Focus on Meiosis

Stops and starts in mammalian oocytes: recent advances in understanding the regulation of meiotic arrest and oocyte maturation

Lisa M Mehlmann

Department of Cell Biology, University of Connecticut Health Center, 263 Farmington Ave., Farmington, Connecticut 06032, USA

Correspondence should be addressed to L M Mehlmann; Email: lmehlman@neuron.uchc.edu

Abstract

Mammalian oocytes grow and undergo meiosis within ovarian follicles. Oocytes are arrested at the first meiotic prophase, held in meiotic arrest by the surrounding follicle cells until a surge of LH from the pituitary stimulates the immature oocyte to resume meiosis. Meiotic arrest depends on a high level of cAMP within the oocyte. This cAMP is generated by the oocyte, through the stimulation of the Gs G-protein by the G-protein-coupled receptor, GPR3. Stimulation of meiotic maturation by LH occurs via its action on the surrounding somatic cells rather than on the oocyte itself. LH induces the expression of epidermal growth factor-like proteins in the mural granulosa cells that act on the cumulus cells to trigger oocyte maturation. The signaling pathway between the cumulus cells and the oocyte, however, remains unknown. This review focuses on recent studies highlighting the importance of the oocyte in producing cAMP to maintain arrest, and discusses possible targets at the level of the oocyte on which LH could act to stimulate meiotic resumption.

Introduction

Meiosis is the process by which diploid germ cells – oogonia or spermatogonia – reduce their number of chromosomes in half in preparation for combining with a haploid cell of the opposite sex to create a genetically new, diploid individual. In female mammals, meiosis occurs over a prolonged period of time; oogonia enter meiosis but become arrested at the diplotene stage of the first prophase (Eppig et al. 2004). Meiosis resumes in response to a surge of luteinizing hormone (LH) from the pituitary gland during the estrous or menstrual cycle, shortly before ovulation. The process by which the oocyte completes the first meiotic division and undergoes other cytoplasmic changes, and progresses to metaphase II is called oocyte maturation. Because the mature, fertilizable oocyte has a relatively short lifespan in the female reproductive tract, the timing of oocyte meiotic arrest, as well as maturation, must be tightly regulated. This review will highlight recent studies examining how the oocyte maintains arrest, and will discuss potential mechanisms whereby LH acts to stimulate meiotic resumption.

The functional unit within the ovary is the follicle, which is comprised of one or more layers of granulosa cells surrounding the oocyte (Fig. 1) (Gougeon 1996, Zeleznik 2004). Ovarian follicles form during embryonic development (Gougeon 1996, Eppig et al. 2004). During follicular growth, the somatic cells divide to form several layers, the oocyte enlarges, and a fluid-filled antrum begins to form. Some follicles at the early antral stage are ‘recruited’ to continue growing; this growth is dependent on the pituitary gonadotropin, follicle-stimulating hormone (FSH) (Gougeon 1996, Zeleznik 2004). During this phase, the antrum divides the granulosa cells into two separate compartments: mural granulosa cells form the outer layers, while the cumulus cells surround the oocyte. The oocyte grows to its full size (~75 μm diameter in the mouse, ~100 μm in the human), but remains arrested in prophase I. If an oocyte is removed from an antral follicle, it spontaneously resumes meiosis and progresses to second metaphase (Pincus & Enzmann 1935). This indicates that the follicle cells hold the oocyte in prophase arrest. Recent progress in clarifying the nature of this arrest is discussed below.
Meiosis resumes in response to a preovulatory surge of LH. LH receptors are located on the mural granulosa cells but not on the cumulus cells or the oocyte (Peng et al. 1991, Eppig et al. 1997), so the mechanisms by which LH stimulates oocyte maturation is indirect. Although much is known about LH signaling in the mural granulosa cells (Richards et al. 2002), how the LH signal is passed on to the oocyte is incompletely understood. The mechanism by which LH triggers oocyte maturation is currently being studied, and hypotheses of how LH causes the oocyte to resume meiosis are discussed below. Ultimately, LH action on the mural granulosa cells translates to a change in signaling molecules within the oocyte to initiate meiotic resumption.

Because the entire follicle surrounding the oocyte must remain intact in order to preserve its normal function, the oocyte has been largely inaccessible to biochemical studies of its function in a physiological environment. The mechanisms that maintain meiotic arrest of the oocyte, as well as the mechanisms by which LH triggers resumption of meiosis, have therefore been technically difficult to study. Many researchers have utilized oocytes or cumulus–oocyte complexes that have been removed from their follicles and maintained in meiotic arrest artificially. While such experiments have been useful for identifying some of the major components involved in the maintenance of meiotic arrest, they are difficult to interpret in the context of what happens in vivo, and in terms of elucidating the earliest steps in the process of oocyte maturation. Recently, new methods for microinjacting the mammalian oocyte within its follicle have provided a means to directly test hypotheses pertaining to meiotic arrest and resumption (Mehlmann et al. 2002, 2004, Kalinowski et al. 2004). This microinjection method has been used to clarify the important role of the Gs G-protein, as well as the necessity for the Gs-linked receptor, GPR3, in the maintenance of arrest in the mouse oocyte (see below).

**Maintenance of meiotic arrest**

Prior to the midcycle surge of LH, the growing oocyte acquires the ability to undergo oocyte maturation. The acquisition of meiotic competence occurs around the time of antrum formation (Erickson & Sorensen 1974, Sorensen & Wassarman 1976, Mehlmann et al. 2004) and corresponds to a point at which the oocyte achieves a threshold level of maturation-promoting proteins, such as CDK1 (cyclin-dependant kinase) and cyclin (de Vantéry et al. 1996, 1997, Kanatsu-Shinohara et al. 2000). Despite the ability of the fully grown oocyte to mature, it remains arrested in prophase I until the LH surge. It is well established that meiotic arrest is regulated by cAMP levels within the oocyte (Conti et al. 2002, Eppig et al. 2004). Spontaneous maturation of oocytes isolated from their follicles can be prevented by including membrane permeant cAMP analogs or cAMP phosphodiesterase inhibitors, such as hypoxanthine or 3-isobutyl-1-methylxanthine (IBMX), in the culture medium (Cho et al. 1974, Dekel & Beers 1978, Conti et al. 2002). Moreover, cAMP levels decrease in oocytes following removal from their follicles (Törnell et al. 1990), as well as in isolated oocytes after removal of IBMX (Schultz et al. 1983a, Vivarelli et al. 1983). The decrease in oocyte cAMP occurs within 2 h after washing out IBMX, a time during which the oocyte becomes committed to resuming meiosis (Schultz et al. 1983a, Vivarelli et al. 1983).

The downstream pathway(s) by which high cAMP levels prevent meiotic maturation is incompletely understood, and a detailed discussion is beyond the scope of this review (Fig. 2 and see Eppig et al. 2004). Ultimately, the cAMP level within the oocyte affects the activity of the CDK/cyclin B (CYB) protein complex, also known as maturation, meiosis or mitosis promoting factor (MPF). High cAMP levels within the oocyte result in the phosphorylation of CDK1 on Thr14 and Tyr15, rendering it unable to promote cell division. The downstream pathway(s) by which high cAMP levels prevent meiotic maturation is incompletely understood, and a detailed discussion is beyond the scope of this review (Fig. 2 and see Eppig et al. 2004). Ultimately, the cAMP level within the oocyte affects the activity of the CDK/cyclin B (CYB) protein complex, also known as maturation, meiosis or mitosis promoting factor (MPF). High cAMP levels within the oocyte result in the phosphorylation of CDK1 on Thr14 and Tyr15, rendering it unable to promote cell division.
inactive (Duckworth et al. 2002). A decrease in oocyte cAMP early in oocyte maturation leads to the dephosphorylation of CDK1 on Thr14 and Tyr15, and the MPF complex becomes active such that the oocyte can re-enter meiosis. The discrete set of steps through which cAMP activates or inactivates MPF are still under investigation. The major players are protein kinase A (PKA), which, through an undetermined number of steps, regulates the activities of the phosphatase CDC25 and the kinase WEE1/MYT1 (Eppig et al. 2004). CDC25 dephosphorylates CDK1, while WEE1/MYT1 phosphorylates it. Oocytes from mice lacking the Cdc25b gene are unable to activate MPF and cannot undergo meiotic resumption, highlighting the importance of WEE1 or MYT1. Future studies are needed to clarify the entire pathway by which cAMP levels affect the activity of MPF.

cAMP could be produced either by the oocyte or by the follicle cells that surround it. One long-standing hypothesis is that cAMP is produced by follicle cells and diffuses through gap junctions to the oocyte (Anderson & Albertini 1976, Bornslaeger & Schultz 1985, Piontkewitz & Dekel 1993, Webb et al. 2002b). Gap junctions are present between the cumulus cells and the oocyte (Albertini & Anderson 1974, Anderson & Albertini 1976). However, the lack of specific inhibitors against gap junctions in the oocyte has complicated efforts to clarify their possible role in the maintenance of meiotic arrest. For further discussion, see Piontkewitz & Dekel (1993), Webb et al. (2002b) and Eppig et al. (2004).

An alternative hypothesis for how high levels of cAMP are maintained in competent, fully grown oocytes is that the oocyte produces its own cAMP through a G-protein-linked receptor in the oocyte plasma membrane that stimulates Gs and, subsequently, adenylyl cyclase (AC) (Fig. 2). Several lines of evidence support this hypothesis. (1) Mouse oocytes contain all of the components necessary to produce cAMP, including the Gs G-protein (Mehlmann et al. 2002), a Gs-coupled G-protein receptor, GPR3 (see below) (Mehlmann et al. 2004), and AC (Horner et al. 2003). (2) Stimulation of oocyte AC with forskolin raises cAMP levels in isolated rodent oocytes and delays the onset of germinal vesicle breakdown (GVBD) (Olsiewski & Beers 1983, Schultz et al. 1983a, Uner et al. 1983, Bornslaeger & Schultz 1985). (3) Microinjection of the non-hydrolyzable GTP analog, GTPγS, which activates G-proteins including Gs, transiently and dose-dependently maintains meiotic arrest in isolated mouse oocytes (Downs et al. 1992). (4) cAMP levels increase in isolated oocytes maintained in meiotic arrest with the phosphodiesterase inhibitors, IBMX or hypoxanthine (Bornslaeger & Schultz 1985, Webb et al. 2002b). (5) Cholera toxin, which irreversibly activates Gs (De Haan & Hirst 2004), has been shown to delay oocyte maturation in isolated oocytes (Dekel & Beers 1978, Schultz et al. 1983b, Uner et al. 1983, Vivarelli et al. 1983, Downs et al. 1992, Grondahl et al. 2000a). The inability of cholera toxin to completely prevent maturation may be a result of Gs degradation within the oocyte following its activation (Levis & Bourne 1992, Fong & Milligan 1999, Moravcova et al. 2004).

Direct evidence for an essential role of Gs in the maintenance of meiotic arrest has been obtained recently by microinjecting either a function-blocking antibody or a dominant negative form of the α subunit of Gs into follicle-enclosed oocytes (Mehlmann et al. 2002, Kalinowski et al. 2004). This pathway is supported further by the finding that oocytes from mice lacking the AC3 AC isofrom, which is present in the oocyte, spontaneously undergo GVBD within ovarian follicles (Horner et al. 2003). Because Gs activity requires stimulation by a G-protein-coupled receptor, it has been postulated that such a receptor exists in the mouse oocyte membrane. This receptor could exhibit constitutive activity, and/or could be stimulated by a ligand produced by the surrounding follicle cells. Inhibiting Gs in isolated oocytes maintained in meiotic arrest with hypoxanthine stimulates meiotic resumption (Mehlmann et al. 2002, Kalinowski et al.

Figure 2 Cell signaling leading to the maintenance of meiotic arrest. GPR3, activated either constitutively or by an unknown ligand from the follicle cells, activates Gs, which stimulates AC to cause an elevation of cAMP. cAMP activates protein kinase A (PKA), which ultimately causes the cell cycle regulatory complex, CDK1/cyclin B (CYB), to be phosphorylated (P) and thereby inactivated. This results because PKA leads (directly or indirectly) to the phosphorylation of the phosphatase CDC25b (CDC25b-P), which inactivates it. PKA may also stimulate the activity of the WEE1/MYT1 kinase that phosphorylates CDK1 to keep it inactive and therefore prevent meiotic resumption. The activity of the cAMP phosphodiesterase, PDE3A, is thought to be kept low in the immature oocyte, thus preventing the breakdown of cAMP and maintaining high levels of cAMP within the oocyte.
2004), supporting the hypothesis that the receptor in the oocyte has some constitutive activity.

Recently, the Gs-coupled receptor, GPR3, has been identified as an essential regulator of meiotic arrest in the mouse oocyte (Mehlmann et al. 2004). Gpr3 RNA is localized in oocytes, with ~14 times lower expression in the follicle cells. Oocytes from mice lacking the Gpr3 gene undergo spontaneous oocyte maturation within fully grown, intact follicles, independent of an increase in LH. Competence to undergo meiosis develops when an oocyte reaches its full size and when the follicle begins to form an antral space (Sorensen & Wassarman 1976, Mehlmann et al. 2004). Correspondingly, ~40% of the oocytes within smaller, early antral follicles from Gpr3−/− mice also undergo spontaneous oocyte maturation. The ability of oocytes from Gpr3−/− mice to maintain meiotic arrest can be rescued by microinjecting Gpr3 RNA into incompetent Gpr3−/− oocytes within preantral follicles, followed by a 4-day culture period during which an antrum forms, indicating that the presence of Gpr3 is needed specifically in the oocyte rather than in the follicle cells (Mehlmann et al. 2004).

GPR3 is an orphan receptor that exhibits a high degree of constitutive activity when overexpressed in numerous tissue culture cell lines, resulting in a high level of cAMP production (Eggerickx et al. 1995, Uhlenbrock et al. 2002). This indicates that it is coupled to Gs. It is currently not known whether constitutive activity of GPR3 in the oocyte is sufficient to produce the amount of cAMP required to maintain meiotic arrest, or whether the follicle cells surrounding the oocyte produce a ligand that increases the activity of GPR3. Structurally, GPR3 is closely related to the lysophosphatidic acid receptors, siphingosine-1-phosphate (edg) receptors, cannabinoid receptors, and melanocortin receptors (Uhlenbrock et al. 2002, Ignatov et al. 2003, Kostenis 2004a, 2004b). With the exception of the melanocortin receptors, these receptor families are activated by lipids. It is therefore possible that a lipid present in the regions of membrane contact between cumulus cells and oocyte stimulates GPR3.

Another possibility for how the follicle cells might keep the oocyte arrested in prophase I until the LH surge is that they may inhibit oocyte phosphodiesterase(s) (Conti et al. 2002). Both of these possibilities need to be explored further to determine how the follicle cells interact with the oocyte to keep cAMP levels high prior to the LH surge.

How does LH trigger meiotic resumption?

The mechanism(s) by which LH, acting on the granulosa cells, triggers the oocyte to resume meiosis is still unknown. As mentioned previously, LH acts on the outermost, mural granulosa cells of the follicle; cumulus cells and oocytes lack LH receptors (Peng et al. 1991, Eppig et al. 1997). The LH signal must therefore be transmitted from the mural granulosa cells to the oocyte. The action of LH could either remove an inhibitory, or maturation-arresting, substance or it could provide a positive, maturation-promoting substance to the oocyte (see Conti et al. 2002, Eppig et al. 2004). This review focuses on some of the current ideas in this field, taking into account recent data elucidating the maintenance of meiotic arrest.

Recent work has shed some light on how the LH signal transmits from the exterior to the interior of the follicle. Mural granulosa cells express RNA encoding epidermal growth factor (EGF)-like proteins within 1–3 h after LH receptor stimulation (Park et al. 2004, Ashkenazi et al. 2005), and these proteins, in particular amphiregulin and epiregulin, cause follicle-enclosed oocytes to mature as effectively as LH, though with a faster time-course. They do not cause maturation of isolated oocytes. Pharmacological inhibition of the EGF receptor in cultured follicles completely inhibits LH-induced oocyte maturation, further supporting a link between these EGF-like proteins and LH (Park et al. 2004). These results are in agreement with previous studies showing that EGF promotes meiotic maturation of cumulus-enclosed oocytes (Das et al. 1991, De La Fuente et al. 1999, Coticchio et al. 2004). The signaling pathway between cumulus cells and oocytes remains unknown however.

LH acting on follicle cells surrounding frog and fish oocytes has long been known to stimulate the production of steroid hormones that trigger oocyte maturation (Masui & Clarke 1979, Nagahama et al. 1995, Maller 1998, Thomas et al. 2002, Hammes 2004, Tsafiri et al. 2005). However, steroids have little if any stimulatory effect on mammalian oocyte maturation (Dekel & Beers 1978, Schultz et al. 1983b, Andersen & Byskov 2002, Gill et al. 2004), and in some cases have a slight inhibitory effect (Schultz et al. 1983b, Kaji et al. 1987). Moreover, complete inhibition of steroidogenesis in cultured follicles does not prevent oocyte maturation in response to LH (Lieberman et al. 1976).

The sterol, follicular fluid–meiosis-activating sterol (4,4-dimethyl-5α-cholesta-8,14,24-trien-3β-ol; FF-MAS), is a candidate oocyte maturation-inducing substance. FF-MAS, which was first isolated from human follicular fluid (Byskov et al. 1995), is an intermediate in the cholesterol biosynthetic pathway (Schroepfer 1982). In the mouse, FF-MAS levels increase following injection of human chorionic gonadotropin (hCG), which stimulates the LH receptor (Baltsen 2001). Both purified and synthetic FF-MAS stimulate the resumption of meiosis in isolated oocytes of a variety of mammalian species including mouse, rat, and human (Byskov et al. 1995, Grondahl et al. 1998, 2000b, Hegele-Hartung et al. 1999, 2001). However, it is not clear whether FF-MAS becomes detectable in mouse ovaries earlier than 3 h after hCG injection (Baltsen 2001). Because GVBD is observed within 1.5 to 3 h after hCG injection (Schultz et al. 1983a), FF-MAS levels should increase earlier if it is an oocyte maturation inducer. In addition, FF-MAS-induced GVBD in isolated oocytes
maintained in hypoxanthine takes 6–20 h (Hegele-Hartung et al. 1999, Downs et al. 2001). FF-MAS is therefore not a likely candidate for the initiation of oocyte maturation. However, there is evidence that it improves the ability of oocytes to complete meiosis to metaphase II, as well as the ability of fertilized oocytes to develop to the two-cell and blastocyst stages (Hegele-Hartung et al. 1999, Cukurcam et al. 2003, Griffin et al. 2004, Marin Bivens et al. 2004a, 2004b). For further discussion, see Bybskov et al. (2002) and Tsafriri et al. (2002, 2005).

A meiosis-inducing factor could affect targets downstream of cAMP, perhaps by interacting with cell cycle-regulatory proteins. However, because cAMP levels fall early in oocyte maturation (Schultz et al. 1999, Bivens et al. 1999, Cukurcam et al. 2003, Griffin et al. 2004, Marin Bivens et al. 2004a, 2004b). For further discussion, see Bybskov et al. (2002) and Tsafriri et al. (2002, 2005).

(1) GPR3. Constitutive activity of G-protein-coupled receptors can be reduced by inverse agonists (Milligan 2003). Such an inverse agonist turning off GPR3 would lower cAMP in the oocyte. Alternatively, LH stimulation could affect the activity of a ligand that in the unstimulated follicle would activate GPR3, either by inactivating the ligand or by decreasing its synthesis, to ultimately lower the activity of GPR3. GPR3 could also be inactivated by other mechanisms, such as phosphorylation by G-protein receptor kinases (GRKs), which would result in its downregu-

Figure 3 Potential targets through which LH could act to stimulate meiotic resumption. Each target is indicated with a green number. See corresponding numbers in the text for details.

ation (Penn et al. 2000, Lefkowitz 2004). Indeed, analysis of the GPR3 protein sequence shows several serine and threonine residues in the carboxyl terminus, as well as in the intracellular loops, that could be targets of GRKs or other kinases, which could cause receptor desensitization (Saeki et al. 1993, Eggerickx et al. 1995).

(2) G\textsubscript{i}. G-proteins can be inactivated by GTPase-activating proteins, or ‘GAPs’, also known as regulators of G-protein signaling (RGS) proteins (Kehrl & Sinnarajah 2002, Cabrera-Vera et al. 2003). These proteins accelerate the exchange of GTP for GDP on the G-protein \( \alpha \) subunit, thereby turning off the G-protein. Although nothing is known about RGS proteins in oocytes, it is interesting to note that RGS2 can inhibit G\textsubscript{i}-mediated cAMP production (Sinnarajah et al. 2001, Kehrl & Sinnarajah 2002, Roy et al. 2003). Thus, an RGS protein could potentially inhibit cAMP production in the oocyte following LH stimulation.

(3) Gi. A well-characterized pathway for inactivating ACs is by stimulating the G\textsubscript{i}, G-protein subunit, which lowers cAMP (Simonds 1999, Hanoune & Defer 2001). Indeed, the three AC isoforms that have been found to be expressed in mouse and rat oocytes, AC1, AC3, and AC9 (Horner et al. 2003), are all inactivated by G\textsubscript{i} (Hanoune & Defer 2001), and a G\textsubscript{i} pathway is known to be responsible for triggering oocyte maturation in echinoderm oocytes (Shilling et al. 1989, Chiba et al. 1992, Tadenuma et al. 1992, Jaffe et al. 1993, Kalinowski et al. 2003). In mammals, a role for G\textsubscript{i} has not been examined with regard to LH signaling to cause oocyte maturation. It is possible that activation of G\textsubscript{i} within mammalian oocytes, in response to LH, stimulates meiotic maturation. This hypothesis could be tested by examining the effects of specifically inhibiting G\textsubscript{i} within follicle-enclosed oocytes, using pertussis toxin or antibodies, on LH-induced maturation.

(4) Calcium. All three of the AC isoforms found in rodent oocytes (Horner et al. 2003) can be inactivated by Ca\textsuperscript{2+} (Deier et al. 2000, Hanoune & Defer 2001, Wang & Storm 2003). In mouse oocytes, forskolin-stimulated cAMP production is prevented by raising intracellular Ca\textsuperscript{2+} (Horner et al. 2003). This effect is reversed by an inhibitor of Ca\textsuperscript{2+}/calmodulin-dependent kinase II, suggesting that mouse oocyte AC is inhibited by Ca\textsuperscript{2+}. It is possible that Ca\textsuperscript{2+} rises in the oocyte following LH stimulation such that it could inactivate ACs. Under some experimental conditions, Ca\textsuperscript{2+} release can be induced in cumulus cells, resulting in a subsequent increase in Ca\textsuperscript{2+} in oocytes as long as functional gap junctions exist between the oocyte and the cumulus cells (Mattioli et al. 1998, Webb et al. 2002a). Measurements of Ca\textsuperscript{2+} within follicle-enclosed oocytes following LH stimulation should
be able to clarify whether Ca\(^{2+}\) has a role in triggering oocyte maturation, perhaps at the level of turning off AC.

(5) cAMP phosphodiesterase (PDE). An attractive hypothesis is that LH stimulation leads to the activation of oocyte PDE, which hydrolyzes cAMP (Conti et al. 2002). The PDE3A isoform is a prevalent PDE in the mouse oocyte (Tsafirri et al. 1996, Shitsukawa et al. 2001). PDE3A activity in cumulus-enclosed mouse oocytes has been shown to increase following LH receptor stimulation (Richard et al. 2001); however, PDE3A activity has not been examined in isolated oocytes following stimulation of the LH receptor. Microinjection of active PDE into isolated mouse oocytes arrested with the PDE inhibitor IBMX causes GVBD (Bornslaeger et al. 1986). A critical role for PDE3A in mouse oocyte maturation has recently been demonstrated by generating PDE3A-deficient mice by homologous recombination (Masciarelli et al. 2004). The oocytes of female Pde3A\(^{-/-}\) mice are unable to undergo meiotic resumption, remaining arrested at prophase I despite normal follicular growth and ovulation. Likewise, oocytes from Pde3A\(^{-/-}\) mice do not spontaneously mature when released from the ovary into culture medium. The ability of these oocytes to resume meiosis is restored, however, by inhibiting PKA or by microinjecting RNA encoding the phosphatase CDC25 (Masciarelli et al. 2004). Although these studies highlight the importance of PDE for meiotic resumption, experiments in which PDE activity is measured specifically in oocytes within intact follicles before and after LH treatment would provide stronger evidence that PDE is a major target of regulation by LH.

**Concluding remarks**

The mechanisms of mammalian meiotic arrest and resumption have been technically challenging to study because the oocyte is embedded in multiple layers of cells and is therefore difficult to manipulate and observe. The follicle must remain intact in order to preserve its normal function. In addition, there is limited material available for biochemical studies of mammalian oocytes. However, new methods for studying oocyte maturation have recently provided useful information about the mechanisms of meiotic arrest. Recent methods have utilized genetically altered mice, as well as directly inhibiting oocyte-specific proteins by microinjecting follicle-enclosed oocytes. With these methods, the pathway leading to high cAMP levels in meiotically arrested oocytes has been clarified, and the receptor GPR3 has been implicated as a major regulator of cAMP production by the oocyte. The ability to manipulate follicle-enclosed oocytes should also pave the way for elucidating the mechanisms whereby LH stimulates mammalian oocyte maturation.

**Acknowledgements**

I thank Laurinda Jaffe and Bruce White for useful discussions and comments on the manuscript. This work was supported by grants from the Center for Interdisciplinary Research in Women’s Health at the University of Connecticut Health Center and the NIH. The authors declare that there is no conflict of interest that would prejudice the impartiality of this scientific work.

**References**


Andersen Y & Bykov AG 2002 Progestosterone and 17 alpha-OH-progesterone in concentrations similar to that of preovulatory follicular fluid is without effect on resumption of meiosis in mouse cumulus enclosed oocytes cultured in the presence of hypoxanthine. Steroids 67 941–945.


Ochsner S & Espey LL 2002 Ovulation: new findings. Topics in Developmental Biology 30 103–145.


Acquisition of meiotic competence in growing mouse oocytes is
controlled at both translational and posttranslational levels.
Developmental Biology 187 43–54.

Vivarelli E, Conti M, De Felici M & Siracusa G 1983 Meiotic
resumption and intracellular cAMP levels in mouse oocytes treated
with compounds which act on cAMP metabolism. Cell Differentiation
12 271–276.

Wang H & Storm DR 2003 Calmodulin-regulated adenylyl cyclases:
cross-talk and plasticity in the central nervous system. Molecular
Pharmacology 63 463–468.

Webb RJ, Bains H, Cruttwell C & Carroll J 2002a Gap-junctional
communication in mouse cumulus–oocyte complexes: impli-
cations for the mechanism of meiotic maturation. Reproduction
123 41–52.

hormone induces a gap junction-dependent dynamic change in
[cAMP] and protein kinase a in mammalian oocytes. Developmental Biology
246 441–454.

Zeleznik AJ 2004 Dynamics of primate follicular growth: a physio-
logical perspective. In The Ovary, edn 2, pp 45–53. Eds PCK

Received 6 May 2005
Accepted 26 May 2005