Oocyte development in bovine primordial follicles is promoted by activin and FSH within a two-step serum-free culture system

Marie McLaughlin and Evelyn E Telfer

Centre for Integrative Physiology, University of Edinburgh, Hugh Robson Building, George Square, Edinburgh EH8 9XD, UK

Correspondence should be addressed to E E Telfer; Email: evelyn.telfer@ed.ac.uk

Abstract

Quiescent follicles of large mammals initiate growth within cultured pieces of ovarian cortex. Systems capable of sustaining in vitro development from this early stage until oocyte maturation would allow investigation of mechanisms regulating oocyte development in its entirety. The aims of this study were 1) to determine whether bovine follicles initiated to grow in vitro could be isolated from the cortical environment, and could undergo further development and 2) to evaluate the effect of activin and FSH on the development of secondary follicles derived from primordial follicles. Fragments of bovine ovarian cortex were cultured in serum-free medium for 6 days; thereafter, secondary follicles were isolated for further culture. After a maximum total of 21 days in vitro, follicles were either processed for histological assessment or opened to release the oocyte–cumulus complexes for inspection by light microscopy. Compared with control, significant follicle and oocyte growth were observed in activin-exposed follicles, with or without FSH, with some oocyte diameters measuring over 100 microns following a total in vitro period of 15 days. Significant oestradiol secretion was observed in follicles cultured in activin alone after a total of 9 days in vitro compared with other treatment groups; however, this effect was not sustained. In summary, this study demonstrates the promotion of primordial bovine follicle development within a two-step serum-free culture system with oocyte diameters > 100 μm achieved over 15 days in vitro. Further development of this system is needed to support complete oocyte growth and thereafter in vitro maturation.

Reproduction (2010) 139 971–978

Introduction

Current understanding of the mechanisms underpinning oocyte development in immature follicles is incomplete. The ability to support the development of primordial bovine follicles through to acquisition of oocyte competency in vitro would not only advance the understanding of germ cell development in non-rodent species, but could also provide mature oocytes for assisted reproductive technologies in species understood to require a prolonged follicular growth period.


Multi-step systems have already been designed to support in vitro follicle development in mice with the birth of pups from in vitro matured oocytes derived from cumulus–oocyte complexes (Eppig & Schroeder 1989), and primordial (Eppig & O’Brien 1996) and primary follicles (Spears et al. 1994); however, to date, it has not been possible to repeat this success in large mammals using oocytes from such immature follicles. A major obstacle to progress in large species is oocyte degeneration associated with loss of oocyte and somatic...
cell contact particularly over a prolonged culture period; therefore, it is imperative that an understanding of the impact of culture conditions on the maintenance of intercellular associations is realised.

In the following experiment, a two-step culture system was designed to promote the activation of follicle growth within fragments of ovarian cortex followed by isolation and individual culture of secondary follicles. The transforming growth factor-β superfamily member activin, previously associated with the maintenance of oocyte–somatic cell adhesion and a recognised promoter of preantral follicle development (Hulshof et al. 1997, Liu et al. 1998, Zhao et al. 2001, Thomas et al. 2003, Silva et al. 2006, Telfer et al. 2008), and FSH were chosen as culture medium supplements. Activation and early follicle growth occur in the absence of gonadotrophins (Peters et al. 1973, McNatty et al. 1990). Development beyond the early antral stage is critically dependent on the presence of FSH (Oktay et al. 1998); however, the precise effect of FSH on preantral follicle development is unclear. It has been demonstrated that FSH can augment the action of activin, which in turn simulates FSH receptor expression in cultured murine granulosa cells (Xiao et al. 1992), confirming a relationship between the follicular actions of FSH and activin. In this experiment, the effect of activin and activin in combination with FSH on the development of secondary bovine follicles grown in vitro was assessed with regard to follicle and oocyte growth, steroidogenesis and maintenance of normal oocyte morphology.

Results

Follicle growth and differentiation

To confirm the developmental stage of follicles in fresh tissue, a total of 12 fragments were randomly selected prior to culture, fixed in Bouin’s solutions and processed as described for histological assessment. The mean distribution of follicles in freshly fixed tissue (day 0) was compared to that of ten fragments cultured for 6 days. Follicles were categorised by a modification of the system used by Wandji et al. (1996) as follows: 1) quiescent follicles: oocytes surrounded by a complete or incomplete single layer of cells, either all flattened (primordial) or a mixed layer of flattened and cuboidal cells (transitory), 2) primary follicles: follicles comprising an oocyte surrounded by a single complete layer of cuboidal granulosa cells, and 3) secondary follicles: multilaminar follicles with more than a complete single layer of granulosa cells. Follicle survival was high in cultured tissue fragments, with <10% follicle degeneration observed in any of the treatment groups. As shown in Fig. 1i, a significantly greater percentage (P<0.01) of growing follicles (primary) and a corresponding significant decrease in the percentage of quiescent follicles (P<0.01) were observed in cultured fragments compared with freshly fixed tissue, which contained predominantly quiescent follicles (Fig. 1ii). Secondary follicles (mean diameter 111±1.46 μm) were dissected from cultured cortical strips (Fig. 1iii and iv), and grown for 6 days in control medium (n=31) or in medium supplemented with 100 ng/ml recombinant human activin A (rhAct A) alone (n=70) or 100 ng/ml rhFSH in combination with 50 ng/ml recombinant human FSH (rhFSH; n=57). Significant growth occurred in follicles supplemented with 100 ng/ml rhAct A in the presence (P<0.01) or absence (P<0.01) of 50 ng/ml rhFSH compared with control over a 12-day in vitro period. Activin-exposed follicles were cultured for a maximum of 21 days; due to poorer growth rates, control follicles were not cultured beyond 12 days. Follicle growth increased significantly at days 15 and 18 in the presence of 100 ng/ml rhAct A and 50 ng/ml rhFSH compared with activin alone (P<0.05); however, by day 21 of the

Figure 1 Effect of culture on the quiescent and growing follicle population in bovine cortical fragments. (i) a, b, c and d indicate differences between quiescent and primary follicle populations. A significant decrease (P<0.01) in the percentage of quiescent follicles was observed after 6 days in culture with a corresponding increase in the percentage of primary follicles (P<0.01). No secondary follicles were observed in freshly fixed (day 0) tissue which contained predominantly quiescent follicles (ii). Secondary follicles were dissected from strips of bovine cortex following 6 days in vitro (iii) and (iv), bar=100 μm and (iv), bar=25 μm.
in vitro period, mean follicle diameters in both groups had declined sharply (Fig. 2i). The presence of an antrum was determined on histological inspection (Fig. 2ii), though occasionally cavity formation could be seen by light microscopy during the in vitro period (Fig. 2iii). Significant antral formation was observed in follicles cultured in 100 ng/ml rhAct A alone ($P < 0.05$) or 100 ng/ml rhAct A and 50 ng/ml rhFSH ($P < 0.05$) compared with follicles cultured in control, with approximately a third of the follicles in both activin-exposed groups forming cavities.

**Oocyte growth and cumulus complex morphology**

Follicles grown in vitro were isolated and cultured for up to 21 days in the presence or absence of 100 ng/ml rhAct A, or in the presence of 100 ng/ml rhAct A in combination with 50 ng/ml rhFSH. As shown in Fig. 3i, significant oocyte growth occurred over 12 days in isolated secondary follicles cultured in the presence of 100 ng/ml rhAct A with or without 50 ng/ml rhFSH compared with oocytes within cortical fragments cultured for 6 days only ($P < 0.01$) and isolated secondary follicles cultured in control medium ($P < 0.05$). The normality of oocyte morphology was assessed by determining the general circularity of the oocyte and the integrity of the oolemma and zona pellucida, and the presence of a germinal vesicle and nucleolus in each section was evaluated as illustrated in Fig. 3ii and iii. Therefore, grossly spherical oocytes with an intact oolemma, distinguishable zona pellucida, a visible germinal vesicle and defined nucleolus were considered morphologically normal. Oocytes not meeting these criteria were deemed abnormal. After 12 days

![Figure 2](image1.png)

**Figure 2** Effect of rhAct A and FSH on follicle development over 21 days in vitro. (i) Values are mean follicle diameters ± S.E.M. * and ** indicate significant differences between treatments. Mean follicle diameters were greater in the presence of 100 ng/ml rhAct A and 50 ng/ml FSH (open circles) on days 15 ($P < 0.05$) and 18 ($P < 0.05$) compared with 100 ng/ml rhAct A alone (closed circles). Follicles cultured in control medium for a total of 12 days were significantly smaller than those cultured in the presence of activin over the same time period (not shown). (ii) Antral cavity formation in a follicle cultured in the presence of 100 ng/ml rhAct A and 50 ng/ml FSH for 15 days; bar = 50 μm. (iii) Formation of antra could occasionally be seen during the in vitro period; bar = 100 μm.

![Figure 3](image2.png)

**Figure 3** Effect of rhAct A and FSH on oocyte growth and morphology over 12 days in vitro. (i) Values are mean oocyte diameters ± S.E.M.; a, b and c indicate differences between treatments. Mean oocyte diameters were significantly greater in the presence of 100 ng/ml rhAct A with or without 50 ng/ml rhFSH compared with the diameters of oocytes cultured in control medium ($P < 0.05$) and with those of oocytes of follicles cultured in strips for 6 days only ($P < 0.01$). (ii) Morphologically normal oocyte grown in vitro in the presence of 100 ng/ml rhAct A alone for 12 days; bar = 25 μm. (iii) Histological image of morphologically normal in vitro grown oocyte cultured in the presence of 100 ng/ml rhAct A and 50 ng/ml rhFSH for 12 days; bar = 50 μm.
**in vitro**, no significant difference in oocyte morphology was observed between the treatment groups (control \( n = 31 \), activin \( n = 20 \), and activin and FSH \( n = 20 \), with 55–65% of all the oocytes cultured appearing morphologically normal. Oocyte–cumulus complexes were released from follicles cultured in 100 ng/ml rhAct A with or without 50 ng/ml rhFSH on days 15 (activin \( n = 15 \), and activin and FSH \( n = 15 \)), 18 (activin \( n = 10 \), and activin and FSH \( n = 8 \)) and 21 (activin \( n = 5 \), and activin and FSH \( n = 6 \)) of the total *in vitro* period, and the oocyte complexes were measured and assessed for the presence of adherent cumulus cells and an intact zona pellucida. Follicle degeneration increased steeply with time. At day 15, <5% of follicles cultured in the presence of activin alone and <7% of those cultured in activin combined with FSH were degenerate, whereas by day 18, this had risen to 32 and 39% respectively. By day 21, almost 50% of the oocytes from both treatment groups were degenerate. As shown in Fig. 4, maximum oocyte diameters (>90.1 \( \mu m \)) were achieved in follicles cultured for a total of 15 days in the presence of activin with or without FSH. A significantly greater proportion of oocytes had adherent cumulus cells when FSH was present in the culture medium \( (P<0.05; \text{Fig. 5i}) \) compared with oocytes cultured in activin alone at days 15 and 18; however, by day 21, nearly all the oocytes from both treatment groups were denuded on release \( (\text{Fig. 5ii}) \). Zona pellucidae were present and intact in all oocytes released from follicles on days 15, 18 and 21; however, by day 21, oocytes from both treatments appeared grossly misshapen \( (\text{Fig. 5iii}) \).

**Oestradiol secretion**

Media from follicles cultured individually in control medium \( (\text{day 9}, \ n = 7; \text{day 12}, \ n = 7) \), activin alone \( (\text{day 9}, \ n = 9; \text{day 12}, \ n = 9) \) or activin with FSH \( (\text{day 9}, \ n = 7; \text{day 12}, \ n = 7) \) were analysed for oestradiol (OE2) content by immunoenzyme assay. Significant OE2 secretion was observed in follicles cultured in activin alone \( (P<0.05) \) compared with the other treatment groups; however, this effect was not sustained, and by day 12, no significant difference was detected between treatments \((\text{Table 1})\).

**Discussion**

If mechanisms regulating germ cell development in non-rodent mammals are to be fully understood and competent oocytes are to be routinely produced *in vitro*, systems supporting the growth of large mammalian follicles from the earliest stages are a prerequisite. Although major milestones in bovine follicle growth have been realised *in vitro*, i.e. primordial follicle activation \( (\text{Wandji et al. 1996, Fortune et al. 1998, Gigli et al. 2006, Yang & Fortune 2006, 2007}) \), preantral follicle growth \( (\text{Wandji et al. 1996, Hulshof et al. 1997}) \),
Table 1 Oestradiol secretion in bovine follicles activated and grown in vitro.

<table>
<thead>
<tr>
<th>Mean oestradiol secretion (pg/ml)</th>
<th>Day 9</th>
<th>Day 12</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>28.45</td>
<td>32.61</td>
</tr>
<tr>
<td>100 ng/ml rhAct</td>
<td>41.31*</td>
<td>35.28</td>
</tr>
<tr>
<td>100 ng/ml rhAct + 50 ng/ml rFSH</td>
<td>30.78</td>
<td>33.33</td>
</tr>
</tbody>
</table>

*Denotes a significant difference between treatment groups at day 9 \((P<0.05)\).

In this study, activin with or without FSH promoted follicle growth up to 18 days in vitro, and the formation of antral cavities in cultured secondary follicles was observed after 12 days. This demonstrates a markedly enhanced rate of development, as it is estimated that in vivo bovine primordial follicles take several months to reach the antral stage (Lussier et al. 1987, Gosden et al. 1994). Previous studies have achieved antral cavity formation after 7 and 10 days in vitro (Gutierrez et al. 2000, Itoh et al. 2002); however, both studies cultured larger preantral follicles (166 ± 2.15 μm). Activin and activin with FSH promoted follicle growth with follicle diameters increasing from 111 up to 200 μm (activin alone) and 247 μm (with FSH) with concomitant increase in oocyte diameters from 39 μm to a maximum of 107 μm over 15 days in vitro. Normal oocyte morphology was maintained in the presence of activin with or without FSH; however, their competency is yet to be tested. Hirao et al. (2004) successfully cultured, matured and fertilised bovine oocyte–cumulus complexes in vitro with a mean oocyte diameter of 95 μm on aspiration (Hirao et al. 2004). In this study, the largest oocytes were observed on day 15; moreover, in the presence of 100 ng/ml rhAct A and 50 ng/ml rFSH, oocytes retained adherent cumulus cells until day 18, indicating that a close association between the germ and somatic cells was being maintained by culture conditions; however, extending the culture to 21 days resulted in oocyte degeneration. It is well established that maintenance of bidirectional communication between the oocyte and its surrounding somatic cells is vital to the production of a healthy oocyte (Albertini et al. 2001, Eppig 2001). Our results support the idea that to facilitate complete oocyte development in vitro, an additional culture step would need to be introduced to support the development of oocytes outwith the large antral follicle (Telfer & McLaughlin 2007).

Follicles supplemented with 100 ng/ml rhAct A alone secreted significant O₂, which together with antral formation is indicative of follicle differentiation. Increased O₂ peaked at day 9, and was not maintained throughout the culture period; however, the reduced steroidogenesis observed at day 12 did not have a deleterious effect on oocyte growth or morphology.

In summary, this study demonstrates the potential of secondary bovine follicles derived from primordial follicles activated in vitro to support continued oocyte development within isolated follicles. The presence of activin and FSH enhanced follicle and oocyte growth in vitro, with some follicles containing oocytes with diameters >100 μm after a 15-day culture period. These results provide another encouraging step towards the realisation of complete in vitro growth of large mammalian oocytes.

Materials and Methods

Tissue preparation and fragment culture

Bovine ovaries from freshly killed heifers were transported from the local abattoir at 33–38 °C in HEPES-buffered M199 media (Invitrogen Ltd) supplemented with amphotericin B (2.5 μg/ml; Invitrogen Ltd), pyruvic acid (25 μg/ml), penicillin G (75 μg/ml) and streptomycin (50 μg/ml; all chemicals obtained from Sigma Chemicals). Under laminar flow conditions, ovaries were rinsed in 70% alcohol, and fine cortical strips were removed using a scalpel. The strips were placed in a dissection medium (Leibovitz medium, Invitrogen Ltd) supplemented with sodium pyruvate (2 mM), glutamine (2 mM; both obtained from Invitrogen Ltd), BSA (Fraction V, 3 mg/ml), penicillin G (75 μg/ml) and streptomycin (50 μg/ml; Sigma Chemicals). Under light microscopy, any damaged, haemorrhaged or excess stromal tissue was excised, allowing the pieces to flatten and enhancing microscopic visualisation. The flat tissue pieces were dissected into fragments of between 0.5 and
1 mm², and re-examined under light microscopy, and all visible follicles were measured. Any follicle of mean diameter > 40 µm was excised from the tissue fragments to ensure that the presumptive population of follicles was unilaminar, i.e. primordial or transitory. Seven replicate cultures were established with 12–33 cortical fragments in each. The fragments were cultured individually in 24-well cell culture plates (Corning B.V. Life Sciences Europe, Amsterdam, The Netherlands) containing McCoy's 5a medium with bicarbonate supplemented with HEPES (20 mM; Invitrogen Ltd), glutamine (3 mM; Invitrogen Ltd), BSA (Fraction V 0.1%), penicillin G (0.1 mg/ml), streptomycin (0.1 mg/ml), transferrin (2.5 µg/ml), selenium (4 ng/ml), insulin (10 ng/ml) and ascorbic acid (50 µg/ml); all obtained from Sigma Chemicals). Fragments were cultured for 6 days at 37 °C in humidified air with 5% CO₂, with half the media being removed and replaced every second day. A number of cortical fragments were randomly selected, and fixed on each day a culture was established (day 0) to allow comparison between cultured and in vivo follicular parameters.

**Follicle isolation and culture**

After 6 days, a number of cortical fragments were removed from each culture and fixed in Bouin's solution prior to secondary follicle dissection to allow comparison of follicle growth between day 0 and 6 (total n = 10). The remaining cortical fragments (n = 233) were transferred to pre-warmed Leibovitz dissection medium supplemented as described earlier, and preantral follicles (111 ± 1.46 µm) were mechanically dissected with 25-gauge needles, and those with an intact basement membrane and no antral cavity were selected for culture. A total of 170 follicles were dissected, 12 follicles were fixed immediately as day 6 follicle controls, and 158 follicles were randomly assigned to treatment groups. Selected follicles were individually placed in 96-well V-bottomed culture plates (Corning Costar Europe, Badhoevedorp, The Netherlands) in 150 µl of the culture medium. For the control group (n = 31), follicles were cultured in McCoy's 5a medium supplemented as described earlier. For the treatment groups, follicles were cultured in McCoy's 5a medium as described before with either 100 ng/ml rhAct A (R&D Systems, Abingdon, UK; n = 70) or 100 ng/ml rhAct A in combination with 50 ng/ml rhFSH (Sigma Chemicals; n = 57). The follicles were cultured individually for a maximum of 15 days at 37 °C in humidified air with 5% CO₂, and on every third day, half the culture medium was removed and replaced concomitant with follicle diameter measurement using a dissecting microscope with a crossed micrometer. Upon microscopic inspection, any damaged or degenerate follicles were excluded from the study. The prevalence of follicle deterioration was not significantly different between the treatment groups.

**Assessment of histology**

A total of 12 fragments of fresh cortical tissue and ten fragments of tissue cultured for 6 days were fixed for 24 h in Bouin's solution and processed for histological inspection as described below. The developmental stage of the follicles that were present in the tissue was assessed by light microscopy, and the proportions of quiescent and growing follicles were compared between the groups. After a total of 12 days in vitro (6 days within cortical fragments and 6 days in individual follicle culture), 20 follicles each from both activin-supplemented treatment groups and all control-cultured follicles (n = 30) were fixed in Bouin's solution for 24 h, and were then dehydrated in an eosin/alcohol solution in increasing concentrations (70, 90 and 100%). Following dehydration, the follicles were immersed in cedarwood oil (BDH Laboratory Supplies, Poole, UK) for 24 h and were then placed in toluene (Fisher Scientific UK Ltd, Loughborough, UK) for 30 min to remove all traces of the oil. The follicles were then embedded in paraffin wax at 60 °C for 4 h, with the wax being renewed every hour to ensure complete clearance of all the toluene. Six-micrometre-thick sections were sectioned, mounted on gelatine-coated slides and left to dry overnight prior to staining with haematoxylin and eosin. Using a light microscope with a crossed micrometer, sections of follicles containing the nucleolus were assessed, and the following parameters were evaluated: follicle and oocyte diameter, antral cavity formation and oocyte integrity. In total, 1–2 follicles from each activin-treated group cultured beyond a total of 12 days were fixed and processed as described above; the remaining follicles from these groups cultured for more than 12 days were assessed as detailed below.

**Assessment of oocyte–cumulus complexes**

A number of follicles cultured in 100 ng/ml rhAct A with (n = 35) or without (n = 45) 50 ng/ml rhFSH remained in vitro for a maximum total period of 21 days. Following 15, 18 and 21 days in vitro, a number of follicles from each treatment group were removed to preheated Leibovitz dissection media supplemented as described earlier, and were gently opened using 25-gauge needles. Using a light microscope with a crossed micrometer, the released oocyte complexes were inspected, and oocyte diameter, zona pellucida integrity, cumulus cell adherence and oocyte morphology were evaluated.

**Measurement of OE₂ by enzyme immunoassay**

To evaluate follicular steroidogenesis in vitro, concentrations of OE₂ in reserved culture media were measured against standard dilutions using OE₂ ELISA kit (DRG Instruments, GmbH, Marburg, Germany). Media removed from follicles cultured in control (n = 7) or in the presence of 100 ng/ml rhAct A with (n = 7) or without (n = 9) 50 ng/ml rhFSH on days 9 and 12 of the total in vitro period were analysed. As per the immunoassay kit instructions (DRG Instruments), reserved media were diluted with freshly prepared culture medium (1 in 10), placed in microtiter wells coated with polyclonal (rabbit) antibody raised against the OE₂ antigenic site, mixed with OE₂–HRP conjugate and incubated for 120 min. After incubation, the unbound conjugate was washed off, and a substrate solution of tetramethylbenzidine was added to allow development of colour. After 15 min, the reaction was stopped using dilute sulphuric acid, the wells were washed, and the absorbance of
each well at 450±10 nm was measured using a microtiter plate reader. The absorbencies were converted to \( \text{OE}_2 \) concentrations using Sigma Plot Version 9, with four parametric logistic functions (Systat Software Inc., Hounslow, UK) and \( \text{OE}_2 \) sensitivity of 25 pg/ml. Specificity of the assay was as follows: \( \text{OE}_2 \) 100%, oestradiol 0.055% and oestrone 0.2%. The inter- and intra-coefficients of variation were 6.81 and 7.25% respectively.

**Statistical analyses**

Inter- and intra-treatment differences were compared by measuring mean follicle diameters, mean oocyte diameters and \( \text{OE}_2 \) production on every third day of culture using a one-way ANOVA with subsequent t-tests. The proportions of quiescent and primary follicles and normal oocyte morphology were compared using \( \chi^2 \) analysis. The proportions of antral cavity and zona pellucida formation and cumulus cell adherence were compared using Fisher’s exact test.

**Declaration of interest**

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

**Funding**

Funding for this study was provided for M McLaughlin by a joint CASE PhD studentship between the Biotechnology and Biological Sciences Research Council and Trans Ova Genetics, Iowa, USA.

**Acknowledgements**

The authors wish to thank Mr John Binnie for excellent technical assistance.

**References**


www.reproduction-online.org


Thomas FH, Armstrong DG & Telfer EE 2003 Activin promotes oocyte development in ovine preantral follicles in vitro. Reproductive Biology and Endocrinology 1 76.


Received 15 January 2010
First decision 23 February 2010
Accepted 4 March 2010