Development, embryonic genome activity and mitochondrial characteristics of bovine–pig inter-family nuclear transfer embryos

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Abstract

The best results of inter-species somatic cell nuclear transfer (iSCNT) in mammals were obtained using closely related species that can hybridise naturally. However, in the last years, many reports describing blastocyst development following iSCNT between species with distant taxonomical relations (inter-classes, inter-order and inter-family) have been published. This indicates that embryonic genome activation (EGA) in xeno-cytoplasm is possible, albeit very rarely. Using a bovine–pig (inter-family) iSCNT model, we studied the basic characteristics of EGA: expression and activity of RNA polymerase II (RNA Pol II), formation of nucleoli (as an indicator of RNA polymerase I (RNA Pol I) activity), expression of the key pluripotency gene NANOG and alteration of mitochondrial mass. In control embryos (obtained by IVF or iSCNT), EGA was characterised by RNA Pol II accumulation and massive production of poly-adenylated transcripts (detected with oligo dT probes) in blastomere nuclei, and formation of nucleoli as a result of RNA Pol I activity. Conversely, iSCNT embryos were characterised by the absence of accumulation and low activity of RNA Pol II and inability to form active mature nucleoli. Moreover, in iSCNT embryos, NANOG was not expressed, and mitochondria mass was significantly lower than in intra-species embryos. Finally, the complete developmental block at the 16–25-cell stage for pig–bovine iSCNT embryos and at the four-cell stage for bovine–pig iSCNT embryos strongly suggests that EGA is not taking place in iSCNT embryos. Thus, our experiments clearly demonstrate poor nucleus–cytoplasm compatibility between these animal species.


Introduction

During early embryogenesis, a precise and accurate communication between nucleus and cytoplasm determines the success of such critical step of embryo development as embryonic genome activation (EGA). In inter-species somatic cell nuclear transfer (iSCNT), the situation is even more complicated as the donor nucleus and recipient cytoplasm originate from different species. Thus, during the maternal-to-embryonic transition (MET), activation of embryonic nuclei, which bear the genetic information of one species, critically depends on the stock of proteins and transcripts in the recipient cytoplasm (Schultz 1993, Minami et al. 2007), which is derived from another species. Initiation of gene transcription involves many factors, and the absence or the species incompatibility of just one of them can block the subsequent transcription (Heix et al. 1997).

Nevertheless, despite the possibility of nucleus–cytoplasm incompatibility, iSCNT represents a plausible experimental scheme especially when oocytes are extremely scarce. In 1999, the work by Dominko et al. (1999) raised hopes about the possibility of using oocytes from different species as recipients of somatic cell nuclei to save endangered species or to create embryonic stem cells for human cell therapy. Species that hybridise naturally are more likely to perform well in iSCNT experiments. This is understandable, since the natural production of living hybrid offspring shows that a certain nuclear–cytoplasmic compatibility exists between the two species (Mastromonaco et al. 2007). As a rule, iSCNT in mammals is more efficient when donor and recipient cells are from closely related species. For instance, inter-subspecies SCNT produced healthy offspring of Boer goat (Jian-Quan et al. 2007) and grey wolf (Kim et al. 2007). iSCNT embryos derived...
from mouflon (Ovis orientalis musimon), nucleus donor cells and sheep (Ovis aries) oocytes (Loi et al. 2001) could also develop to term. Wildcat (Felis silvestris lybica; Gomez et al. 2004) and sand cat (Felis margarita; Gomez et al. 2008) were produced using domestic cat (Felis catus) oocytes. iSCNT embryos derived from gaur (Bos gaurus) adult nucleus donor cells and bovine (Bos taurus) oocytes implanted and foetuses developed up to 200 days (Lanza et al. 2000). Inter-genus SCNT embryos derived from leopard cat (Prionailurus bengalensis) nucleus donor cells, and domestic cat oocytes were able to implant and form foetuses (Yin et al. 2006).

On the other hand, many studies have reported production of iSCNT morulae and blastocysts (BL) also when nucleus donor cells and recipient oocytes had a very distant taxonomical relation, like in the case of inter-family bovine–pig (Dominko et al. 1999, Uhm et al. 2007), inter-order cat- and panda–rabbit (Wen et al. 2005), camel- and Tibetan antelope–rabbit (Zhao et al. 2006), human–rabbit (Shi et al. 2008), dog–pig (Sugimura et al. 2009), tiger–pig (Hashem et al. 2007), human–bovine (Chang et al. 2003, Illmensee et al. 2006, Li et al. 2008) or inter-classes chicken–rabbit (Li et al. 2004) combinations. However, this approach remains ineffective.

The aim of the present study was to investigate pre-implantation development in bovine–pig iSCNT embryos that have been previously shown to develop to blastocyst stage at the rate of 4% (Uhm et al. 2007) to 8% (Dominko et al. 1999). Indeed, very few data are available on embryonic gene function before and at the time of EGA in iSCNT embryos (Arat et al. 2003, Bjerregaard et al. 2007, Chung et al. 2009, Hamilton et al. 2004, Kim et al. 2004, Li et al. 2008, Wang et al. 2009), and all show abnormal genome activation and gene expression profiles in the iSCNT embryos studied. It is likely that the ineffective ness of the iSCNT approach may be linked to abnormal (or absence of) EGA in such embryos due to aberrant nucleus–cytoplasm cross-talk. Since Bos taurus and Sus scrofa domesticus have a rather distant (inter-family) taxonomical relation and EGA occurs at different cell stages (8–16-cell stage in bovine and four-cell stage in porcine embryos), the pig–bovine iSCNT model may thus represent a useful model for investigating the nucleus–cytoplasm interaction during early embryo development. We have focused especially on nucleus remodelling, embryo development, donor cell-specific gene (collagen type VI α 1 gene (COL6A1)) fate and EGA during MET, with special attention to RNA polymerase II (RNA Pol II) activity, nucleoli formation (as an indicator of RNA polymerase I (RNA Pol I) activity) and activation of the pluripotency gene NANOG. Finally, we have investigated the behaviour of mitochondria under xeno-nuclear control.

**Results**

**Nucleus remodelling in bovine and porcine ooplasm**

To investigate pre-implantation development, we created both intra-species (bovine or porcine NT) and inter-species (bovine–pig or pig–bovine iSCNT) embryos by NT. Remodelling is believed to be an essential event in reprogramming of the genetic material after NT. This phenomenon involves structural changes in the donor nucleus, such as nuclear envelope breakdown, premature chromosome condensation (PCC) as well as pronuclear development and swelling (Tsunoda et al. 1989). The analysis of NT and iSCNT embryos (Supplementary Table 1, see section on supplementary data given at the end of this article) showed that about 90–100% of fused embryos, irrespectively of the origin of the recipient cytoplasm and of the nucleus donor cell, exhibited PCC 1 h post-fusion and swollen nuclei with a pronucleus-like appearance 3 h after activation. These results indicate that remodelling of the donor cell nucleus in the recipient cytoplasm is highly efficient and not species specific.

**In vitro development of iSCNT embryos**

In the first set of experiments, we investigated the developmental potential of iSCNT embryos reconstructed from porcine donor cells and bovine recipient oocytes. As a control, intra-species porcine NT embryos were used. We tested two porcine adult cell lines (PAF1 and PAF2) and one foetal fibroblast line (PFF) as nucleus donors for both NT and iSCNT embryos. In the control group, the proportion of embryos which had developed to the blastocyst stage was 20–38% at day 6 post-activation (Supplementary Table 2, see section on supplementary data given at the end of this article). Conversely, iSCNT embryos did not develop beyond the 16–25-cell stage (72 h after activation), and eventually, development was blocked. There was no sign of compaction or blastulation during further culture till day 8. No difference between SOFaa and CR1aa medium in the ability to support the development of iSCNT embryos was found.

We then repeated the same experiments using bovine foetal (BFF) as well as adult male (BAFm) and female (BAFf) fibroblasts as nucleus donors and porcine oocytes as recipients. The proportion of control bovine NT embryos that developed to blastocyst stage was about 50% at day 7 indicating a high efficiency of bovine NT and the suitability of the donor cell lines used (Supplementary Table 3, see section on supplementary data given at the end of this article). At 72 h after activation, all advanced embryos were at the four-cell stage in both NT and iSCNT groups. However, at 96 h, almost half (47%, n = 38) of control NT embryos possessed eight or more cells while all advanced iSCNT embryos (50–67.9%) were blocked at the 4–6-cell
stage (4.1 ± 0.2, n=30; 4–6 nuclei) indicating developmental arrest. There was no sign of compaction or blastulation during further culture till day 7.

Next, we counted the number of cells and nuclei in each pig–bovine iSCNT embryo because they just passed the porcine EGA stage. This step was necessary also as high level of cellular abnormality can be expected in iSCNT embryos and blastomere fragmentation can occur (Lorthongpanich et al. 2008). We analysed the most advanced embryos at 48 h (i.e. embryos made up of ≥ 4 cells), 72 h (≥ 8 cells) and 96 h (≥ 8 cells) after activation. There was no difference in the number of cells (10.2 ± 2.1) and nuclei (9.6 ± 2.3) between bovine NT and pig–bovine iSCNT embryos at 48 h. At 72 h, the average number of nuclei in NT embryos (Table 1) was equal to cell number and was significantly lower than in pig–bovine iSCNT embryos. At the same time, the average number of nuclei in pig–bovine iSCNT embryos (Table 1) was significantly higher than the number of nuclei with condensed chromosomes were detected (Fig. 1). Since chromosome condensation can also occur during apoptosis and one of the earliest events in the outer membrane, we stained fixed embryos with Annexin V, which binds to phosphatidylserine, to detect apoptosis is the translocation of phosphatidylserine to the embryos analysed, indicating that the bovine cytoplasm not only blocked de novo transcription of a fibroblast-specific gene irrespectively of the species affiliation of the nuclear donor but also effectively degraded transcripts inherited with the fibroblast cytoplasm. These preliminary experiments seem to confirm the data by Li et al. (2007) on the silencing of the fibroblast-specific vimentin and collagen genes in early yak–bovine iSCNT embryos.

Table 1 Number of cells and nuclei in bovine nuclear transfer (NT) and pig–bovine inter-species somatic cell nuclear transfer (iSCNT) embryos. Mean ± s.d.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>NT</th>
<th>iSCNT</th>
<th>Number of embryos</th>
<th>Number of cells/embryo, M ± s.d.</th>
<th>% of embryos with:</th>
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<tr>
<td></td>
<td>46</td>
<td>62</td>
<td>8–17</td>
<td>11.2 ± 2.6</td>
<td>Anucleate cells</td>
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<tr>
<td></td>
<td>45</td>
<td>32</td>
<td>8–25</td>
<td>13.8 ± 3.9</td>
<td>Binucleate cells</td>
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<tr>
<td></td>
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<td></td>
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<td>≥ 16 cells</td>
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<td>72 h</td>
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<td>96 h</td>
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Values with different letters in one row (a, b, c) are significantly different by t-test, *P*<0.01. Values with ‘a’ versus ‘a*’ in one row describe a tendency to be different, *P*=0.12. Values with different letters in one column (A, B) are significantly different, *P*<0.01.
Assessment of EGA

RNA Pol II

Since the previous experiments indicate that iSCNT embryos’ development is blocked at the time of EGA, we decided to analyse the events associated with the activation of transcription. In *Xenopus laevis*, EGA initiates with the activation of class II genes (mRNA), followed by class III genes (tRNA) and finally class I genes (rRNA; Shiokawa et al. 1989, 1994). In eukaryotes, RNA Pol II is responsible for transcription of mRNAs and of most of the small nuclear RNAs. Thus, we used an antibody that recognises RNA Pol II irrespectively of its phosphorylation status to visualise its accumulation, and RNA-FISH to estimate RNA Pol II activity through the detection of poly-adenylated (poly) mRNA accumulation in the nuclei. Moreover, since some aberrations can be found in genes’ expression of intra-species NT embryos, IVF embryos were included in this set of experiments as an additional control group.

We studied the most advanced IVF, NT and iSCNT embryos at 24, 48, 72 and 96 h after activation. In the nuclei of IVF and NT embryos, RNA Pol II (Fig. 2; Supplementary Figure 2, see section on supplementary data given at the end of this article) and poly(A) RNAs (Fig. 3; Supplementary Figure 3, see section on supplementary data given at the end of this article) were found at all studied stages of early embryo development and labelling intensity clearly increased at 72 and 96 h post fertilisation/activation in comparison with the earlier time points. Conversely, iSCNT embryos were characterised by low level of RNA Pol II expression and low transcriptional activity in the nuclei starting from the two-cell stage, and both RNA Pol II expression and activity did not increase at the time of EGA (Figs 2 and 3; Supplementary Figures 2 and 3).

Activation of gene expression in iSCNT embryos

Next, we assessed whether functional transcripts could be generated in iSCNT embryos, despite the low level of RNA Pol II activity by using porcine green fluorescent protein (GFP)-positive foetal fibroblasts (GFP+ cells under the control of the constitutive PGK promoter) as nucleus donor cells for pig–bovine iSCNT embryos. Accumulation of GFP was detected in iSCNT embryos starting from the one-cell stage (24 h after activation), and more than 80% of pig–bovine iSCNT embryos (Supplementary Table 2) showed accumulation of GFP at day 7 (Supplementary Figure 4, see section on supplementary data given at the end of this article). This indicates that low level of RNA Pol II transcription was present in iSCNT embryos.

Another event associated with RNA Pol II activity but crucially dependent on EGA is the expression of NANO2, a key pluripotency gene (Brevini et al. 2006, Gandolfi et al. 2006). We were unable to detect NANO2 mRNA in pig–bovine (Supplementary Figure 5D) iSCNT embryos at any of the developmental stages (2-, 4-, 8- and 16-cell) analysed. These data confirm that no or highly aberrant genome activation takes place at the porcine MET (4–8-cell) or at the bovine MET (8–16 cell).

RNA Pol I activation and nucleolar maturation

In mammals, the fact that transcriptional activation of class I genes (rRNA) depends on the previous activation of class II genes (mRNA; Shiokawa et al. 1989, 1994) was recently confirmed in bovine (Maddox-Hyttel et al. 2007, Svarcova et al. 2007) and porcine embryos (Maddox-Hyttel et al. 2007). Specifically, RNA Pol I as well as other key nucleolar proteins (Svarcova et al. 2007) are de novo expressed from embryonic genome, and consequently, rRNA gene activation and the associated nucleolus formation have been suggested as
markers of EGA in mammalian embryos (Bjerregaard et al. 2007). Nucleolin is a multifunctional maternally inherited nucleolar protein (Svarcova et al. 2007) engaged in the late processing of rRNA that colocalises with RNA Pol I to nucleoli. We thus used an antibody against nucleolin (C23) to evaluate nucleolus formation in the more advanced NT and iSCNT embryos at 96 h after activation (Figs 1 and 4). NT embryos were characterised by the presence of intensely labelled polygonal and shell-like mature nucleoli. Conversely, pig–bovine (Fig. 1C1) and bovine–pig (Fig. 4D) iSCNT embryos were characterised by prevailing nucleoplasmic localisation of nucleolin with occasional concentration in faintly stained half-rings or rings in bovine nuclei or in several dots or rings in some porcine nuclei.

This result, together with the data on RNA Pol II and NANOG, suggests that the embryonic genome is not properly activated in iSCNT embryos. To further test this hypothesis, we incubated bovine NT embryos with actinomycin D (AD) and porcine NT embryos with α-amanitin (AA). Long-term (48 h) incubation with AD or AA effectively blocked EGA in NT embryos as indicated by the absence of mature nucleoli (Figs 1D and 4C) and the appearance of nuclei with condensing chromatin in

Figure 2 RNA Pol II was visualised with the CTD4H8 antibody that detects both phosphorylated and unphosphorylated RNA Pol II in bovine IVF (A and A1), NT (B and B1) and pig–bovine iSCNT (C and C1) embryos. The embryos were fixed at the two-cell stage (A–C) and at the most advanced stages at 96 h after insemination/activation (A1, B1 and C1). CTD4H8 labelling revealed the presence of RNA Pol II in the nuclei of (A) IVF, (B) NT and (C) iSCNT two-cell stage embryos, significant accumulation of RNA Pol II in the nuclei of (A1) IVF and (B1) NT embryos, and (C1) the lack of RNA Pol II accumulation in iSCNT embryos. Scale bar, 100 μm.

Figure 3 RNA Pol II activity, as determined by the accumulation of polyadenylated transcripts in the nuclei, was assessed by RNA FISH with a fluorescein-labelled oligo dT probe in bovine IVF (A, A1 and A2), NT (B, B1 and B2) and pig–bovine iSCNT (C, C1 and C2) embryos. Embryos were fixed and labelled at the two-cell stage (A–C) and at the most advanced stages at 96 h after insemination/activation (A1, B1 and C1). Nuclei were counterstained with DAPI for DNA visualisation. Dot-like labelling of polyadenylated transcripts in the nuclei of two-cell stage (A) IVF, (B) NT and (C) iSCNT embryos and localisation of polyadenylated transcripts in DAPI-stained nuclei of (A1) IVF, (B1) NT and (C1) iSCNT embryos. Accumulation of polyadenylated transcripts in the nuclei of 8–16-cell stage (A2) IVF, (B2) NT embryos and (C2) dot-like labelling in the nuclei of iSCNT embryos. Scale bars, 100 μm.
bovine NT embryos (not shown). Few faintly stained ring-like structures labelled with anti-nucleolin antibody could be found in some nuclei both in bovine and porcine NT embryos. Short incubation (3 h) with AD, to block RNA Pol I activity, resulted in complete disappearance of mature nucleoli (Figs 1B and 4B), nucleolin translocation into the nucleoplasm and few weekly anti-nucleolin-stained rings in some blastomeres. Thus, the formation of mature nucleoli in NT embryos depends on the synthesis of RNA Pol I during EGA and RNA Pol I-guided transcription of rRNA. The similar appearance of iSCNT embryos and AA- or AD-treated NT embryos suggests that in iSCNT embryos, the embryonic genome is not properly activated, RNA Pol I cannot be transcribed and as a result functional nucleoli cannot be formed.

**Assessment of mitochondrial changes in NT and iSCNT embryos during development**

Another important issue in iSCNT embryo development is the interaction of the nucleus with complex cytoplasmic organelles like mitochondria. Since mitochondria are involved in a broad range of critical functions (energy supply, cell signalling and apoptosis), inadequate or perturbed mitochondrial function may adversely affect SCNT efficiency (Hiendleder et al. 2005). Moreover, Lee et al. (2002) have shown that the increase in mitochondrial mass depends on cytoplasmic protein synthesis. Thus, changes in the mitochondrial mass in embryos are most probably linked to EGA.

To follow changes in mitochondria, we stained iSCNT and NT embryos with the cationic dye JC-1 at 48, 72 and 96 h after activation. JC-1 exists in a monomeric form, emitting at 527 nm (green fluorescence) after excitation at 490 nm. In the cell, it corresponds to the low mitochondrial membrane potential (<100 mV) and characterises relative mitochondrial mass (Mancini et al. 1997). In the presence of high membrane potential (>140 mV), JC-1 forms so called J-aggregates that are associated with a large shift in emission (590 nm, red fluorescence). The ratio of red/green fluorescence characterises the inner mitochondrial membrane potential (Reers et al. 1991). We have found that at 48 h, NT and iSCNT embryos possessed equal relative mitochondrial mass (Fig. 5) and J-aggregate content (not shown). However, at 72 and 96 h, both parameters were significantly lower in iSCNT embryos than in their NT counterparts (P < 0.05). We then asked whether this discrepancy could be linked to the arrest of development and aberrant EGA observed in iSCNT embryos. To this aim, we inhibited genome transcription with AA in porcine and bovine NT embryos at 48 h for 24 h, and then stained them with JC-1 at 72 h. AA treatment led to significantly lower values of mitochondrial mass than in untreated NT embryos confirming that the most likely reason of low mitochondrial parameters in iSCNT embryos is the aberrant EGA.

**Discussion**

In the present manuscript, we have tried to characterise the main events of EGA, namely activation of RNA Pol II transcription at the time of MET that should lead to subsequent transcription of RNA Pol I and pluripotency genes. Any serious aberrations of these events cause the developmental failure of the embryos. Our results indicate that the most likely cause of pig–bovine and bovine–pig iSCNT embryos developmental failure is the
absence of EGA or its aberrant implementation due to nucleus–cytoplasm incompatibility.

Differently from other laboratories (Dominko et al. 1999, Uhm et al. 2007, Song et al. 2008), we did not manage to obtain pig–bovine and bovine–pig iSCNT embryos that could develop to the blastocyst stage, whereas intra-species NT proved to work effectively in our hands. Since the efficiency of NT (Boiani et al. 2005, Lagutina et al. 2006) and even more of iSCNT (Liu et al. 2004, Zhao et al. 2006, Sugimura et al. 2009) embryo development crucially depends on embryo culture conditions, we tested both SOFaa and CR1aa, which have been previously reported to support bovine (Galli et al. 2003) and porcine (Lagutina et al. 2006) embryo development and pig–bovine iSCNT blastocyst development (Dominko et al. 1999, Uhm et al. 2007) respectively. No difference was observed, and pig–bovine/bovine–pig iSCNT embryos were not able to escape the block at the 16–25-cell/four-cell stage, which corresponds to the time of bovine MET (8–16-cell stage; Kanka 2003) and porcine MET (four-cell stage; Tomanek et al. 1989, Freitag et al. 1991, Jarrell et al. 1991). The reasons of such developmental failure in our culture conditions are not clear, but it should not be ascribed to the quality of our in vitro maturation–in vitro culture (IVC) media or to the NT technique used, since several offspring were born following bovine and porcine NT.

The analysis of remodelling of the donor nucleus in the cytoplasm of a recipient oocyte from a different species indicates that the normal pattern of nucleus remodelling was followed as previously reported in different studies on mammalian NT and iSCNT embryos (Tarkowski & Balakier 1980, Dominko et al. 1999, Arat et al. 2003, Uhm et al. 2007, Lee et al. 2008). Thus, the biochemical mechanism of this process is universal, and it works also in intra-class NT embryos derived from chicken blastodermal cells and rabbit oocytes (Liu et al. 2004).

On the other hand, the cleavage kinetics of NT and iSCNT embryos was comparable only up to 48 h of IVC, and during the following 24 h, the development of iSCNT embryos was arrested at the MET stage: that is, at the four-cell stage for bovine–pig embryos and at the 8–16-cell stage for pig–bovine embryos. However, pig–bovine iSCNT embryos divided significantly faster than bovine NT control embryos, and many binucleate blastomeres were observed in about 50% of them. Such accelerated cleavage was reported by Memili & First (1998) in bovine embryos treated with AA to prevent EGA. This finding together with the observation that our iSCNT embryos stopped dividing at the moment of the oocyte MET suggests that EGA did not take place in our iSCNT embryos.

To confirm this hypothesis, we then assessed RNA Pol II expression and activity. Low level of activity was observed in the nuclei of IVF, NT and iSCNT embryos as early as the two-cell stage. This confirms the data on low-grade transcription during the first three cell cycles in bovine embryos (Barnes & First 1991, Plante et al. 1994, Hyttel et al. 1996, Viuff et al. 1996, 1998, Memili & First 1998, Svarcova et al. 2007, Kues et al. 2008) and extends our knowledge about porcine embryos that were considered transcriptionally inactive until late in the third cell cycle, i.e. at the four-cell stage (Tomanek et al. 1989, Freitag et al. 1991, Jarrell et al. 1991). However, during further IVC, iSCNT embryos did not show the important increase in RNA Pol II activity observed in control embryos, which went through normal EGA.

Moreover, 96 h after activation, iSCNT embryos did not present mature nucleoli. Indeed, immunostaining with an anti-nucleolin antibody revealed faint nucleoplasmic labelling with weakly stained half-rings or rings in bovine nuclei or several dots or rings in some porcine nuclei without formation of mature nucleoli. The same pattern of staining was reported in pig–ovine (Hamilton et al. 2004) and rhesus monkey–bovine iSCNT embryos (Song et al. 2009) that were not able to properly activate the genome and form BL. Inhibition of RNA Pol I transcription in control NT embryos led to disappearance of nucleolin-labelled mature nucleoli and dispersion of nucleolin into the nucleoplasm. The dispersed nucleoplasmic nucleolin and inability to form nucleoli were also observed in NT embryos after inhibition of EGA as a result of the absence of de novo transcription of RNA Pol I that depends on RNA Pol II activity. Therefore, taking into account the developmental block of iSCNT embryos and the absence of RNA Pol II accumulation and its low activity, we conclude that the failure to form mature nucleoli is the result of the absence of EGA.

We also studied the expression pattern of NANOG (Brevini et al. 2006, Gandolfi et al. 2006), which depends on EGA. Porcine NANOG mRNA was not
detected even in the best (≥16-cell) pig–bovine iSCNT embryos at the time of porcine and bovine EGA as previously reported for chimpanzee–bovine iSCNT embryos (Wang et al. 2009). Comparison of gene expression in 8–16-cell stage human–bovine or human–rabbit embryos and human IVF or NT embryos using single-embryo transcriptome profiling revealed general downregulation of human genes in the bovine and rabbit recipient cytoplasm (Chung et al. 2009). These data and the absence of Alpl2 (eAp), Pou5f1 (Oct4) and Cdb1 (c-Cad) expression in mouse embryonic fibroblast (MEF)–bovine iSCNT embryos (Arat et al. 2003, Kim et al. 2004) suggests that bovine ooplasm does not reprogram properly donor nuclei from other species.

On the other hand, we extended our knowledge about the cytoplasm control in iSCNT embryos over xenonucleus genes using constitutive and tissue-specific inducible promoters. The permanent low RNA Pol II activity in iSCNT embryos was sufficient for the observed GFP accumulation in iSCNT embryos derived from fibroblasts that carry the GFP gene under the control of the constitutive PGK promoter, as previously described by Uhm et al. (2007) using GFP controlled by the rat β-actin promoter. This permanent activity of RNA Pol II is enough to sustain GFP expression in MEF–bovine (Arat et al. 2003) and panda-rabbit iSCNT embryos (Wen et al. 2005), avian feather keratin expression in chicken–rabbit intra-classes iSCNT embryos (Liu et al. 2004), housekeeping gene expression in mouse–bovine iSCNT embryos (Arat et al. 2003, Kim et al. 2004). However, the abnormal pattern of housekeeping genes expression (Arat et al. 2003, Kim et al. 2004, Wang et al. 2009) confirmed the inadequate embryonic genome activity in all tested iSCNT embryos.

On the contrary, our data on the absence of transcription of COL6A1 in iSCNT embryos (Supplementary Figure 5) indicate that bovine cytoplasm can block de novo transcription of a fibroblast-specific gene irrespectively of the species affiliation of the donor nucleus. This agrees with the results by Inoue et al. (2006) and Green et al. (2007) on the silencing of donor cell-specific genes in NT embryos using muscle and hematopoietic cells as nucleus donors and with the statement by Vassena et al. (2007) that the donor genome is markedly silenced by the ooplasm at the one-cell stage of NT embryo development. The silencing of porcine genes in bovine cytoplasm does theoretically prove a high homology of elements controlling gene silencing. Conversely, the expression of avian feather keratin in chicken–rabbit intra-class NT embryos (Liu et al. 2004) as early as in eight-cell embryos is the example of the inability of mammalian ooplasm to correctly recognise avian tissue-specific inducible promoters or silencers.

Another interesting question of iSCNT embryo development is the interaction of the xeno-nucleus with the cytoplasmic mitochondria that have their own DNA and can synthesise RNAs and proteins. It is known that the number of mtDNA copies/embryo is thought to be stable during cleavage until replication begins at an undefined time post-implantation (Aiken et al. 2008) and the total number of mitochondria does not increase during the preimplantation stage of development (Szlöcsi 1965). However, during early embryo development, mitochondria undergo changes in shape and structure and the number of cristae increases (Betteridge 1988, Van Blерkom 2009). The detailed study (Mtango et al. 2008) of temporal expression of mRNAs related to mitochondrial biogenesis revealed a persistent expression of maternally encoded mRNAs in combination with transcriptional activation and mRNA accumulation around the time of EGA. All proteins involved in mitochondrial biogenesis and functions, currently estimated at ~1500, are encoded by nuclear DNA, synthesised in the cytosol, and then targeted and imported into mitochondria by specific mechanisms (Cotter et al. 2004, Hiendlieker et al. 2005). Lee et al. (2002) demonstrated that the increase in mitochondrial mass does not depend on mtDNA presence or function but on cytoplasmic protein synthesis. Thus, changes of mitochondrial mass in embryos are most probably dependent on EGA. We have found that the relative mitochondrial mass and J-aggregate content were comparable in NT and iSCNT embryos at 48 h of IVC when only maternally inherited transcripts were available. However, both parameters in iSCNT embryos at 72 h post-activation were significantly lower than in control NT embryos and resembled those of NT embryos after inhibition of EGA. These results further confirm that EGA is not taking place in iSCNT embryos, and suggest that comparison of the relative mitochondrial mass in iSCNT and control NT embryos at the time of the host oocyte’s EGA can serve as an express test to assess genome activation in iSCNT embryos. Another interesting question of the interaction of the xeno-nucleus with the mitochondria in iSCNT embryos that was not studied here is the level of compatibility between nuclear and mitochondrial genomes in iSCNT embryos as the existence of species-specific incompatibility between nuclear and mitochondrial genomes of closely related species has been demonstrated in cybrids of primates (Kenyon & Moraes 1997, Barrientos et al. 2000) and murids (Dey et al. 2000, McKenzie & Trounce 2000, McKenzie et al. 2003).

To summarise, our data strongly indicate that bovine and porcine oocytes cannot properly reprogram the genome of each other in iSCNT embryo and activate embryonic genome transcription at the time of MET. The causes of such species incompatibility should be investigated more carefully. One of the reasons might be some specific differences in gene structure. The high homology of mammalian proteins and their coding genes in the case of iSCNT seems not to guarantee proper gene function. For instance, the mouse Pou5f1 promoter correctly activates GFP transcription at the
time of EGA in bovine NT embryos derived from bovine fibroblasts carrying a mouse Pou5f1 promoter-driven GFP reporter gene (Wuensch et al. 2007), while it does not in porcine NT embryos derived from porcine fibroblasts carrying the same transgene (D Brunetti, personal communication). An aberrant DNA methylation pattern in iSCNT embryos may be another reason. Chen et al. (2004) demonstrated that demethylation of donor repetitive sequences is driven by the recipient ooplasm, and ooplasms from different species have different capabilities to demethylate genes. The results of our study raise the question of inter-species nuclear–cytoplasmic or genomic compatibility. The role of nuclear–cytoplasmic interactions in EGA, gene activation and dependence of the activation of a given gene on the function of others is a large field that needs to be investigated.

Materials and Methods

Oocyte maturation

Unless otherwise stated, all chemicals were purchased from Sigma–Aldrich. Oocytes were aspirated from 3 to 8 mm in diameter ovarian follicles of killed animals. In vitro manipulation was carried out essentially as described previously (Lagutina et al. 2006). Bovine oocytes were matured in TCM199 supplemented with 10% (v/v) FCS, 1 μl/ml ITS Media Supplement (insulin, transferrin, selenite, Sigma, cat. no. I1884), 1 mM sodium pyruvate, 0.5 mM L-cystein, 10 mM glycine, 100 μM β-mercaptoethanol, gonadotropins (0.05 IU/ml FSH and 0.05 IU/ml LH; Pergovet 75, Serono). Porcine oocytes were matured in DMEM-F12 supplemented with 10% (v/v) FCS, 110 μg/ml sodium pyruvate, 75 μg/ml ascorbic acid, 100 μg/ml glutamine, 5 μg/ml myoinositol, 0.4 mM cystine, 0.6 mM cysteamine, ITS Media Supplement (1 μl/ml), gonadotropins (0.05 IU/ml FSH and 0.05 IU/ml LH; Pergovet 75, Serono), 100 ng/ml LONGR3 insulin-like growth factor 1 (IGF1; recombinant IGF analogue), 50 ng/ml LONG epidermal growth factor (EGF; recombinant EGF analogue) and 5 ng/ml bovine fibroblast growth factor (bFGF; human recombinant). Bovine and porcine oocytes were cultured at 38.5 °C in 5% CO2 in humidified air for 24 h respectively.

Production of bovine and porcine IVF embryos

Bovine IVF embryos were produced as previously described (Lagutina et al. 2002). Porcine IVF embryos: after 42 h of maturation, oocytes were partially denuded of the cumulus cells and cultured in four-well plates containing 0.3 ml/well of TALP prepared without glucose, supplemented with 1 mg/ml heparin, 20 mM α-penicillamine, 100 mM hypotaurine, 1 mM adrenaline, and 0.1×106 spermatozoa/ml. Motile spermatozoa were obtained by centrifugation of frozen–thawed semen on a Percoll discontinuous density gradient for 40 min at 750 g, washed in Ca-free TALP and pelleted by centrifugation for 10 min at 400 g.

Preparation of donor cells

Bovine and porcine adult and foetal fibroblasts and porcine foetal GFP-positive fibroblasts (GFP under the control of the PGK promoter) at passages 1–10 were used as donor cells. Fibroblasts were derived from ear biopsies according to standard procedures and cultured in DMEM+TCM 199=1:1 with 10% FCS. The medium for porcine fibroblasts was supplemented with 5 ng/ml bFGF (human recombinant). All donor cells were tested for efficient blastocyst development after intra-species NT. Two days before NT, donor cells were passaged, and one day before NT, they were serum starved (0.5% FCS) for 24 h.

Nuclear transfer

NT was carried out essentially as described previously (Lagutina et al. 2006). Briefly, oocytes were denuded of cumulus cells by vortexing in SOF-HEPES in the presence of hyaluronidase, and oocytes with extruded polar bodies were selected. NT embryos were reconstructed following a zona-free method (Oback et al. 2003). After zona pellucida digestion with 0.5% pronase in PBS, oocytes were washed in SOF-HEPES with 10% FCS and returned to maturation medium. All further manipulations were performed in SOF-HEPES with 10% FCS. Zona-free oocytes were exposed to cytochalasin B (7.5 μg/ml) and Hoechst (5 μg/ml) for 5 min. Metaphase chromosomes were removed under u.v. light with a blunt enucleation pipette. Then, each cytoplast was individually washed for few seconds in 300 μg/ml phytohemagglutinin P in PBS and quickly dropped over a single donor cell (Vajta et al. 2003) settled at the bottom of a microdrop of donor cell suspension diluted in SOF-HEPES. Formed cell couples were washed in 0.3 M mannitol (Ca++-,free, 100 μM Mg++) solution, fused by double DC-pulse of 1.2 kV/cm applied for 30 μs and returned to maturation medium.

Activation

Bovine oocytes

Embryos (2–3 h after fusion) were activated 27–28 h after the onset of oocyte maturation with 5 μM ionomycin in SOF-HEPES for 4 min followed by 4-h culture in individual 2-μl drops of 2 mM 6-dimethylaminopurine in SOF supplemented with MEM essential and non-essential amino acids and 4 mg/ml BSA (m-SOFaa) at 38.5 °C in humidified atmosphere of 5% CO2, 5% O2 and 90% N2.

Porcine oocytes

Embryos (2–3 h after fusion) were activated 48–50 h after the onset of oocyte maturation by double DC-pulses of 1.2 kV/cm for 30 μs in 0.3 M mannitol solution, containing 1 mM Ca++, Mg++, (Cheong et al. 2002) and 100 μM Mg++, followed by 4-h culture in maturation medium with 5 μg/ml cytochalasin B at 38.5 °C in humidified atmosphere of 5% CO2, 5% O2 and 90% N2.
Embryo in vitro culture

NT and iSCNT embryos were cultured according to the WOW method (Vajta et al. 2000) individually in microwells in 20-μl drops of m-SOFaa in 5% CO₂, 5% O₂ and 90% N₂ humidified atmosphere at 38.5 °C. Half of the medium was renewed with fresh m-SOFaa at days 4 and 6 (day 0 was the day of NT). Cleavage was assessed 48 h after activation, the occurrence of BL at day 6 and 7. All experiments were done using three or more replicates (unless otherwise indicated). However, some experiments were performed in CR1 medium supplemented with MEM essential and non-essential aminoacids that were claimed to be efficient for the production of iSCNT BL in several species using bovine oocytes (Dominko et al. 1999, Uhm et al. 2007).

To assess the effect of EGA inhibition on the formation of nucleoli, bovine and porcine embryos were cultured in medium supplemented respectively with 25 μg/ml AA or 0.2 μg/ml AD in the window of time going from 48 to 96 h of IVC.

To assess the effect of inhibition of RNA Pol I activity on the function of nucleoli, bovine and porcine embryos were cultured in medium supplemented with 0.2 μg/ml AD for 3 h starting at 93 h of IVC.

To evaluate the effect of inhibition of EGA on mitochondria, bovine and porcine NT embryos were cultured in medium supplemented with 25 μg/ml AA starting at 48 h and till 72 h after activation.

RT-PCR

Unless otherwise indicated, all chemicals were purchased from Invitrogen.

RNA extraction from pig–bovine iSCNT embryos

Poly(A)+ RNA was isolated from three separate groups of five iSCNT embryos at the 2-, 4-, 8- and 16-cell stage using the Dynabeads mRNA direct MicroKIT (Dynal-Invitrogen), following the manufacturer’s instruction (Supplementary Materials and Methods, see section on supplementary data given at the end of this article).

RT

For each sample, RT was carried out in a total volume of 20 μl of reaction mixture using 1 μl of Oligod(T)12–18 primer and 1 μl dNTP Mix (10 mM each). RNA was denatured at 65 °C for 5 min, then 4 μl of first-strand buffer (250 mM Tris–HCl pH 8.3, 375 mM KCl and 15 mM MgCl₂), 1.5 μl 50 mM MgCl₂, 2 μl of 0.1 M dithiothreitol and 1 μl of RNaseOUT Recombinant Ribonuclease Inhibitor (40 U/μl) were added. RT was performed with 200 U (1 μl) Superscript II RT for 1 h at 42 °C, followed by a denaturation step at 70 °C for 15 min. Tubes without RNA or RT were prepared as negative controls (data not shown).

PCR

Accession number, primer sequences, annealing temperature, amplification product sizes and number of amplification cycles for each gene are presented in Supplementary Table 4, see section on supplementary data given at the end of this article. Rabbit globin mRNA was used as exogenous standard to normalise reactions for the amount of RNA in each sample. PCR amplification was performed with cDNA concentrations equivalent to one embryo from different pools of iSCNT embryos. The reaction mix consisted of 11 μl of 50 mM MgCl₂, 0.3 μl of Taq DNA polymerase (5 U/μl), 2 μl of 10 X PCR buffer (200 mM Tris–HCl pH 8.4, 500 mM KCl), 1 μl NTP Mix (10 mM each), 1 μl (100 pmol) of each sequence-specific primer and sterile water up to 20 μl. Amplifications were carried out in an automated thermal cycler (iCycler, Bio-Rad). To confirm the identity of the RT-PCR fragments, amplification products from each primer pair were separated by gel electrophoresis in 2% agarose gels, purified using Spin-X centrifuge tube filters (Corning, Milan, Italy; Sigma-Aldrich), sequenced (SEQLAB, Gottingen, Germany) and aligned using Clustal W 1.82 (EMBL-EBI service).

Immunocytochemistry

IVF, NT and iSCNT embryos at 24, 48, 72 and 96 h after activation were fixed in 4% paraformaldehyde in PBS with 0.1% Triton X-100 at 4 °C for 3 h and subsequently stained with antibodies against nucleolin (C23, an indirect marker of RNA Pol I activity; Santa Cruz Biotechnology Inc., Santa Cruz, CA, USA, sc-8031) and RNA Pol II (CTD4H8; Covance Research Products Inc., Princeton, NJ, USA) overnight. Embryos were washed in PBS/BSA and incubated with the secondary antibody conjugated with FITC (Santa Cruz Biotechnology) at room temperature for 2 h, washed in PBS/BSA and mounted in Cytifluor with DAPI.

FISH

RNA Pol II activity was assessed in IVF, NT and iSCNT embryos at 24, 48, 72 and 96 h after activation by detection of polyadenylated transcripts in the nuclei using RNA FISH with a fluorescein-labelled oligo dT probe (Sigma). This probe was 45 bases long with fluorescein inserted every ten position. The method was based on the protocol by Tam et al. (2002) with slight modifications for embryos (Supplementary Materials and Methods).

Analysis of GFP reporter gene expression

To visualise RNA Pol II activity in pig–bovine iSCNT embryos derived from porcine foetal fibroblasts that carry the GFP reporter gene under the control of the constitutive promoter PGK, we estimated the expression of the GFP gene 24 h after activation and at day 7 under a fluorescence microscope using a standard FITC filter set.

Assessment of mitochondrial changes

We used JC-1 fluorescent dye (Invitrogen) that accumulates in mitochondria to assess the behaviour of mitochondria in bovine and porcine NT and iSCNT embryos. Embryos at 48, 72 and 96 h after activation were stained with 2 μM JC-1 in SOF-HEPES with 10% FCS at 37 °C for 1 h. Images were collected using a double FITC/Rodamine filter. Both red and yellow coloured fields were considered as a sign of mitochondrial activity.
green fluorescence emissions from the area of the embryo where dye distribution artefacts have been removed were analysed with Adobe Photoshop Elements 2. The data were presented as a mean fluorescence intensity of the embryo. In total, not less than 20 embryos per every group/time point were analysed in three replicates.

**Nucleus remodelling, number of nuclei and cells**

NT and iSCNT embryos 1 h after fusion and 3, 48, 72 and 96 h after activation were stained with Hoechst 33342 to estimate morphology, number and distribution of nuclei.

**ApoAlert Annexin V staining**

Staining of NT and iSCNT embryos at 96 h after activation was performed according to the manufacturer’s instructions (Clontech Laboratories, Inc). Images were analysed using FITC and Rodamine filters.

**Images**

Embryos were viewed with the fluorescence microscope Nikon Eclipse 80i using FITC, Rodamine or dual filter set for FITC/Rodamine, and images were collected using a DS Camera Head DS-5M with DS Camera Control Unit DS-L1 (Nikon Instruments, Milan, Italy).

**Statistical analysis**

Differences between the experimental groups were verified using Student's t-test and the χ² test. A value of P<0.05 was considered significant.

**Supplementary data**

This is linked to the online version of the paper at http://dx.doi.org/10.1530/REP-09-0578.

**Declaration of interest**

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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**References**

Aiken CEM, Cindrova-Davies T & Johnson MH 2008 Variations in mouse mitochondrial DNA copy number from fertilization to birth are associated with oxidative stress. Reproductive Biomedicine Online 17 806–813.


Barrientos A, Muller S, Dey R, Wienberg J & Moraes CT 2000 Cytochrome c oxidase assembly in primates is sensitive to small evolutionary variations in amino acid sequence. Molecular Biology and Evolution 17 1508–1519.


Chang KH, Lim JM, Kang SK, Lee BC, Moon SY & Hwang WS 2003 Blastocyst formation, karyotype, and mitochondrial DNA of interspecies embryos derived from nuclear transfer of human cord fibroblasts into enucleated bovine oocytes. Fertility and Sterility 80 1380–1387.


Hiendleder S, Zakhartchenko V & Wolf E 2005 Mitochondria and
Hyttel P, Viuff D, Avery B, Laurincik J & Greve T
Jarrell VL, Day BN & Prather RS
Jian-Quan C, Juan C, Xu-Jun X, Guo-Hui L, Si-Guo L, Hong-Ying S,
Kanka J
Green AL, Wells DN & Oback B
Heix J, Zomerdijk JC, Ravanpay A, Tjian R & Grummt I
Kim N-H, Shin MR & Park SH 2004 Bovine oocyte cytoplasm supports
Kim MK, Jang G, Oh HJ, Yuda F, Kim HJ, Hwang WS, Hossein MS, Kim JJ,
Gomez MC, Pope CE, Giraldo A, Lyons LA, Harris RF, King AL, Cole A,
Godke RA & Dresser BL 2004 Birth of African wildcat cloned kittens
Gomez MC, Pope CE, Kuther RH, Ricks DM, Lyons LA, Ruhe M, Dumas C,
Lyons J, Lopez M, Dresser BL et al. 2008 Nuclear transfer of sand cat cells
to enucleated domestic cat oocytes is affected by cryopreservation of
Green AL, Wells DN & Oback B 2007 Cattle cloned from increasingly
Hamilton HM, Peura TF, Laurincik J, Walker SK, Maddocks S & Maddox-
Hytell P 2004 Ovine ooplasm directs initial nuclear assembly in
embryos cloned from ovine, bovine, and porcine cells. Molecular Reproduction
and Development 69 117–125.
Hashem MA, Bhandari DP, Kang SK & Lee BC 2007 Cell cycle analysis and
interspecies nuclear transfer of in vitro cultured skin fibroblasts of the
Siberian tiger (Panthera tigris altaica). Molecular Reproduction and
Development 74 403–411.
Heiz J, Zomerdijk JC, Ravanpay A, Tjian R & Grummt I 1997 Cloning of
murine RNA polymerase II-specific TAF factors: conserved interactions
between the subunits of the species-specific transcription initiation factor
TIF-II/B1/S1. PNAS 94 1733–1738.
Hiedler S, Zakhartchenko V & Wolf E 2005 Mitochondria and the
success of somatic cell nuclear transfer cloning; from nuclear-
mitochondrial interactions to mitochondrial complementation and
mitochondrial DNA recombination. Reproduction, Fertility, and Development
17 69–83.
Hytell P, Viff D, Avery B, Laurincik J & Greve T 1996 Transcription and cell
cycle-dependent development of intranuclear bodies and granules in
263–270.
Illnensee K, Levanduski M & Zavos PM 2006 Evaluation of the embryonic
preimplantation potential of human adult somatic cells via an embryo
interspecies bioassay using bovine oocytes. Fertility and Sterility 85
(Supplement 1) 1248–1260.
Inoue K, Ogunki N, Miki H, Hirose M, Noda S, Kim JM, Aoki F, Miyoshi H
& Ogura A 2006 Inefficient reprogramming of the hematopoietic stem cell
Jarrell VL, Day BN & Prather RS 1991 The transition from maternal to
zygotic control of development occurs during the 4-cell stage in the
domestic pig. Sus scrofa: quantitative and qualitative aspects of protein
Jian-Quan C, Juan C, Xu-Jun X, Guo-Hui L, Si-Guo L, Hong-Ying S,
You-Bing W & Guo-Xiang C 2007 Effect of cytoplasm on the development of inter-subspecies nuclear transfer reconstructed goat embryo.
Molecular Reproduction and Development 74 568–573.
Kanka J 2003 Gene expression and chromatin structure in the
pre-implantation embryo. Theriogenology 59 3–19.
Kenyon L & Moraes CT 1997 Expanding the functional human
mitochondrial DNA database by the establishment of primate xenonochondrial
hybrids. PNAS 94 9131–9135.
Kim N-H, Shin MR & Park SH 2004 Bovine oocyte cytoplasm supports
nuclear remodeling but not reprogramming of murine fibroblasts.
Reproduction, Fertility, and Development 16 145–146. Abstract 47.
Kim MK, Jang G, Oh HJ, Yuda F, Kim HJ, Hwang WS, Hossein MS, Kim JJ,
Shin NS, Kang SK et al. 2007 Endangered wolves cloned from adult
somatic cells. Cloning and Stem Cells 9 130–137.
Kues WA, Sudheer S, Herrmann D, Carnwath JW, Havlicek V,
Besenfelder U, Lehrach H, Adjaye J & Niemann H 2008 Genome-
wide expression profiling reveals distinct clusters of transcriptional
regulation during bovine preimplantation development in vivo. PNAS
105 19768–19773.
Lagutina I, Ponderato N, Lazzari G & Galli C 2002 Kinetics of oocyte
maturation and subsequent development of IVF, parthenogenetic, and
NT bovine embryos after meiotic inhibition with roscovin. Cloning
and Stem Cells 4 113–119.
Lagutina I, Lazzari G & Galli C 2006 Birth of cloned pigs from zona-free
nuclear transfer blastocysts developed in vitro before transfer. Cloning
and Stem Cells 8 283–293.
Lanza RP, Cibelli JB, Diaz F, Moraes CT, Farin PW, Farin CE, Hammer CJ,
West MD & Damiani P 2000 Cloning of an endangered species (Panthera tigris altaica) using interspecies nuclear transfer.
Lee HC, Yin PH, Chi CW & Wei YH 2002 Increase in mitochondrial mass in
human fibroblasts under oxidative stress and during replicative cell
Lee E, Kim JH, Park SM, Jeong YI, Lee JY, Park SW, Choi J, Kim HS,
Jeong YW, Kim S et al. 2008 The analysis of chromatin remodeling and
the staining for DNA methylation and histone acetylation do not provide
definitive indicators of the developmental ability of inter-species cloned
2007 Nuclear reprogramming in embryos generated by the transfer of yak (Bos grunniens) nuclei into bovine oocytes and comparison with bovine–bovine SCNT and bovine IVF embryos. Theriogenology 67
1331–1338.
Li Y, Li S, Zhang Q, Li R, Chen X, Fang Z, Xue K, Chen da Y & Sheng HZ
2008 Activation of human embryonic gene expression in cytoplasmic
hybrid embryos constructed between bovine oocytes and human
fibroblasts. Cloning and Stem Cells 10 297–305.
Liu SZ, Zhou ZM, Chen T, Zhang YL, Wen DC, Kou ZH, Li ZD, Sun QY &
Chen DY 2004 Blastocysts produced by nuclear transfer between chicken
blastodermal cells and rabbit oocytes. Molecular Reproduction and
Development 69 296–302.
rescue of an endangered mammal by cross-species nuclear transfer using
Lorthongpanich C, Laowtammathron C, Chan AW, Ketudat-Cairns M &
Parnpai R 2008 Development of interspecies cloned monkey embryos
reconstructed with bovine enucleated oocytes. Journal of Reproduction
and Development 54 369–373.
Maddox-Hytell P, Svarcova O & Laurincik J 2007 Ribosomal RNA and
nucleolar proteins from the oocyte are to some degree used for
embryonic nucleolar formation in cattle and pig. Theriogenology 68
(Supplement 1) S63–S70.
Mancini M, Anderson BO, Caldwell E, Sedghinasab M, Paty PB &
Hockenbery DM 1997 Mitochondrial proliferation and paradoxical
membrane depolarization during terminal differentiation and apoptosis
in a human colon carcinoma cell line. Journal of Cell Biology 138
449–469.
Mastromonaco GF, Favetta LA, Smith LC, Filion F & King WA 2007 The
influence of nuclear content on developmental competence of gaur (Bos
antelopes) in vitro fertilized and somatic cell nuclear transfer embryos.
Biological Reproduction 76 514–523.
McKenzie M & Trounce IA 2000 Expression of Rattus norvegicus mtDNA in
Mus musculus cells results in multiple respiratory chain defects. Journal of
Biological Chemistry 275 31514–31519.
McKenzie M, Chiotis M, Pinkett CA & Trounce IA 2003 Functional
respiratory chain analyses in murid xenomitochondrial cybrids expose
coevolutionary constraints of cytochrome b and nuclear subunits of
complex III. Molecular Biology and Evolution 20 1117–1124.
Memelli E & First NL 1998 Developmental changes in RNA polymerase II in
bovine oocytes, early embryos, and effect of α-amanitin on embryonic
Minami N, Suzuki T & Tsukamoto S 2007 Zygotic gene activation and
maternal factors in mammals. Journal of Reproduction and Development
53 707–715.
Mtango NR, Harvey AJ, Latham KE & Brenner CA 2008 Molecular control
of mitochondrial function in developing rhesus monkey oocytes and
preimplantation-stage embryos. Reproduction, Fertility, and Development
20 846–859.
Oback B, Wiersma AT, Gaynor P, Laible G, Tucker FC, Oliver JE,
Miller AL, Troskie HE, Wilson KL, Forsyth JT et al. 2001 Cloned cattle
derived from a novel zona-free embryo reconstruction system. Cloning
and Stem Cells 5 3–12.
Plante I, Plante C, Shepherd DL & King WA 1994 Cleavage and 3H-uridine
incorporation in bovine embryos of high in vitro developmental potential.
Reers M, Smith TW & Chen LB 1991 Aggregate formation of a
carboxycyanine as a quantitative fluorescent indicator of membrane


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