Aquaporin 9 expression in the developing rat epididymis is modulated by steroid hormones

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Abstract

Fluid and solute transport across the epithelium of the male excurrent duct is important for sperm maturation and storage. Aquaporin 9 (AQP9), which allows permeation of water and neutral solutes, is abundant throughout the male reproductive tract, where it is expressed at the apical membrane of rat epididymal principal cells as early as at 1 week of age. We evaluated the effect of neonatal exposure to: 1) a GnRH antagonist (GnRHa); 2) diethylstilbestrol (DES); 3) ethinyl estradiol (EE); 4) DES plus testosterone (DES+TE); and 5) the anti-androgen flutamide on AQP9 expression in the epididymis of peripubertal rats. Control groups received the vehicle alone. In 25-day-old rats, quantification of the mean pixel intensity of immunofluorescence-stained sections showed a significant decrease in AQP9 staining in the apical membrane of epididymal principal cells after treatments with GnRHa, DES, or flutamide, compared to controls. These results were confirmed by western blotting. While EE induced a marked decrease in AQP9 levels by western blotting, the decrease in AQP9-associated fluorescence was not significant compared to controls. DES+TE-treated rats showed levels of AQP9 protein similar to controls, indicating maintenance of AQP9 expression by testosterone treatment in the presence of DES. Our data show that expression of AQP9 in the developing rat epididymis is downregulated by neonatal DES, GnRHa, EE, and flutamide, and that the effects mediated by estrogens can be prevented by testosterone administration.

Introduction

Interestingly, one of the solutes that can permeate through AQ9 is glycerol (Tsukaguchi et al. 1998), a spermatozoa metabolic substrate that accumulates in the lumen of the distal epididymis (Cooper & Brooks 1981). We have shown significant AQ9-dependent glycerol permeability in the apical membrane of epididymal epithelial cells (Pietrement et al. 2008). AQ9 was, thus, identified as a major apical AQ in the epididymal epithelium, where it could provide a route via which transepithelial fluid and solute transport could occur.

In the efferent ducts of the adult male excurrent duct, reabsorption of luminal fluid is regulated by both androgens and estrogens (Wong & Yeung 1978, Hess et al. 1997, Hess 2003, Ruz et al. 2006). We have shown that treatment of rodents and primates with diethylstilbestrol (DES) during the neonatal period strongly inhibits AQ1 expression in the efferent ducts of prepubertal animals (Fisher et al. 1998), indicating that maintaining the estrogen/androgen balance during postnatal development is crucial for the maturation of these segments.

In the epididymis, AQ9 regulation is less well documented. We have shown that AQ9 permeability is acutely regulated by luminal bradykinin (Belleannee et al. 2009). However, the role of sex hormones in regulating its expression has not been fully characterized. The promoter region of AQ9 contains a putative steroid hormone receptor-binding site (Tsukaguchi et al. 1998). In addition, sex-linked differences of AQ9 expression have previously been reported in the liver, suggesting that different hormonal profiles during and after development may play a role in the expression of this protein (Nicchia et al. 2001). We and others have previously shown that orchidectomy (Pastor-Soler et al. 2002, Oliveira et al. 2005), or treatment of adult male rats with the anti-androgen flutamide (Pastor-Soler et al. 2002), caused a significant reduction in the level of AQ9 protein expression in the adult epididymis. The downregulation of AQ9 expression in castrated animals was reversed by testosterone in the cauda epididymis (Pastor-Soler et al. 2002) and by dihydrotestosterone in the initial segments (Oliveira et al. 2005), indicating that androgens are important for AQ9 expression in the epididymis of adult animals. Androgen and estrogen (α and β) receptors are expressed in the epididymis during development and in adults (McKinnell et al. 2001, Zhou et al. 2002, Hess 2003). Neonatal exposure to the estrogenic compound DES significantly affects the morphology of the male efferent duct, including efferent ducts, epididymis, and vas deferens (McKinnell et al. 2001, Williams et al. 2001, Rivas et al. 2002, 2003, Goyal et al. 2003, Atanassova et al. 2005). Disturbances in the estrogen/androgen balance were proposed to be the key determinant in causing these adverse effects on male reproductive parameters. In addition, our previous study showed a marked reduction in the number of clear cells in the epididymis of prepubertal rats after neonatal exposure to DES (Fisher et al. 2002), a finding that further supports the notion that the epididymis is also susceptible to changes in the androgen/estrogen balance.

We have shown that AQ9 begins to appear in the apical membrane of epididymal principal cells as early as 1 week of age, and that its expression continues to increase throughout the postnatal period to reach the highest levels shortly before puberty (Pastor-Soler et al. 2001). This progressive increase in principal cell AQ9 expression during the prepubertal period was also confirmed by another laboratory (Badran & Hermo 2002). These findings suggest that initiation of AQ9 expression in the developing epididymis does not depend exclusively on the higher androgen levels that are triggered at puberty, and might require the participation of other hormones, including estrogens. The present study, therefore, assesses whether neonatal treatment with compounds known to affect the androgenic/estrogenic balance alters the expression of AQ9 in the epididymis in the peripubertal period. Understanding the effect of hormone manipulation during the neonatal period is crucial, because overexposure of young males to exogenous estrogens, including those from various environmental sources, can result in major abnormalities and reproductive dysfunction in adulthood (Atanassova et al. 1999, Goyal et al. 2003).

Results

Hormonal effects on epididymal AQ9 expression

Effect of GNRH antagonist, DES, ethinyl estradiol, and DES+ TE treatments

The expression of AQ9 was assessed by immunofluorescence staining of cryostat sections of the proximal cauda epididymidis, a region that shows high AQ9 expression. Control incubations were first performed using anti-AQ9 antibodies that had been preabsorbed with the AQ9 immunizing peptide. As we have previously described, AQ9 was localized to the apical membrane of principal cells in the epididymis of adult, as well as prepubertal rat epididymis (Fig. 1). This apical staining reached maximum intensity between the third and fourth postnatal weeks. Pre-incubation of the antibody with the AQ9 peptide completely abolished principal cell apical staining at all time points, demonstrating specificity of the antibody for AQ9.

In a separate cohort of animals, a strong AQ9 staining, comparable to that seen in adult animals, was observed in the apical membrane of principal cells of control 25-day-old rats (Fig. 2A and F). GNRH antagonist (GNRHa) and DES treatments during the neonatal period induced a marked decrease in AQ9 staining intensity, leading to an almost complete disappearance of staining in some tubules (Fig. 2B and C). Ethinyl estradiol (EE) also induced a reduction in AQ9 staining, although less
marked than the reduction observed with GNRHa or DES (Fig. 2D and G). Animals treated with DES together with testosterone showed AQP9 levels similar to that of control animals (Fig. 2E).

The level of AQP9 expression was quantified for each treatment by digital quantitative immunofluorescence labeling. Images from immunostained sections were acquired with a digital camera using identical parameters, and the mean pixel intensity (MPI) of the apical AQP9-associated fluorescence was quantified. For each rat, the MPI was normalized relative to the average control group value. Figure 3 shows the averaged normalized MPI for each animal group. A one-way ANOVA analysis revealed a highly significant effect of treatment ($P<0.0005$). A Tukey’s post hoc test showed marked reductions in AQP9 protein expression in the GNRHa- and DES-treated groups, compared to the control group ($P<0.0005$). The EE-treated rats showed a reduction in AQP9 MPI, although the effect was not statistically significant compared to controls ($P=0.149$). Animals treated with both DES and testosterone (DES + TE) showed significantly brighter AQP9 epididymal apical membrane staining compared to DES-treated animals ($P<0.0005$), with MPI values that were slightly, but not significantly, increased compared to controls ($P=0.479$).

**Effect of flutamide**

A separate cohort of rats was treated with the androgen receptor antagonist flutamide or with the vehicle during the neonatal period. Visual inspection of epididymis sections from 25 day-old rats showed a decrease in AQP9 immunofluorescence in the apical membrane of principal cells of the cauda epididymidis after flutamide treatment, compared to control (Fig. 4). This impression was confirmed by quantification of AQP9-associated MPI (Fig. 5), which revealed a significant decrease in AQP9 protein expression in the apical membrane of principal cells in the flutamide-treated group, compared to controls ($P<0.05$).

**Immunoblotting (SDS-PAGE and western blotting)**

Western blots were performed using total epididymal homogenates in order to confirm the effects of the different hormonal treatment in the expression of AQP9 expression seen by immunofluorescence labeling (Fig. 6). As we have previously described, the affinity-purified AQP9 antibody detected a main band at an apparent molecular weight of 30–35 kDa, as well as additional higher molecular weight bands in total homogenates of epididymis (caput, corpus, and cauda) of control animals (Pastor-Soler et al. 2001, 2002, Pietrement et al. 2008). No AQP9 bands were detected in lanes containing the same amount of protein from total epididymal extracts of animals treated with GNRHa, DES, and EE. The DES + TE group showed a stronger signal for AQP9 compared to the DES-treated animals, indicating maintenance of AQP9 protein expression by testosterone in the presence of DES. A very faint AQP9 band was observed in flutamide-treated animals.

**Discussion**

Previous studies have shown that AQP9 can be detected in the developing excurrent ducts as early as 1 week postnatally, indicating that its expression does not strictly depend on the high androgen levels that are triggered during puberty. Accordingly, either the low androgen levels found in prepubertal males or other factors were proposed to be involved in AQP9 expression. The present study shows that AQP9 expression in principal cells of peripubertal rats, as quantified by immunofluorescence and visualized by western blotting, is greatly decreased following perinatal exposure to either the GNRHa, antarelix, or the estrogenic compound, DES. Exposure of neonatal rats to EE also decreased the level of expression of AQP9 in epididymal homogenates by...
western blot. However, the decrease that we observed by immunofluorescence staining failed to reach statistical significance in EE-treated animals, probably reflecting the different sensitivities of the two techniques. Quantification of AQP9 expression by immunofluorescence was restricted to the proximal cauda epididymidis, a region that expresses high levels of AQP9, while assessment of AQP9 expression by western blotting was performed on whole epididymis extracts. Thus, it remains possible that other epididymal regions were more strongly affected by EE, leading to the marked reduction in AQP9 signal observed by western blotting compared to our immunofluorescence measurements of the cauda region. The same cohort of animals showed a marked reduction in testis weight and plasma testosterone levels after treatment with GNRHa, DES, and EE (Fisher et al. 2002). In addition, expression of AR was almost completely inhibited by DES, but it was not affected by GNRHa (McKinnell et al. 2001). Both aER (ESR1) and bER (ESR2) expression remained intact following these treatments (McKinnell et al. 2001). Thus, the reduced expression of AQP9 that we observed here following DES exposure could have been attributed to either inhibition of androgen activation, increase in estrogen stimulation, or both.

Flutamide alone prevented the induction of AQP9 in neonatal rats (this study) and significantly reduced AQP9 levels in adult rats (Pastor-Soler et al. 2002) showing the androgen requirement of AQP9 expression in the epididymis. Flutamide did not affect testis weight or plasma testosterone levels in the same animals (Fisher et al. 2002), showing that it did not suppress the hypothalamo-pituitary axis, and further indicating that direct inhibition of epididymal androgen action is sufficient to inhibit AQP9 expression. In addition, when testosterone was administered together with DES, AQP9 expression was maintained at control levels. Altogether, these findings suggest that the effects elicited by DES could be solely attributed to a marked inhibition of androgen stimulation secondary to elevation of estrogen levels.

The fact that normal AQP9 expression necessitates androgen stimulation implies that, during normal postnatal development, the lower levels of circulating androgens in prepubertal male rats compared to adults are essential and sufficient to trigger the progressive expression of AQP9 that was previously observed between postnatal weeks 1 and 4 (Pastor-Soler et al. 2001, Badran & Hermo 2002). The developmental pattern of expression of AQP9 in principal cells mimics that of V-ATPase in clear cells in the epididymis (Breton et al. 1999). The effects elicited by GNRHa, DES, EE, and flutamide on principal cell AQP9 expression also correlate with the marked decrease that we observed...
Steroid hormones modulate epididymal AQP9

Figure 3 Quantification of AQP9-associated fluorescence staining in the cauda epididymis of 25-day-old control and treated rats (groups represented in Fig. 1). Significant decreases in AQP9 staining intensity were induced by GNRHa and DES. EE also induced a reduction in AQP9 staining but the difference was not significant versus control. Co-administration of testosterone with DES completely restored AQP9 staining compared to DES alone, with AQP9 staining intensity comparable to control. Data are expressed relative to control, and represent the means ± S.E.M. obtained from 4 to 7 animals per group. **P < 0.0005 versus control; **P < 0.005 versus DES; NS, not significant versus control.

Figure 4 Downregulation of AQP9 expression in the cauda epididymis of 25-day-old rats by the anti-androgen flutamide. Immunofluorescence for AQP9 in the cauda epididymis from a control rat (A) and a flutamide-treated rat (B) showing a marked decrease in apical AQP9 staining in the flutamide-treated animal. Panels C and D are higher magnification representations of some of the tubules shown in A and B respectively. Digital images were acquired using identical parameters. Bars = 25 μm.

In conclusion, this study emphasizes the role of steroid hormones in the establishment and maintenance of AQP9 in the epididymis, and further illustrates the importance of monitoring environmental factors that affect hormonal balance as potential mediators of male fertility.
Materials and Methods

Animals and treatments

All the animal studies were performed under license from and in accordance with the legal requirements of the UK Home Office. Wistar male rats received postnatal treatments (day of birth is assigned as day 1). Irrespective of the treatment received, pups were weaned at 22 days and fed on standard rat breeding diet no. 3 (SDS, Dundee, Scotland). All animals in this study were killed on day 25, a time at which maximal AQP9 expression was reached in nontreated rats (Pastor-Soler et al. 2001, Da Silva et al. 2006b).

Neonatal rats treated with androgens and/or estrogens

The first group of animals had been treated previously with modulators of androgens and estrogens, and were characterized for the efficacy of each treatment (Fisher et al. 2002). Archival epididymis sections were used in the present study. These animals received one of the following s.c. treatments:

1) Control group (n=6): 20 μl corn oil administered on alternate days (2–12 inclusive)
2) GNRHa group (n=6): 10 mg/kg of a long acting GNRHa (Antarelix; Europeptides, Argenteuil, France) in 5% mannitol administered on postnatal days 2 and 5.
3) DES group (Sigma Chemical Company) (n=4): 10 μg in 20 μl corn oil administered on alternate days (2–12 inclusive).
4) Ethinyl estradiol (EE group; Sigma) (n=8): 10 μg in 20 μl corn oil administered on alternate days (2–12 inclusive).
5) DES + testosterone (DES + TE group) (n=5): 10 μg DES co-administered with 200 μg testosterone esters (Sustanon, Organon Labs, Cambridge, UK) in 20 μl corn oil on alternate days (2–12 inclusive).

Flutamide-treated neonatal rats

A second series of animals had also been treated and characterized previously (Fisher et al. 2002). They received s.c. injections of either:

1) Flutamide (androgen receptor antagonist, Sigma) (n=3): 50 mg/kg in 20 μl of corn oil on alternate days (2–12 inclusive) or
2) Corn oil (20 μl) administered on alternate days from day 2 to 12 inclusive (control group; n=3).

Testosterone levels measured at postnatal day 25 were significantly reduced in the GNRHa, DES, and EE groups (Fisher et al. 2002). In addition, a significant testis weight reduction (>85%) was observed in the GNRHa, DES, EE, and DES + TE groups, demonstrating the effectiveness of the respective treatments in retarding testicular development (Fisher et al. 2002). In contrast, neonatal administration of flutamide was without significant effects on plasma testosterone levels and testicular weight.

Tissue fixation and immunocytochemistry

Tissue preparation

Rats were killed by inhalation of CO₂ followed by cervical dislocation. The right epididymis from each animal was fixed by immersion in periodate-lysine-paraformaldehyde (PLP) containing 2% paraformaldehyde for 5 h at room temperature. The left epididymis was snap frozen and stored at −80°C.
prior to protein extraction. The right testis was weighed after necropsy, and their weights are reported in our previous publication (Fisher et al. 2002).

Antibodies

An affinity-purified rabbit primary antibody against AQ99 was used for western blotting and immunofluorescence labeling. This antibody was raised against a C-terminal peptide from rat AQ99 (PSENNLEKHEL5VIM-C) and has been previously characterized (Pastor-Soler et al. 2001, 2002, Da Silva et al. 2006b, Pietrement et al. 2008).

Immunofluorescence labelling

PLP-fixed tissues were cryoprotected in a solution of 30% sucrose in PBS. Tissues were embedded in OCT compound (Tissue-Tek; Sakura, Finetek USA, Torrance, CA, USA), mounted on a cutting block, and frozen in a Reichert Frigocut or a Leica 3050 cryotome (spencer Scientific). The tissue was then cut at 4 µm thickness, and sections were placed onto Fisher Superfrost Plus microscope slides (Fisher Scientific, Pittsburgh, PA, USA). After rehydration in PBS at room temperature, all tissue slides were pretreated with 1% SDS for 4 min, as previously described (Brown et al. 1996). The slides were then washed in PBS (3 × 5 min) and pre-incubated in 1% BSA in PBS/0.02% sodium azide for 15 min at room temperature to block nonspecific staining. The anti-AQP9 antibody was prepared at a dilution of 1:3200 in antibody diluent (DAKO, Carpinteria, CA, USA) and applied for 75 min at room temperature. The slides were then washed twice for 5 min in high salt PBS (2.7% NaCl) to reduce nonspecific staining, and once in normal PBS. Some slides were also incubated with anti-AQP9 antibody that had been premixed with a tenfold excess of the immunizing peptide (PSENNLEKHEL5VIM-C). The secondary antibody (a goat anti-rabbit IgG coupled to CY3) was applied for 1 h at room temperature. The slides were then incubated with anti-AQP9 antibody that had been premixed with a tenfold excess of the immunizing peptide (PSENNLEKHEL5VIM-C). The tissue was left on ice for 4 min, as previously described (Brown et al. 1996). The slides were then cut at 4 µm thickness, and sections were placed onto Fisher Superfrost Plus microscope slides (Fisher Scientific, Pittsburgh, PA, USA). After rehydration in PBS at room temperature, all tissue slides were pretreated with 1% SDS for 4 min, as previously described (Brown et al. 1996). The slides were then washed in PBS (3 × 5 min) and pre-incubated in 1% BSA in PBS/0.02% sodium azide for 15 min at room temperature to block nonspecific staining. The anti-AQP9 antibody was prepared at a dilution of 1:3200 in antibody diluent (DAKO, Carpinteria, CA, USA) and applied for 75 min at room temperature. The slides were then washed twice for 5 min in high salt PBS (2.7% NaCl) to reduce nonspecific staining, and once in normal PBS. Some slides were also incubated with anti-AQP9 antibody that had been premixed with a tenfold excess of the immunizing peptide (PSENNLEKHEL5VIM-C). The secondary antibody (a goat anti-rabbit IgG coupled to CY3) was applied for 1 h at room temperature. The slides were then incubated with anti-AQP9 antibody that had been premixed with a tenfold excess of the immunizing peptide (PSENNLEKHEL5VIM-C). The tissue was left on ice for 4 min, as previously described (Brown et al. 1996). The slides were then washed twice for 5 min in high salt PBS (2.7% NaCl) to reduce nonspecific staining, and once in normal PBS. Some slides were also incubated with anti-AQP9 antibody that had been premixed with a tenfold excess of the immunizing peptide (PSENNLEKHEL5VIM-C). The secondary antibody (a goat anti-rabbit IgG coupled to CY3) was applied for 1 h at room temperature. The slides were then incubated with anti-AQP9 antibody that had been premixed with a tenfold excess of the immunizing peptide (PSENNLEKHEL5VIM-C). The tissue was left on ice for 4 min, as previously described (Brown et al. 1996). The slides were then washed twice for 5 min in high salt PBS (2.7% NaCl) to reduce nonspecific staining, and once in normal PBS. Some slides were also incubated with anti-AQP9 antibody that had been premixed with a tenfold excess of the immunizing peptide (PSENNLEKHEL5VIM-C). The secondary antibody (a goat anti-rabbit IgG coupled to CY3) was applied for 1 h at room temperature. The slides were then incubated with anti-AQP9 antibody that had been premixed with a tenfold excess of the immunizing peptide (PSENNLEKHEL5VIM-C). The tissue was left on ice for 4 min, as previously described (Brown et al. 1996).

Quantification of AQ99 expression by immunofluorescence labeling

Immunofluorescence labeling of each tissue was performed in at least three independent incubations, each incubation including slides from all treatment groups. All slides for a particular incubation were treated under identical conditions, and digital images were obtained using identical acquisition parameters. Each image was corrected for its own luminal unstained background value, and the MPI of apical AQ99-associated fluorescence was measured using IPLab Spectrum software. The segmentation function of IPLab spectrum was used to set the minimum pixel intensity for inclusion in the quantification so that the selected pixels corresponded to the brush border membrane area of principal cells observed in the initial, non-manipulated image (see Results). We have previously used this method to quantify the downregulation of epididymal AQ99 expression induced by flutamide in adult rats (Pastor-Soler et al. 2002).

Statistical analysis

GNRHa, DES, EE, and DES+ TE treatments

Overall, 28 rats were treated in the first experimental setting (control rats and rats treated with GNRHa, DES, EE, and DES+TE). In order to minimize procedural variations, 15 epididymis slides including three animals from each group were treated per incubation. Thus, the epididymis from one given rat was sampled multiple times. The MPI of AQ99-associated fluorescence for each rat was normalized to the average control group value for the corresponding incubation. The purpose of this normalization procedure was to correct for any potential variations that might occur between incubations. All normalized values for each rat were averaged and analyzed. A one-way ANOVA was conducted to determine whether the treatment groups differed. A Tukey’s post hoc test was then conducted to determine which group(s) was/were significantly different from one another. The mean difference was considered significant at P=0.05.

Flutamide treatment

In this experimental setting, statistical significance of the flutamide effect on the AQ99-associated MPI was compared to its own control group, using a one-tailed Student’s t-test at the 0.05 level of significance.

Protein extraction and immunoblotting (SDS-PAGE and western blotting)

Protein extraction

For each treatment group, whole epididymides of 25-day-old rats were powdered in a porcelain mortar under liquid nitrogen, and the powder was stored on dry ice. Protein was extracted by the addition of 200 ml cold extraction buffer containing 10 mM HEPES, pH 7.9; 0.1 mM EGTA; 1 mM dithiothreitol; 0.5 mM phenylmethylsulphonyl fluoride; and Complete protease inhibitors (Roche). The tissue was left on ice for 15 min prior to the addition of 25 ml of 10% Nonidet P-40 (Sigma) to the tube, which was then vortexed thrice for 10 s each. After centrifugation at 12 000 g for 1 min at 4 °C, the supernatant was decanted, and 100 mg aliquots were frozen on dry ice before being stored at −40 °C.

Western blot analysis

Protein samples (from all treatment groups) and protein markers (Bio-Rad) were separated by SDS-PAGE using 4–20% gradient gels (Invitrogen). The gels were run at 110 V for 180 min in Tris/glycine transfer buffer (Invitrogen). The final images were imported into and printed from Adobe Photoshop.
Membranes were blocked with 5% milk in TBS for 1 h at room temperature, and then washed in TBST (Tris-buffered saline; 50 mM Tris–HCl, 150 mM NaCl containing 0.05% Tween-20; Sigma). The membranes were incubated with the AQP9 antibody, diluted 1:2000 in TBS/0.02% sodium azide at 4 °C overnight. After extensive washes in TBST, a goat anti-rabbit IgG conjugated to HRP (Sigma) was applied to membranes at a dilution of 1:10 000. After washing, AQP9 was detected using the ECL kit (Amersham), and membranes were exposed to film (Kodak) until optimal development of the signal was detected.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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