – in contrast with the pattern of the other hormones – concentrations remained low during the sampling period (Fig. 5C and D). BCS at parturition affected leptin concentrations (Table 1). Leptin concentrations (nmol/l) in fat cows fell more sharply than in lean cows (4.36 ± 0.039x + 0.00078x² vs 3.96 ± 0.026x + 0.00065x², P < 0.05; a vs b, P < 0.05; y vs z, P = 0.094).

Correlations among variables

T4, T3, IGF-I and leptin were correlated with BCS; the latter presented the highest correlation coefficient (r = 0.51, r = 0.68, r = 0.61 and r = 0.8 respectively, n = 292, P < 0.001 for all). Milk production was associated with T3 and with a higher significance to IGF-I (r = 0.22, n = 152, P < 0.05 and r = 0.26, n = 152, P < 0.01 respectively). All hormones were negatively correlated with NEFA and BHB and high correlation coefficients for NEFA vs T3 (r = −0.67, n = 446, P < 0.001) and NEFA vs IGF-I (r = −0.7, n = 446, P < 0.001) were found. All hormone concentrations were positively correlated and T3 vs T4 (r = 0.83, n = 446, P < 0.001), T3 vs IGF-I (r = 0.78, n = 446, P < 0.001) and IGF-I vs leptin (r = 0.7, n = 446, P < 0.001) were the ones most closely related.

Regression analyses

The regression analyses between plasma leptin and BCS before and after parturition in lean and fat cows are shown in Fig. 6. While leptin concentrations were consistently related to BCS during the experimental period in fat cows this was present only before parturition in lean cows.

Plasma leptin concentration (nmol/l) was less related to NEFA concentration in fat than in lean cows, either before or after parturition (fat cows: y = 0.669 − 0.431x, n = 91, R² = 0.14 or y = 0.444 − 0.125x, n = 156, R² = 0.05; lean cows: y = 0.612 − 0.538x, n = 70, R² = 0.32 or y = 0.444 − 0.194x, n = 129, R² = 0.21 respectively, P < 0.005 for all).

Similar observations occurred when analyzing leptin vs IGF-I concentration. The functions for fat cows were y = 0.325 + 0.005x, n = 91, R² = 0.19 before parturition and y = 0.25 + 0.0038x, n = 156, R² = 0.2 after parturition; lean cows y = 0.125 + 0.0088x, n = 70, R² = 0.51 and y = 0.238 + 0.0044x, n = 129, R² = 0.33, P < 0.0001 for all).

Reproductive parameters

There was an effect of parity on the initiation of ovarian cyclicity, parturition to first service and parturition to conception intervals. The interaction effect of BCS at parturition and parity was significant only for the reinitiation of ovarian cyclicity (Table 3). Postpartum anestrus was longer in primiparous than in multiparous cows: 45 vs 21 days (P < 0.0001). Primiparous lean cows presented a longer interval from parturition to first ovulation than primiparous fat cows, but this was not observed for multiparous cows.
Overall, the interval from parturition to first service was 131 and 97 days for primiparous and multiparous cows respectively \((P, 0.01)\). The interval from parturition to conception was 146 and 109 days for primiparous and multiparous cows \((P, 0.01)\). An interesting finding was that multiparous lean cows reinitiated ovarian cyclicity before the primiparous fat cows. This was reflected in the parturition to conception interval (Table 3).

**Table 3** Mean ± S.E.M. days to first ovulation, parturition first service interval and parturition conception interval in primiparous (L1) or multiparous (L2) cows with BCS at parturition of \(<3\) or \(\geq 3\). Numbers are shown in parentheses.

<table>
<thead>
<tr>
<th>Parity/BCS at parturition</th>
<th>First ovulation</th>
<th>Parturition first service interval</th>
<th>Parturition conception interval</th>
</tr>
</thead>
<tbody>
<tr>
<td>L1/BCS &lt; 3 (12)</td>
<td>52.8 ± 4.8 (12)</td>
<td>139.0 ± 11.8 (10)</td>
<td>143.0 ± 13.0 (8)</td>
</tr>
<tr>
<td>L1/BCS ≥ 3 (9)</td>
<td>37.4 ± 5.6 (9)</td>
<td>122.2 ± 12.8 (9)</td>
<td>149.4 ± 12.2 (8)</td>
</tr>
<tr>
<td>L2 BCS &lt; 3 (8)</td>
<td>19.0 ± 6.3 (8)</td>
<td>99.9 ± 13.7 (8)</td>
<td>114.1 ± 13.0 (8)</td>
</tr>
<tr>
<td>L2/BCS ≥ 3 (13)</td>
<td>23.0 ± 4.5 (13)</td>
<td>93.4 ± 10.0 (11)</td>
<td>104.0 ± 10.0 (11)</td>
</tr>
</tbody>
</table>

Values with different letters within the same column differ \((P < 0.05)\) except e vs f \((P = 0.085)\) and g vs h \((P = 0.058)\).

Overall, the interval from parturition to first service was 131 and 97 days for primiparous and multiparous cows respectively \((P < 0.01)\). The interval from parturition to conception was 146 and 109 days for primiparous and multiparous cows \((P < 0.01)\). An interesting finding was that multiparous lean cows reinitiated ovarian cyclicity before the primiparous fat cows. This was reflected in the parturition to conception interval (Table 3).

BCS at parturition, BCS, insulin, leptin, BHB and total protein (all with \(P < 0.05\)) and urea and T3 \((P < 0.08)\) were associated with the reinitiation of ovarian cyclicity as determined by regression analysis \((R^2 = 0.78, P = 0.0001)\).

**Discussion**

This study has demonstrated that parity affects NEFA, T3, T4 and IGF-I concentrations as well as reproductive parameters in dairy cows, while BCS at parturition has an effect on IGF-I and leptin concentrations and reproductive parameters.

Body condition decreased from 30 days before parturition, and this trend was steeper during the first 4 weeks after parturition. NEFA and BHB levels increased around parturition, reflecting the negative EB of the animals (Ingvarsten & Andersen 2000). Primiparous cows had lower BCS than multiparous cows and this was consistent
with the higher NEFA levels and the greater number of BHB samples above 1 mmol/l in this category (subclinical ketosis according to Whitaker et al. (1999)). This is probably related to the increased needs for growth in primiparous cows occurring simultaneously with the demands of lactation and their lower feed intake capacity as described previously (Rémond et al. 1991).

Studies investigating the potential metabolic signals for the reproductive axis have been focused primarily on blood metabolites and metabolic hormones known to fluctuate during altered states of energy metabolism. Decreased insulin concentrations around parturition were found in agreement with previous reports (Holtenius et al. 2003). Insulin plays a central role in the homeostatic control of energy metabolism and its concentration is positively correlated with energy intake (Chilliard et al. 1998). The diminished concentration is consistent with the reduction in DM intake that characterizes this period (Bertics et al. 1992). Both insulin and IGF-I are known to stimulate follicular growth (Spicer et al. 1993), but in this study insulin did not seem to be related to reinitation of ovarian cyclicity: insulin concentrations were fully recovered 30 days postpartum and while multiparous cows ovulated before that period, primiparous cows ovulated afterwards.

The peripheral tissues try to fit their current local energy metabolism to the postpartum catabolic condition, for example by interfering in the path of thyroid hormones, resulting in diminished circulation as observed here as well by others (Pethes et al. 1985, Heyden et al. 1993). Lower concentrations of T3 were found in primiparous cows which is in disagreement with the data of Cissé et al. (1991) who suggested that higher T3 concentrations in the first lactation could be due to a lower milk yield since thyroid hormones are excreted by the mammary gland. On the other hand, a lower level of T3 was reported to be associated with lower EB (Blum et al. 1983), which seems to be the case in our study when taking into account the fact that primiparous cows presented comparatively greater body condition loss and an overall unbalanced metabolic condition. This could explain the apparent discrepancy with the study of Cissé et al. (1991), in which heifers and cows had the same EBs. Low thyroid hormone concentrations have been suggested to be associated with low reproductive performance in the postpartum cow (Huszenicza et al. 2002).

In our study, parity affected IGF-I concentrations as primiparous cows had lower concentrations of this hormone. This contrasts with previous findings (Wathes et al. 2003) where IGF-I concentrations were higher in

Figure 6 Relationships between plasma leptin and BCS in lean or fat dairy cows (i.e. with BCS at parturition of BCS < 3 or BCS ≥ 3 respectively) measured either 1 month before or 2 months after parturition.
the younger animals. Since it has been suggested that low insulin and IGF-I are the metabolic signals that delay ovulation (Beam & Butler 1999) and IGF-I concentrations were high in primiparous cows, Taylor et al. (2003) suggested that in this category – which is still growing – insulin concentrations may be limiting, whereas in older cows a closer association is observed between IGF-I and fertility parameters. Our results do not support this hypothesis since primiparous cows presented lower IGF-I but similar insulin concentrations and presented a delayed ovulation when compared with multiparous cows. Primiparous cows with lower mean IGF-I concentrations presented a steeper decline in IGF-I than multiparous cows. Lean cows with lower IGF-I concentrations than fat cows presented a slower decline. IGF-I concentrations are good indicators of the capacity to resume cyclicity after parturition in agreement with Roberts et al. (1997): fat cows and multiparous cows had higher IGF-I concentrations and better reproductive performance.

As previously reported in adult dry non-pregnant ruminants (Delavaud et al. 2000, 2002), plasma leptin content in this study was a good indicator of body fatness (BCS) in peripartum dairy cows, especially when measured 3 weeks before calving, i.e. at a stage where leptin is highly expressed. On the other hand, Holtenius et al. (2003) reported no relation between leptin concentrations and BCS after parturition. Differences among studies could be due to the different production system, energy intake, EB, range of BCS among cows, genetic background and/or frequency of sampling. Leptin concentrations started to decrease by 20 days before parturition in agreement with other studies (Kadokawa et al. 2000, Block et al. 2001, Liefers et al. 2003) and then remained low until 2 months after parturition which is at variance with the reported increase around day 10 postpartum (Kadokawa et al. 2000), the progressive increase from week 1 to week 12 (Reist et al. 2003), the unchanged concentrations up to week 5 (Huszenicza et al. 2001) and the transient increase at week 2 (Liefers et al. 2003), but consistent with the lower concentrations found by Block et al. (2001) and Holtenius et al. (2003). Baseline leptin levels were reached later postpartum in fat cows consistent with a later maximal loss in BCS in these animals, suggesting that the decrease in leptin concentrations due to negative EB may be masked in fat cows by the amount secreted by the adipose tissue reserves that are still high during the first month of lactation. This hypothesis is supported by the regression analyses of plasma leptin during the pre- and postpartum period in lean and fat cows: while no association was found between leptin levels and BCS after parturition in lean cows, this was maintained in fat cows during the same period. Overall, our results suggest interactions between the effects of BCS, parity and lactation stage on peripartum leptin regulation.

The reinitiation of ovarian cyclicity was delayed in primiparous cows and in lean animals and this was consistent with longer intervals from parturition to first service and to conception in these animals. The anestrous duration was associated with BCS loss and was longer in primiparous cows as shown previously (Butler & Smith 1989). These authors and others (Huszeniczca et al. 1987, 1988) demonstrated that the sooner the cows restore the EB, the sooner they will start cycling and become pregnant. It is interesting to note that multiparous lean cows resumed cyclic activity sooner than primiparous fat cows and this could be due to the patterns of the endocrine signals or to negative EB due to lower intake, ascendant lactation curve (Fig. 1) and/or growth requirements in heifers. All these parameters could indeed be related to the high body lipid mobilization (Verité & Chilliard 1992), BCS loss (Rémond et al. 1991) and higher plasma NEFA (Cissé et al. 1991, Fig. 3) that were observed in primiparous cows vs multiparous cows, despite the fact that the former yielded less milk. In this study, parity was a more important effect for reproductive performance than BCS at parturition; it should be noted that these animals were under grazing conditions where an effect such as dominance for food availability is present (Grant & Albright 2001).

Cows with better reproductive performance had higher IGF-I and leptin concentrations. Similarly, a negative relationship between IGF-I concentrations after parturition and the interval to the resumption of ovarian cyclicity has been reported (Butler 2000). Data regarding leptin and reproductive performance are conflicting. Cows with decreased leptin have been linked to delayed onset of cyclicity or longer intervals to first estrus during the postpartum period (Kadokawa et al. 2000, Liefers et al. 2003). Similarly, a slightly increasing tendency of the leptin pattern was seen in cows resuming their ovarian cyclicity within 35 days (Huszeniczca et al. 2001). The data presented here do not support the theory that the restoration in leptin concentrations activates the hypothalamic–pituitary–ovarian axis since, unlike the other hormones, it remained low until 60 days postpartum. On the other hand, not only are the hormone concentration itself and the tissue sensitivity to it (receptors) important for the reproductive axis, but also the hormonal dynamics (e.g. leptin decay – as for IGF-I – was more precipitous in primiparous cows) could be read by the endocrine system as a different signal. As previously reported (Bocquier et al. 1998, Clarke & Henry 1999) our results also suggest that leptin may play a permissive role, when increased above a critical threshold, in the activation of the hypothalamic–pituitary axis and consequent reinitation of ovarian activity in the postpartum cow. This would allow cows which are in good condition at parturition and thus have a higher leptinemia during early lactation (Fig. 5), to have a facilitated reproductive activity.

In summary, primiparous cows showed a metabolic/endocrine profile more unbalanced than that of multiparous cows, reflecting that they are recovering from the negative EB period with more difficulty. The endocrine signals that
could most likely inform the reproductive axis with regard to the negative EB and/or the level of body reserves that may explain the reproductive performance found in this study were IGF-I and leptin.

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