Signalling pathways involved in the control of sperm cell volume

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Abstract

The ability to maintain cellular volume is an important general physiological function, which is achieved by specific molecular mechanisms. Hypotonically induced swelling results in the opening of K+ and Cl− ion channels, through which these ions exit with accompanying water loss. This process is known as regulatory volume decrease (RVD). The molecular mechanisms that control the opening of the ion channels in spermatozoa are as yet poorly understood. The present study investigated pathways of osmo-signalling using boar spermatozoa as a model. Spermatozoa were diluted into isotonic and hypotonic Hepes-buffered saline in the presence or absence of effector drugs, and at predetermined intervals volume measurements were performed electronically. Treatment with protein kinase C (PKC) inhibitors staurosporine, bismaleimide I and bismaleimide X led to dose-dependent increases of both isotonic and hypotonic volumes (P<0.05). However, as the isotonic volume was affected more than the hypotonic volume, the kinase inhibitors appeared to improve RVD, whereas activation of PKC with phorbol dibutyrate blocked RVD. The increase in isotonic cell volume induced by bismaleimide X was observed in chloride-containing medium but not in the medium in which chloride was replaced by sulphate, implying that PKC was involved in the control of chloride channel activity, e.g. by closing the channel after volume adjustment. The protein phosphatase PP1/PP2 inhibitors calyculin and okadaic acid increased the isotonic volume only slightly but they greatly increased the relative cell volume and blocked RVD. The activation of RVD processes was found to be cAMP-dependent; incubation with forskolin and papaverine improved volume regulation. Moreover, papaverine was able to overcome the negative effect of protein phosphatase inhibitors. The mechanism of sperm RVD appears to involve (a) alterations in protein phosphorylation/dephosphorylation balance brought about by PKC and PP1 and (b) a cAMP-dependent activating pathway.

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Introduction

When cells encounter hypo- or hypertonic environments, they tend to swell or shrink due to the influx or efflux of water during re-establishment of osmotic equilibrium. Various animal cell types are able to maintain their volume after osmotic shock, thereby avoiding the consequences of excessive volume changes (Lang et al. 1998). During its life, the spermatozoon experiences considerable changes in its environment, most notably during maturation within the epididymis and at ejaculation (Yeung et al. 2004a and references therein). To be able to maintain cellular functionality in face of these osmotic changes, spermatozoa of several mammalian species (boar, mouse, bull and human) have been found to exhibit volume regulatory abilities, particularly regulatory volume decrease (RVD) in response to hypotonic challenge (Petrunkina et al. 2004a, and references therein).

It has been reported for a variety of cell types that the activation of independent K+ and Cl− channels is swelling-sensitive, as is that of organic osmolyte channels (Lang et al. 1998). Similar swelling-dependent activation of these channels has been demonstrated for spermatozoa of a variety of species, such as cattle, swine, human and dog (Kulkarni et al. 1997, Yeung et al. 2003, Petrunkina et al. 2001a, 2004a, 2004b). In somatic cells, RVD mediated by chloride and potassium currents is known to involve signalling cascades mediated by protein phosphorylation and dephosphorylation (Klein et al. 1993, Bize & Dunham 1994, Robson & Hunter 1994, Crepel et al. 1998, Musch et al. 1998, Voets et al. 1998, Thoroed et al. 1999, Hubert et al. 2000). In contrast, very little is known about whether phosphorylation processes are linked to the activation of osmo-sensing regulatory mechanisms in spermatozoa. Yeung et al. (1999, 2000) have hypothesized...
that the ability to regulate volume is important for achieving fertility and that tyrosine phosphorylation may be involved in RVD. Sperm from c-ros tyrosine kinase receptor knockout mice were infertile and showed impaired motility and other characteristics commensurate with a volume regulatory disturbance (though, as pointed out by the authors (Yeung et al. 2004b), these changes may have been due to an effect of c-ros knockout on the epididymis itself and thence disturbance of the maturation process). In domestic species (bull, boar and dog), it has been demonstrated that the control of sperm cell volume is closely related to the sperm’s ability to bind to the oviductal epithelium, and/or to their fertility competence in vivo; volumetric behaviour is also a crucial parameter for evaluating effects of cryoprotectants (Petrunkina et al. 2001b, 2004a, 2005a, Khalil et al. 2006).

Initial pharmacological and molecular evidence suggests that regulation of sperm cell volume under hypertonic conditions is mediated by protein tyrosine kinase (PTK)-dependent pathways (Petrunkina et al. 2005b). In that study, differential effects of the PTK inhibitor lavendustin and the general protein kinase inhibitor staurosporine (especially potent against protein kinase C, PKC) were observed, suggesting that phosphorylation reactions mediated by PTKs and/or by serine-threonine kinases such as the PKCs may play a major role in volume regulation of mammalian sperm. Thus far, there has not been any investigation of a putative role for cAMP in the control of cell volume. This study is the first seeking to carry out a comparative investigation of the roles of cAMP, the protein kinases PKA, PKC and PTK, and the protein phosphatases PP1, PP2A and TP (tyrosine phosphatase) in volume control in mammalian sperm, under both isotonic and hypotonic conditions.

Our study was based upon the boar sperm model and used electronic cell sizing to detect volume responses. Commonly used techniques such as protein profiling in sperm extracts revealed by Western blotting may be helpful in detecting and identifying the proteins likely to be involved in the mediation of cellular functions. However, these techniques neither allow characterization of the cell function in its physiological environment nor allow single-cell-based analysis: an appropriate screening of signalling pathways at the cellular level must be completed prior to performing focused studies at the molecular level. In the last decade, flow cytometry has been widely used to characterize sperm volumetric behaviour (Yeung et al. 2003, 2004a, 2005a, 2005b); although this technique is obviously more advantageous for functional studies on cell physiology, it allows only relative quantification of the cell volume; it is also greatly dependent on instrument settings. Volume analysis by electronic cell sizing, on the other hand, as well as providing single-cell information, allows exact calibration for cell volume quantification in physical units (Petrunkina et al. 2004a, 2004c). Moreover, the subtle changes in cell volume due to osmo-sensing are often below the threshold at which they can be detected by flow cytometry; such a drawback is especially critical for studies on spermatozoa (Petrunkina et al. 2004b, 2005a, 2005b). In our study, therefore, we have used electronic cell sizing to enable accurate and sensitive monitoring of osmotically and pharmacologically induced changes in sperm volume.

Materials and Methods

Chemicals

Unless otherwise stated, chemicals were obtained from Merck AG (Darmstadt, Germany), Alexis GmbH (Grüneberg, Germany), Merck KgaA (Calbiochem, Darmstadt, Germany) and Sigma AG (Sigma-Aldrich).

Semen sources

Semen was obtained from boars of fertile crossbred animals from the Institute of Reproductive Medicine’s boar colony. These samples were collected (generally twice a week) by the so-called gloved hand method via a sterile gauze (to remove gel). Immediately after collection, the semen was transferred to the laboratory and diluted in Beltsville thawing solution (BTS) extender (Johnson et al. 1988) to a concentration of between 0.8 and 1.0×10⁶ cells/ml, and stored at 17°C. Semen samples stored up to 72 h were used. In some experimental series, semen samples diluted in Androhep (Waberski et al. 1994) were also used.

The initial quality of ejaculates (volume, concentration, motility and morphology) was investigated as described by Weitze (2001). The minimal requirements for boar sperm quality were set according to the following criteria: semen volume 100 ml; sperm concentration 0.1×10⁶per µl; motility 70%; the total proportion of morphological alterations including plasma droplets 45%; the proportion of sperm with attached plasma droplets <25% (Weitze 2001). Only ejaculates which fulfilled the minimal requirements of boar sperm quality were subject to further experiments.

Just before experimentation, aliquots of diluted semen (3–5 ml) were washed through a two-step gradient of 35 and 70% isotonic Percoll-saline (Harrison et al. 1993). After removal of the supernatant layers, the loose sperm pellet was resuspended in residual 70% Percoll to a final concentration of about 2×10⁶ cells/ml. Semen samples were maintained throughout this procedure at a minimum of 25°C, and processed samples were used within 1 h.

Media

Two variants of a Hepes-buffered saline medium (HBS) were used as the vehicles for experimentation. The isotonic variant (isoHBS; 300 mosmol/kg) consisted of 137 mmol/l NaCl, 10 mmol/l glucose, 2.5 mmol/l KOH and 20 mmol/l Hepes buffered with NaOH to pH 7.4 at
39 °C. The hypotonic variant (hypoHBS; 180 mosmol/kg) was prepared by adjusting the NaCl content to approximately 76 mmol/l (Petrunkina et al. 2004a).

The isotonic medium based on sodium sulfate instead of sodium chloride was used for substitution experiments (Petrunkina et al. 2004a, 2005b). This medium (isoSulf, 300 mosmol/kg) consisted of 115 mmol/l sodium sulfate, 10 mmol/l glucose, 20 mmol/l Hepes buffered with NaOH to pH 7.4 at 39 °C. To minimize the detection of particulate noise during cell volume measurements, all media were passed through a 0.2 μm filter before use.

**Experimental procedure**

In essence, the experimental approach consisted of diluting washed sperm samples at 39 °C in iso- or hypotonic medium containing specific effectors, and then taking subsamples at timed intervals for analysis of the population volume distributions. Effects were largely judged by comparing the modal volumes of such distributions. The pharmaceuticals, their effects and the ranges of concentration used are given in Table 1.

The general experimental procedure was as follows: aliquots of washed sperm samples were diluted in 1 ml volumes of isoHBS at 39 °C containing predetermined concentrations of effectors (final sperm cell concentration between 0.5 and 1 × 10⁷ cells/ml). After 10-min pre-incubation, samples were transferred to 5 ml of either hypoHBS or isoHBS containing the same concentration of effector as the parent medium, and incubated further at 39 °C (final sperm concentration about 1 × 10⁶ cells/ml). The isotonic and hypotonic sperm suspensions were

### Table 1 Effectors used in this study.

<table>
<thead>
<tr>
<th>Compound</th>
<th>Abbreviation</th>
<th>Established effects</th>
<th>Range tested</th>
<th>Previously detected effects on sperm function</th>
</tr>
</thead>
<tbody>
<tr>
<td>Staurosporine</td>
<td>Sta</td>
<td>Inhibits protein kinase C (IC₅₀ = 0.7 nM), protein kinase A (IC₅₀ = 7 nM), protein kinase G (IC₅₀ = 8.5 nM), CaM kinase (IC₅₀ = 20 nM)</td>
<td>0.1–1 μM</td>
<td>0.1–2 μM inhibits ZP-induced AR⁸ and 1–10 μM inhibits increase in merocyanine binding due to bicarbonate⁹</td>
</tr>
<tr>
<td>N-(2-[p-Bromocinnamylamino) ethyl]5-isoquinolinesulfonamide</td>
<td>H-89</td>
<td>Inhibits protein kinase A (Kᵢ = 48 nM) and other kinases at high concentrations: PKC (Kᵢ = 32 μM)</td>
<td>5–15 μM</td>
<td>10–50 μM inhibits increase in merocyanine binding⁹</td>
</tr>
<tr>
<td>Bisindolylmaleimide I, GF-109203x (2-[1-(3-methylaminopropyl)-1H-indol-3-yl]-3-[1H-indol-3-yl]-maleimide)</td>
<td>Bis I</td>
<td>Inhibits protein kinase C with improved selectivity as compared to staurosporine (Kᵢ = 10 nM) and protein kinase A (&gt; 2 μM)</td>
<td>1–5 μM</td>
<td>100 nM decreases calcium influx and acrosome reaction⁶</td>
</tr>
<tr>
<td>Bisindolylmaleimide X 2-[8-aminomethyl-6,7,8,9-tetrahydropyrido[1,2-a]indol-3-yl]-N-methylmaleimide</td>
<td>Bis X</td>
<td>Selective inhibitor of PKC (IC₅₀ = 15 nM)</td>
<td>0.3–3 μM</td>
<td></td>
</tr>
<tr>
<td>Phorbol-12,13-dibutryrate</td>
<td>PDBu</td>
<td>Activates protein kinase C</td>
<td>0.3–3 μM</td>
<td></td>
</tr>
<tr>
<td>Forskolin (7β-Acetoxy-8,13-epoxy-1α, 6β, 9α-trihydroxy-14 en-11-one)</td>
<td>Forskolin</td>
<td>Activates adenylate cyclase (EC₅₀ = 4 μM), leading to elevated cAMP levels</td>
<td>1–10 μM</td>
<td>10 μM increases sperm cAMP levels⁴; 100 μM increases sperm capacitation rates &amp; phosphorylation⁶</td>
</tr>
<tr>
<td>1,9-Dideoxyforskolin (7β-Acetoxy-6β-hydroxy-8,13-epoxy-14 en-11-one)</td>
<td>DDFS</td>
<td>Does not activate adenylate cyclase. Inhibits chloride channels</td>
<td>1–10 μM</td>
<td>1 μM inhibits regulatory volume decrease⁴</td>
</tr>
<tr>
<td>Papaverine 6,7-dimethoxy-1-teratrylisoquinoline hydrochloride</td>
<td>Pap</td>
<td>Inhibitor of cAMP phosphodiesterase</td>
<td>20 μM</td>
<td>20 μM enhances merocyanine binding in the presence of PP inhibitors⁵; enhances iso- and hypertonic volume⁵</td>
</tr>
<tr>
<td>Lavendustin A 5-[[2,5-dihydroxy-phenylmethyl][2-hydroxy-phenylmethyl]amino]-2-hydroxybenzoic acid</td>
<td>Lav A</td>
<td>Potent inhibitor of the protein tyrosine kinases EGFR kinase and p60c-src (IC₅₀ = 11 nM and 500 nM). Does not inhibit PKA or PKC</td>
<td>1–5 μM</td>
<td>1–15 μM enhances isotonic and hypertonic volume⁵</td>
</tr>
<tr>
<td>Okadaic acid</td>
<td>OA</td>
<td>Potent inhibitor of protein phosphatases PP1 (IC₅₀ = 0.1–10 nM) and PP2A (IC₅₀ = 0.1 nM). Does not affect tyrosine phosphatase activity</td>
<td>10–100 nM</td>
<td>20–200 nM enhances merocyanine binding in the presence of papaverine⁹</td>
</tr>
<tr>
<td>Calyculin</td>
<td>Cal</td>
<td>Potent inhibitor of PP1 (IC₅₀ = 2 nM) and PP2A protein phosphatases (IC₅₀ = 0.5–1 nM)</td>
<td>10–100 nM</td>
<td>50 nM increases motility⁹; if papaverine present, 20–200 nM enhances merocyanine binding⁹</td>
</tr>
<tr>
<td>Sodium orthovanadate</td>
<td>Vanadate</td>
<td>Inhibitor of protein tyrosine phosphatases with broad specificity</td>
<td>100 μM</td>
<td></td>
</tr>
</tbody>
</table>


*Petrunkina et al. (2005b); ⁶Smith et al. (1996).
sampled for cell volume measurement after predetermined periods (between 5 and 20 min). IsoHBS and hypoHBS without additives were used as controls. When an effector was added from a stock solution prepared in alcohol or DMSO, the final concentration of solvent in the incubation medium was always <0.5% (v/v), a level which had no significant effect on the volumetric behaviour of the spermatozoa under the experimental conditions tested.

**Electronic volume analysis**

At each sampling time, a single sample from each incubated sperm suspension was passed through a CASY 1 cell counter (Schaerfe Systems GmbH, Reutlingen, Germany). This instrument produces cell volume information on the basis of cell frequency distribution. During the measurements, the cells suspended in the electrolyte solution pass through a capillary pore. The changes in electric resistance caused by each cell passing through the pore result in voltage changes proportional to the cell volume. Because the measurement signal is registered at a frequency of 1 MHz, cell passage produces a series of individual signal pulses; the voltage change characteristics for each cell are calculated by integration of these pulses (pulse area analysis). This methodology guarantees high accuracy and resolution of measurements; the pulse area values are neither dependent on the orientation of the sperm cells as they pass through the capillary pore nor on the geometrical shape of the cells. The recorded electric signals are converted to 'volume equivalent' values using a channel analyser with 512 000 volume measurement channels corresponding to the pulse 'areas' produced by each cell. The resultant values are then recalculated to a linear distribution within 1024 effective cell size channels.

In our experiments, the sample volume setting was 200 µl and the size scale 10 µm (i.e. the system was set to record particles with diameters up to 10 µm, which corresponds to a volume equivalent value of about 524 fl); each sampling obtained data from more than 10 000 cells so that overall the resolution was approximately 0.1 µm per effective size channel.

Because the electrical conductivity of hypoHBS was different from that of isoHBS (due to different concentrations of electrolyte), data recorded from sperm suspensions in the former medium were multiplied by a correction factor to obtain the real physical volume (see Petrunkina & Töpfer-Petersen 2000). Volume data are presented as real volumes in femtolitres (fl: 10⁻¹⁵ l).

**Analysis of volumetric data**

Unless otherwise stated, treatment effects were judged by comparing the modal values of the sample volume distributions, since modal volume is the most sensitive parameter of response to osmotic stress (Petrunkina & Töpfer-Petersen 2000). By comparing modal volumes of different particle subpopulations within the sample, it was possible to differentiate between spermatozoa and debris (see Petrunkina et al. 2004a), and only the spermatozoa were considered. (Note that the high degree of differentiation between peaks related to spermatozoa and those related to debris is improved by the Percoll-gradient centrifugation used to prepare the original sperm samples: this leads to large-scale elimination of diluent, free plasma droplets and sperm fragments: Petrunkina et al. 2004b).

The relative volume shift \( V_r \) was used as a measure of the volume response to hypotonic conditions. It was defined as \( V_r = V_{\text{hypo}}/V_{\text{iso}} \) where \( V_{\text{hypo}} \) is the modal value of the hypotonic volume distribution and \( V_{\text{iso}} \) the modal value of the isotonic volume distribution. \( V_{15} \) describes the relative volume increase in the face of hypotonic challenge after 5 min and \( V_{20} \) its change (volume decrease) after 20 min. The RVD was defined as the recovery of the relative hypotonic volume \( \text{RVD} = V_{20} - V_{15} \). When several peaks appeared within a volume distribution (see Petrunkina et al. 2004b), the volume of the first most abundant osmotically active subpopulation peak was used for the analysis; the subpopulation was considered to be osmotically active when the relative volume shift \( V_r = V_{\text{hypo}}/V_{\text{iso}} \) was > 1, indicating that the spermatozoa within this population were able to swell, thus were alive (Petrunkina & Töpfer-Petersen 2000).

The absolute cell volumes under both isotonic and hypotonic conditions were compared between the treatments, and the relative volume shifts attributed to the different treatments were compared with those in control samples. The response to effector treatment under isotonic conditions was judged by relating the modal volume of the resultant main peak to that of the sperm peak in the control sample.

**Flow cytometric estimation of cell viability**

Sperm samples were labelled for 10 min with propidium iodide (PI, 5 µg/ml final concentration); this dye binds to DNA in membrane-damaged cells and is now commonly used for recording the membrane integrity (cf. Harrison & Vickers 1990). The stained samples were analysed with a Dako Galaxy flow cytometer (Dako GmbH, Hamburg, Germany). For each sample, a single measurement was made. The cells were excited at 488 nm using an argon laser, and propidium iodide fluorescence was detected using a 610 nm band-pass filter (FL-3) on a logarithmic scale. Forward- and side-scatter data (FSC and SSC) were collected in the linear mode.

For each day's analyses, the control settings were adjusted as follows. An unstained sperm sample was first passed through the instrument. The forward- and side-scatter settings were adjusted so that the sperm population was presented as an L-shaped profile. The fluorescence detector voltages were adjusted after
measuring the sample stained only with PI so that the dead cells were clearly distinguishable from the live (unstained) cells on the Y-axis (FL-3). Unstained cells were detected near the origin with respect to the X-axis. Data were analysed using Flomax software (Vs 2.0, 1999, Partec GmbH, Münster, Germany). The FL-3 fluorescence data were displayed as one-dimensional histograms for determination of proportion of cells responding to the treatment. Ranges were delineated to discriminate between membrane-intact and membrane-defective spermatozoa.

**Data analysis**

The results presented are based on the means from three and six independent experiments. Observed effects were verified by ANOVA, t-test and non-parametrical analysis (SAS Software, SAS Institute Inc., Cary, NC, USA). Unless stated otherwise, the values presented are mean ± S.E.M. Differences were considered to be significant if the calculated probability of their occurring by chance was <5% (P<0.05).

**Results**

**Protein kinase-dependent pathways**

Four inhibitors of protein kinases were probed in this experimental series in the ranges indicated in Table 1: staurosporine (potent general inhibitor of protein kinases with very high potency for PKC, though PTK also inhibited and, to a much lesser extent, PKA), Bis I (PKC inhibitor with less potency for PKA), lavendustin A (inhibitor of PTK) and H-89 (PKA inhibitor). The addition of protein kinase inhibitors to incubation of the sample resulted in a concentration-dependent increase (P<0.01) in the isotonic sperm volume (Fig. 1 and Table 2). Even at 1 μmol/l concentration, staurosporine, Bis I and lavendustin A caused 40–50% increases in isotonic volume when compared with control values (P<0.05), whereas a comparable effect of the PKA inhibitor H89 was observed only at a concentration of 15 μmol/l.

Similarly, incubation with the protein kinase inhibitors resulted in a concentration-dependent increase (P<0.05) in hypotonic cell volume (Table 2, and compare Fig. 2a and b). However, the increase in volume caused by exposure to hypotonic conditions in the presence of staurosporine, lavendustin A or H-89 was similar to the increase in isotonic volume (e.g. Fig. 2a and b). Thus, the relative volume shift after 5 and 20 min was generally comparable to or lower than that in the control, indicating the maintenance of volume regulatory response (Fig. 3). Nevertheless, in our experiments, due to considerable variation between samples, the tendency of acceleration of volume regulatory response was not statistically significant. Treatment with the PKC inhibitor Bis I, on the other hand, resulted in both higher initial swelling and a very strong degree of RVD; after 20 min of exposure to hypotonic medium in the presence of 5 μM Bis I, the relative volume shift was lower (Vr=1.06) than in the control (Vr=1.32; P<0.06) and the regulatory volume change (ΔVr=Vr,20-Vr,5) was significantly higher than in the control (−0.88 in Bis I-treated samples vs −0.38 in control; P<0.05).

Unfortunately, the treatment of sperm with Bis I led to considerable increases in cell death (at a concentration of 5 μmol/l, 60–70% of sperm became PI-positive), whereas incubation with the other inhibitors even at the highest concentrations tested did not significantly affect the membrane integrity (PI-positive cells: 11–17% in staurosporine, 12–24% in lavendustin, 14–23% in H89 as against 10–15% in controls). It should be pointed out that this effect of Bis I did not invalidate our conclusions because we were observing swelling effects, and any swelling observed was necessarily due to the response of membrane intact, i.e. live, cells. However, because of the toxic effect of Bis I, we sought another PKC inhibitor, which was less detrimental. Bisindolylmaleimide X (Bis X), which specifically inhibits PKC with an IC50 similar to Bis I, and bisindolylmaleimide V (Bis V), a negative control, had no

**Table 2** Effect of protein kinase inhibitors on isotonic and hypotonic volumes.

<table>
<thead>
<tr>
<th>Inhibitors</th>
<th>Concentration (μmol/l)</th>
<th>V_iso 20 min (fl)</th>
<th>V_hypo 20 min (fl)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>–</td>
<td>15.7±0.4</td>
<td>20.8±1.2</td>
</tr>
<tr>
<td>Staurosporine</td>
<td>0.1</td>
<td>18.2±1.4</td>
<td>27.6±4.8</td>
</tr>
<tr>
<td></td>
<td>1</td>
<td>22.3±2.6</td>
<td>30.4±4.8</td>
</tr>
<tr>
<td>Bis I</td>
<td>1</td>
<td>21.4±3.1</td>
<td>25.3±2.3</td>
</tr>
<tr>
<td></td>
<td>5</td>
<td>31.9±2.4</td>
<td>32.8±1.7</td>
</tr>
<tr>
<td>Lavendustin A</td>
<td>1</td>
<td>21.2±2.8</td>
<td>29.5±4.2</td>
</tr>
<tr>
<td></td>
<td>5</td>
<td>30.1±1.47</td>
<td>36.0±4.8</td>
</tr>
<tr>
<td>H-89</td>
<td>15</td>
<td>18.5±1.7</td>
<td>24.5±2.9</td>
</tr>
<tr>
<td></td>
<td></td>
<td>21.6±2.4</td>
<td>34.0±3.4</td>
</tr>
</tbody>
</table>

Figure 1 Effect of staurosporine, a protein kinase inhibitor, on sperm cell volume in isotonic medium. Solid line indicates control (no inhibitor) sperm volume distribution; broken line indicates volume distribution in the presence of 5 μmol/l staurosporine. Note that the isotonic volume distribution curve is shifted to the right by the protein kinase inhibitor; in other words, the cell volume of the majority of cells in the population is increased.

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significant effect on cell death in concentrations between 0.3 and 1 µmol/l (PI-positive cells: 12–26% in Bis X, 9–21% in Bis V, 8–23% in controls).

As expected, after addition of Bis X to the incubation suspensions the isotonic sperm volume increased in a dose-dependent fashion. However, at 1 µmol/l the effect of Bis X was not as dramatic as that of Bis I – the former induced a volume increase of about 21% as compared with the control (no inhibitor) volume, whereas the latter induced an increase of about 36%. There were no changes in isotonic cell volume in samples treated with Bis V, but after 20 min of exposure to hypotonic conditions, the hypotonic cell volume (and V_r) still remained high, whereas V_r was decreased in Bis X-treated cell suspensions (Fig. 4a). Although the inactive analogue, Bis V, appeared to produce an enhancement of RVD.

Figure 2 Effect of the protein kinase inhibitor staurosporine on osmotically induced cell volume regulation in spermatozoa. (a) Volume shift in control media (no kinase inhibitor). Solid line indicates sperm volume distribution in isotonic medium; broken line indicates volume distribution in hypotonic medium. The volume distribution is shifted to the right side in hypotonic medium as compared with the distribution in isotonic medium, demonstrating swelling of the cells in response to hypotonic conditions. (b) Volume shift in the presence of 5 µmol/l staurosporine. Solid line indicates volume distribution in isotonic medium, broken line indicates volume distribution in hypotonic medium. Both distributions are shifted to the right as compared with the control distributions in (a). However, the relative degree of swelling resulting from hypotonic stress (shaded area) was less in the presence of staurosporine than it was in the control. This demonstrates an enhancement of RVD.

Figure 3 Effect of protein kinase inhibitors on sperm cell volume regulation. Data are presented as V_r = V_hypo/V_iso. Open bars are V_r, closed bars are V_r. Control, no inhibitor present; Sta, 1 µmol/l staurosporine; Bis I, 5 µmol/l bisindolylmaleimide I; Lav A, 5 µmol/l lavendustin A; H-89, 15 µmol/l H-89. Note that because of sample variation, only the changes brought about by Bis I were statistically significant (P<0.05).

Figure 4 Effect of protein kinase C effectors on sperm cell volume regulation. (a) Effects of bisindolylmaleimides X and V. Bis X (solid symbols), a specific inhibitor of protein kinase C, led to lowered relative volume shift after 20 min of incubation under hypotonic conditions as compared with control (no effector present), i.e. it enhanced volume recovery after initial swelling. In the presence of Bis V (open symbols), an inactive analogue of Bis X, swelling levels after 20 min remained high or even increased. (b) Effects of phorbol esters on volume regulation. Solid symbols, PDBu; open symbols, α-PDBu. While elevated swelling levels were observed in response to 1 µmol/l PDBu, an activator of PKC, the relative volume was considerably lower in the presence of a similar concentration of its inactive analogue, α-PDBu.
elevation in the hypotonic cell volume when compared with the control hypotonic volume, a markedly significant difference was observed between the effects of the active and inactive analogues; at all concentrations tested, the relative volume shift was higher in the cell suspensions incubated with Bis V than in those incubated with Bis X (P<0.05, Fig. 4a).

The results above suggested strongly that PKC played an important role in volume control. To confirm this, we investigated the effect of PKC activation with the phorbol dibutyrate PDBu at concentrations between 0.3 and 3 μmol/l, as the levels of membrane-defective sperm were found not to be significantly increased at these concentrations. The isotonic volume was not affected either by PDBu or by α-PDBu (an inactive phorbol ester), although the latter led to values nominally larger than those of the controls (not significant; P>0.05). After 5 min exposure to hypotonic conditions, there was no difference in Vr between samples treated with either PDBu or α-PDBu (data not shown), but Vr was increased by PDBu in a dose-dependent fashion after 20 min exposure (Fig. 4b). After 20 min in the presence of 3 μmol/l PDBu, the hypotonic volume increased to 20.8 fl as compared with 11.8 fl in control suspensions (P<0.01). At 0.3 μmol/l PDBu, no significant effects were observed due to big sample variations but RVD was inhibited at 1 μmol/l concentration of PDBu; the swelling progressed over 20 min (ΔVr = 0.67), whereas it showed a recovery (ΔVr = −0.45) in control suspensions (P=0.054) and was maintained in place in α-PDBu-treated suspensions (ΔVr = 0.00). Furthermore, the volume set point (i.e. Vr,20) was 1.88 in PDBu-treated suspensions (at 3 μM), whereas it was 1.22 in the control (P<0.05).

To see if PKC was acting to control chloride fluxes, the effects of Bis X were tested in isotonic medium in which chloride was replaced by sulfate (isoSulf). In isoSulf, the increase in Vr,iso induced by Bis X in HBS was abolished and replaced by a slight volume decrease. Whereas 3 μmol/l Bis X produced a Vr,iso,20 of 19.9±1.9 fl in HBS, in isoSulf it produced a Vr,iso,20 of 11.6±1.0 fl (P<0.05 for difference). These Vr,iso,20 values compared with control values of 12.6±0.6 fl in HBS.

These results suggested strongly that the increased isotonic volume caused by inhibition of PKC is due to the opening of chloride channels, and that PKC acts to maintain these channels in an inactive state. The increased Vr resulting from PKC activation would seem to be due to the defective opening of the channels and thus the inability of Cl− to exit with K+ to maintain tonicity across the cell plasma membrane.

**Protein phosphatase-dependent pathways**

Given the effect of protein kinase inhibitors, it seemed logical to investigate the involvement of the dephosphorylation processes in sperm volume control. Two inhibitors of protein phosphatase PP1 and PP2A, okadaic acid and calyculin, were tested at concentrations of 10 and 100 nmol/l (see Table 1). Although both inhibitors led to increased isotonic volumes, the effects (Table 3) were not nearly as large as those of the protein kinase inhibitors (3–4 vs 10–20 fl) and occurred only after longer incubation.

At a dosage of 10 nmol/l, both inhibitors led to increased hypotonic volumes, which varied between 43.7 and 49.2 fl as compared with a control hypotonic volume of 25.4 fl (P<0.05; see Table 3). Whereas protein kinase inhibitors had similar marked effects on both isotonic and hypotonic volumes, the protein phosphatase inhibitors had a much more pronounced effect on hypotonic than on isotonic volume (compare Fig. 5b with a). At 10 nmol/l okadaic acid or calyculin, at which no effect on isotonic cell volume was observed, RVD was blocked (Fig. 6). The relative volume change ΔVr showed an increase in treated samples (+0.3 and +0.50 for okadaic acid and calyculin respectively), whereas in control samples, it showed a decrease (−0.64).

Treatment with ortho-vanadate, an inhibitor of TP, led to somewhat increased isotonic volumes, but this tendency was not significant. There was also a tendency towards reduction of Vr after 5 min, and after 20 min RVD was blocked significantly (P<0.05). There was a progressive increase in the cell volume (ΔVr = +0.21), whereas in the control the normal markedly progressive decrease in cell volume was observed (ΔVr = −0.52; P<0.05).

**cAMP-dependent pathways**

Cyclic AMP effects are widely mediated via its activation of PKA. Yet, during the experiments with protein kinase inhibitors, it was noticeable that inhibition of PKA with H-89 did not have a great effect on isotonic volume (Table 2) although it did tend to reduce hypotonic swelling (non-significantly; Fig. 3). While PKA is the best-known target of cAMP, the second messenger can act by other means. There are various ways to elevate

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<th>Table 3 Effects of protein phosphatase and phosphodiesterase inhibitors on isotonic and hypotonic cell volumes.</th>
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OA, 10 nM okadaic acid; Cal, 10 nM calyculin; Pap, 20 μM papaverine. *Values labelled with asterisks differ significantly from the controls within the same row (P<0.05). Effect of papaverine alone (20 μM) were tested against split-sample controls in a separate experiment.
cAMP levels: for example, direct stimulation of adenylate cyclase activation by forskolin (Leclerc & Kopf 1995) or inhibition of cAMP phosphodiesterase by papaverine (reported particularly for boar sperm, Harrison & Miller 2000). Both methods were used here.

Forskolin (FSK) and its biologically inactive analogue dideoxyforskolin (DDFSK) were tested at concentrations (reported particularly for boar sperm, Harrison & Miller 2000). Both methods were used here. Forskolin (FSK) and its biologically inactive analogue dideoxyforskolin (DDFSK) were tested at concentrations (reported particularly for boar sperm, Harrison & Miller 2000). Both methods were used here. Forskolin (FSK) and its biologically inactive analogue dideoxyforskolin (DDFSK) were tested at concentrations (reported particularly for boar sperm, Harrison & Miller 2000). Both methods were used here. Forskolin (FSK) and its biologically inactive analogue dideoxyforskolin (DDFSK) were tested at concentrations (reported particularly for boar sperm, Harrison & Miller 2000). Both methods were used here. Forskolin (FSK) and its biologically inactive analogue dideoxyforskolin (DDFSK) were tested at concentrations (reported particularly for boar sperm, Harrison & Miller 2000). Both methods were used here.

At the lowest concentration tested (1 μmol/l), both FSK and DDFSK had similar effects on the hypotonic volume. RVD was inhibited. The relative volume shift remained at a stable high level (between 2.19 and 2.12) during 20 min incubation with FSK under hypotonic conditions, and even increased significantly during incubation with DDFSFK (V_{r,5} = 2.32; V_{r,20} = 2.95), while RVD was observed in the control (V_{r,5} = 2.32; V_{r,20} = 1.90; P<0.05). Above this concentration, incubation with FSK caused a decrease in V_r after 5 and 20 min of exposure (i.e. RVD was enhanced; Fig. 7b). Although inhibition of RVD was also reversed at higher concentrations of DDFSFK, this occurred more slowly, i.e. after 20 min and not after 5 min incubation; thus the effect of FSK was faster than that of DDFSFK. At 10 μmol/l, there was a significant difference between forskolin and its inactive analogue in their effect on relative sperm volume after 5 min (V_{r,5} = 1.48 and V_{r,5} = 2.11 respectively).

At a concentration of 20 μmol/l, the inhibitor of cAMP phosphodiesterase, papaverine, caused a significant increase in isotonic volume both after 5 min (17.9 fl; control, 14.5 fl) and after 20 min (19.3 fl; control, 15.1 fl, Table 3); it also caused a marked acceleration of RVD, V_{r,5} = 1.46 as compared with the control, V_{r,5} = 1.93 (P<0.05). The addition of papaverine to the samples incubated with okadaic acid and calyculin induced isotonic swelling when the phosphatase inhibitors alone had no effect (Table 3). Moreover, papaverine reversed the increased hypotonic volume and relative swelling induced by the phosphatase inhibitors (Table 3; Fig. 6). Although resulting cell volumes were much greater than in the control suspensions (P<0.05), and usually greater than with phosphodiesterase inhibitor alone, the relative volume shifts were diminished after 5 min and especially after 20 min (Fig. 6). Although cell death increased significantly during treatment with 100 nmol/l concentrations of the phosphatase inhibitors, the proportion of

Figure 6 Effect of protein phosphatase inhibitors and papaverine on sperm cell volume regulation. OA, 10 nmol/l okadaic acid; Cal, 10 nmol/l calyculin; Pap, 20 μmol/l papaverine. Open bars are V_{r,5}; closed bars are V_{r,20}. Inhibiting protein phosphatase blocked the RVD process. However, Pap accelerated RVD (lower V_{r,5} than control) and inclusion of Pap with OA overcame the latter's blocking of RVD (the effect of Pap with Cal was much less clear). This effect of Pap indicates an involvement of cAMP in the RVD process. Effects of papaverine alone (20μM) were tested against split-sample controls in a separate experiment.

Figure 5 The effects of calyculin, a PP1 protein phosphatase inhibitor, on sperm cell volume. (a) Effect on the isotonic volume distribution curve (20 min exposure, 100 nmol/l calyculin, 300 mosmol/kg). Solid line indicates control (no inhibitor) curve; broken line indicates calyculin curve. (b) Effect on the hypotonic volume distribution curve (20 min exposure, 100 nmol/l calyculin, 180 mosmol/kg). The degree of shift of the curve produced by calyculin is much greater under hypotonic conditions, i.e. RVD is blocked.

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propidium iodide-positive cells remained between 25 and 30%, even after the addition of papaverine (control: 15%), so that most cells in the population may be considered to have been intact. At 10 nmol/l, no effect of calyculin or okadaic acid on membrane integrity was seen. Thus, the effects detected could be clearly attributed to changes in the live cell population.

As a general note regarding the results described above, perceptive readers may wonder whether the presence of subpopulations of spermatozoa bearing attached cytoplasmic droplets might have distorted our findings. Although the additional cellular volume due to attached and/or free cytoplasmic droplets is detectable within population distributions obtained from electronic sizing (9–11 µm³ at 20 °C; O’Donnell 1969, Brotherton 1975), efferents were tested with respect to changes induced in modal volumes, not means or medians, so that the contribution of minor populations of droplets (attached or free) to the osmotic response would not have affected materially the overall response of the main sperm populations.

Discussion

The findings from the experiments described above have been able to link mechanisms for maintaining sperm cell volume under isotonic conditions with those leading to RVD after exposure of the sperm cell to hypotonic conditions. RVD involves swelling-sensitive opening of K⁺ and Cl⁻ channels to allow exit of ions so as to maintain osmotic equilibrium, whereas maintenance of cell volume under isotonic conditions requires that these channels be maintained in an inactive (closed) state. Premature activation of the channels under isotonic conditions results in the entry of Cl⁻ and Na⁺ down concentration gradients, whence increased osmotic pressure and resultant swelling to re-establish osmotic equilibrium. Our results have shown for the first time that PKC is involved in maintaining sperm isotonic volume by suppressing ion transport, whereas cyclic AMP is involved in stimulating sperm RVD by activating ion transport. Activation of sperm ion transport and RVD also involves the serine/threonine phosphatase PP1. A schematic representation of our findings is shown in Fig. 8.

PKC inhibitors led to sperm swelling in both isotonic and hypotonic media (Figs 1 and 2a and b). That inhibitor-induced isotonic swelling did not take place in a medium in which Cl⁻ had been replaced with sulfate suggests that PKC acts largely on chloride ion channels. We have recently shown that certain inhibitors of chloride channels (e.g. tamoxifen) lead to increased isotonic volume, whereas others (e.g. NPPB(5-nitro-2-(3-phenylpropylamino)-benzoic acid)) affect only hypotonic volume (Petrunkina et al. 2004a). An alternative explanation for the effect of kinase inhibitors on isotonic volume in spermatozoa might have been that they caused enhancement of taurine flux (e.g. in glial cells; Deleuze et al. 2000); however, it has recently been shown that taurine efflux is not essentially involved in volume control in boar or human spermatozoa, although it does play a role in mouse sperm RVD (Yeung et al. 2003, 2004, Petrunkina et al. 2004a). While staurosporine, lavendustin and Bis I led to comparable increases in isotonic volume, the effect of the PKC inhibitor Bis X was less. This suggests that the maintenance of transport mechanisms in an inactive state involves both PTK- and PKC-dependent pathways. It has recently been shown that tyrosine phosphorylation is involved in the sequence leading to the deactivation of sperm volume regulatory mechanisms (Petrunkina et al. 2005b), whereas in somatic cells ion channels are opened by tyrosine kinase (Lepple-Wienhues et al. 2000). Moreover, some ion channels in human-derived renal epithelial cell line HEK 293 cells, such as the voltage-gated potassium channel of the KCNQ4 family, have been reported to be inactivated via protein kinase C activity under isotonic conditions (Hougaard et al. 2004).
Reproduction was equivocal.

Results in increased cAMP levels and similar effects. Notably, increased to increased isotonic cell volume and to accelerated swelling recovery activation of volume regulatory mechanisms usually inactive under cAMP-sensitive. An increase in intracellular cAMP is associated with decrease. Activation of the volume regulatory mechanism appears to be forms). (b) cAMP-dependent pathway activating regulatory volume phosphorylation of the residues phosphorylated by PKC?). Thus, inhibition of protein phosphatase PP1 results in blocked RVD. A PTK/TP-dependent system regulated pathways controlling maintenance of cell volume. PKC is involved in the signalling sequence that leads to deactivation of the osmo-dependent regulatory mechanism, probably by closing and keeping closed a volume-sensitive anion channel. Inhibition of PKC results in premature activation of the channel, increased isotonic cell volume and prolonged and/or accelerated volume regulation. Protein phosphatase PP1 is involved in the signalling sequence that leads to the activation of the volume regulatory mechanism, probably by opening the volume-sensitive ion channels (by dephosphorylation of the residues phosphorylated by PKC?). Thus, inhibition of protein phosphatase PP1 results in blocked RVD. A PTK/TP-dependent system may regulate PP1 activity (transition between active and inactive forms). (b) cAMP-dependent pathway activating regulatory volume decrease. Activation of the volume regulatory mechanism appears to be cAMP-sensitive. An increase in intracellular cAMP is associated with activation of volume regulatory mechanisms usually inactive under isotonic conditions: activation of adenylate cyclase by forskolin leads to increased isotonic cell volume and to accelerated swelling recovery in response to hypotonic conditions. Inhibition of phosphodiesterase results in increased cAMP levels and similar effects. Notably, increased cAMP levels can overcome inhibition of the PP1-dependent pathway. Evidence for the involvement of PKA in the cAMP-dependent pathway was equivocal.

Hypotonic conditions can lead to the activation of protein kinase C (Lang et al. 1998). Larsen et al. (1994) reported that this activation occurs during late stages of RVD. The fact that the PKC inhibitor Bis I enhanced the amplitude of sperm RVD and resulted in a lowering of the volume set point, i.e. the resultant volume following RVD (see Fig. 3), agrees with their findings and suggests that PKC may be responsible for the termination of RVD, i.e. by closing an ion channel (see Fig. 8). (Note that the internal accumulation of Cl− and Na+ ions during isotonic swelling caused by the opening of channels (see above and Fig. 1) will tend to amplify the RVD process because the lower NaCl concentration in the external hypotonic medium will initially induce water influx into the cells. Moreover, if the isotonic swelling during drug pretreatment was not due to the premature opening of ion channels, no relative shrinking during hypotonic exposure would take place; rather, a further progressive swelling would be observed under hypotonic conditions.) Our findings mirror those of Hofmann (2000) who reported that another PKC inhibitor, chelerythrine, decreased the volume set point in Ehrlich ascites tumour cells after RVD. Further evidence that PKC acts to close ion channels is provided by our demonstration that the PKC activator phorbol dibutyrate causes elevated levels of relative swelling in response to hypotonic conditions (Fig. 4b).

Although PTK-dependent pathways appear to be involved in sperm volume regulation under both isotonic and hypertonic conditions (Petrunkina et al. 2005b), tyrosine phosphorylation seems to play a less crucial role in sperm exposed to hypotonic conditions (see effects of lavendustin and vanadate).

Under hypotonic conditions, protein serine/threonine phosphatase inhibitors had a marked effect on RVD. Both calyculin A and okadaic acid inhibited RVD at low concentrations (10 nmol/l). It has been suggested that swelling in response to hypotonic conditions inhibits a kinase, favouring dephosphorylation and leading to transport mechanism activation (Dunham et al. 1993, Bize & Dunham 1994). Okadaic acid has been shown to block activation of the volume-sensitive chloride current in a non-pigmented ciliary epithelial cell line (Shi et al. 2002), indicating that dephosphorylation is involved in the opening of chloride channels in this cell type. In our experiments, calyculin was more efficient than okadaic acid in inhibiting RVD: at 10 nmol/l, its inhibitory effect was barely affected by papaverine, whereas the inhibitory effect of 10 nmol/l okadaic acid was largely reversed by papaverine (see Table 3 and Fig. 6). Thus, as both inhibitors have approximately the same potency against PP2A, and calyculin has a higher potency against PP1, we conclude that it is PP1 that is involved in the signalling sequence leading to channel opening in sperm (Fig. 8).

Our findings indicate that cAMP plays a major role in ion channel activation. At low concentrations, forskolin, a potent stimulator of adenylate cyclase, increased isotonic volumes (Fig. 7a) and enhanced RVD (Fig. 7b). The phosphodiesterase inhibitor papaverine, at a concentration sufficient to elevate cAMP levels in boar sperm (Harrison & Miller 2000), also led to a significant increase in both isotonic volume and RVD. As the cAMP effectors had an accelerating effect on RVD as well as an enhancing one, it is likely that cAMP is involved in the signalling pathway, which leads to the opening of the channel (Fig. 8). This hypothesis is supported by the fact that papaverine was able to reverse the inhibition of RVD by okadaic acid at 10 nmol/l (Fig. 6). Our finding is interesting because, although cAMP has been reported to decrease the volume of a wide variety of cells
maintenance of RVD in salivary acinar cells isolated in 2004 channels have been detected in boar and human that activates ion channels in sperm does not appear to involve PKA to any major extent: at 5 μmol/l the PKA inhibitor H-89 had little effect on isotonic volume, whereas similar concentrations of Bis I and lavendustin increased it markedly (Table 2). Moreover, in contrast to the cAMP effectors, H-89 produced no significant effects on volume regulation by endogenous protein kinase C or differential sensitivity to blockage by anti-ClC-3 antibodies (Yamamoto-Mizuma et al. 2004). It is therefore likely that the PKC-regulated volume-sensitive anion channels playing a role in sperm RVD are indeed members of the CLC family.

As a final comment, given the potential importance of volume regulatory control, boar spermatozoa appear to be an excellent model for experimental studies. In contrast to mouse spermatozoa, boar spermatozoa show characteristics similar to human; they are available in large numbers, and, of course, are routinely obtained as ejaculated cells (as are human), whereas mouse spermatozoa are routinely obtained as epididymal cells.

It remains unclear as to whether the various kinases, phosphatases and cAMP identified as being involved in volume regulation act on the same target protein or whether they act on different targets under different conditions, determined by osmolality or cell volume. Hougaard et al. (2004), for example, reported that isotonic volume was affected by PKC, PKA and Ca2+, but that these mechanisms were not responsible for increased activity during cell swelling. However, because isotonic sperm swelling induced by PKC inhibition was abolished in a medium containing sulfate instead of chloride, it appears more likely that PKC is involved in the closing of chloride channels both under isotonic and hypotonic conditions. In this case, the same target is addressed, but the cellular response is determined by internal and external ionic concentrations; this may account for the wide variability of individual response reported in our recent studies (Petrunkina et al. 2001a, 2001b, 2004a, 2005b). The effects of PP1 inhibitors were also consistent with a direct role in channel activation. However, as modulation of phosphotyrosine levels had only minor effects on RVD, it may well be that PTK and TP address a different target, such as intermediate regulation of PP activity (Cohen 1989).

The molecular identity of the channels, whose activities are modulated by the kinase, phosphatase and cAMP mechanisms revealed by our studies remains a matter of speculation and requires further investigation. It was recently demonstrated that volume-sensitive anion channels are responsible for chloride transport during sperm RVD (Petrunkina et al. 2004a, Yeung et al. 2005a). Volume-sensitive Cl− currents with similar functional properties have been reported in somatic cells (Shi et al. 2002); CLC-3 channels (voltage-gated chloride channels of the CLC family) in particular were shown to be regulated by endogenous PKC (Duan et al. 1999, Shi et al. 2002). Chloride channels have been detected in boar and human spermatozoa though not in mouse (Petrunkina et al. 2004a, Yeung et al. 2005a, 2005b). Despite apparent maintenance of RVD in salivary acinar cells isolated from mice with targeted disruption of the Clcn3 gene (Arreola et al. 2002), it has recently been shown that the properties of volume-sensitive anion channels in knockout mice are different from those in wild-type with respect to fundamental channel properties such as regulation by endogenous protein kinase C or differential sensitivity to blockage by anti-ClC-3 antibodies (Yamamoto-Mizuma et al. 2004). It is therefore likely that the PKC-regulated volume-sensitive anion channels playing a role in sperm RVD are indeed members of the CLC family.

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