Elucidating the identity and behavior of spermatogenic stem cells in the mouse testis

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Abstract

Spermatogenesis in mice and other mammalians is supported by a robust stem cell system. Stem cells maintain themselves and continue to produce progeny that will differentiate into sperm over a long period. The pioneering studies conducted from the 1950s to the 1970s, which were based largely on extensive morphological analyses, have established the fundamentals of mammalian spermatogenesis and its stem cells. The prevailing so-called Asingle (Aₚ) model, which was originally established in 1971, proposes that singly isolated Aₚ spermatogonia are in fact the stem cells. In 1994, the first functional stem cell assay was established based on the formation of repopulating colonies after transplantation in germ cell-depleted host testes, which substantially accelerated the understanding of spermatogenic stem cells. However, because testicular tissues are dissociated into single-cell suspension before transplantation, it was impossible to evaluate the Aₚ and other classical models solely by this technique. From 2007 onwards, functional assessment of stem cells without destroying the tissue architecture has become feasible by means of pulse-labeling and live-imaging strategies. Results obtained from these experiments have been challenging the classical thought of stem cells, in which stem cells are a limited number of specialized cells undergoing asymmetric division to produce one self-renewing and one differentiating daughter cells. In contrast, the emerging data suggest that an extended and heterogeneous population of cells exhibiting different degrees of self-renewing and differentiating probabilities forms a reversible, flexible, and stochastic stem cell system as a population. These features may lead to establishment of a more universal principle on stem cells that is shared by other systems.

Morphological basics of mouse spermatogenesis and its stem cells

Mouse spermatogenesis occurs in seminiferous tubule of the testis. Fig. 1A and B illustrate a simplified architecture of the seminiferous tubules (Russell et al. 1990, Yoshida 2010). The anatomical framework of the tubules consists of the peripheral basement membrane and two types of somatic cells, i.e. Sertoli and peritubular cells. Sertoli cells comprise an epithelium inside the basement membrane, whose tight junctions provide an anatomical basis of blood–testis barrier, while peritubular cells cover the outside of the basement membrane. All the stages of germ cells from stem cells to spermatozoa are located between and nourished by Sertoli cells (Fig. 1B and C). Spermatogonia, which are defined as mitotic stages of spermatogenic cells including stem and differentiating cell types, occupy the basal compartment between basement membrane and the Sertoli cells’ tight junction.

Characteristically, spermatogonia form syncytia between their mitotic sisters due to an incomplete cytokinesis, which causes the synchronization of the subsequent divisions. As a result, spermatogonia are typically found as single isolated cells (designated as Aₚ) or syncytia of 2ⁿ (2, 4, 8, 16, etc.) cells. The syncytia continue to extend along with subsequent course of differentiation. Spermatogonia of mouse and other rodent are often classified into two: the so-called ‘undifferentiated’ and ‘differentiating’ spermatogonia. ‘Undifferentiated’ spermatogonia (sometimes designated as AUnd) consist of AUnd, Aₚaired (Aₚ, syncytia of two cells), and Aₚaired (Aₚaired, syncytia of 4, 8, 16, and occasionally 32 cells). ‘Differentiating’ spermatogonia are derived from ‘undifferentiated’ spermatogonia and proceed highly organized differentiation process and include A₁, A₂, A₃, A₄, In, and B spermatogonia, showing synchronous division accompanying the extension of the syncytia. The term ‘undifferentiated’ may be misleading because formation and extension of spermatogonial syncytia are

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considered as an important signature of differentiation (de Rooij & Russell 2000). However, this term will be used in this review not only because no other terms correctly indicate this collective entity but also because this population seems to retain potential of self-renewing and formation of syncytia does not indicate irreversible commitment for differentiation (discussed later). When spermatogonia enter meiosis, large syncytia of germ cells – now designated as spermatocytes – translocate across the tight junction into adluminal compartment. After meiosis, haploid round and then elongating spermatids form and mature into spermatozoa as the cells move towards the lumen. Clearly, stem cells comprise a subpopulation of spermatogonia in the basal compartment, but elucidation of their identity and behaviors has long been a big challenge.

Spermatogenic stem cell models

Several models have been proposed with respect to the identity and behavior of stem cells, essentially on the basis of intensive morphological investigations of fixed specimens (for review Russell et al. (1990), Meistrich & van Beek (1993) and de Rooij & Russell (2000)). Among them the so-called ‘A model’ became prevailing, which proposed that singly isolated A spermatogonium is the only cell type that act as stem cells (Fig. 2A). Whereas, interconnected syncytia of A_pr and A_al, as well as more matured differentiating spermatogonia, are considered no longer capable of self-renewal and irreversibly committed for differentiation (Huckins 1971, Oakberg 1971). A second model hypothesized that some spermatogonial syncytia are fragmented and replenish the stem cell pool, based on the presence of syncytia with the cell numbers of other than 2^n at a higher frequency after irradiation (Erickson 1978).

However, it was theoretically impossible to obtain a final conclusion solely based on the morphology of spermatogonia. First, precise judgment of spermatogonial interconnection in whole-mounted seminiferous tubule specimen (Clermont & Bustos-Obregon 1968) is difficult because intercellular bridges are not always visible under microscope, and identification of syncytia was practically based on the distance between nuclei with similar appearance that reflect their cell cycle phase (de Rooij & Russell 2000). More fundamentally, what one needs to know more is the behavior of spermatogonia over time (i.e. proliferation, differentiation, death, etc.) in order to fully understand the stem cell system.

In the field of mammalian spermatogenesis, the syncytia of spermatogonia have often been called a ‘clone’, which by definition indicates a group of cells derived from a single ancestor cell. This term is appropriate if we rely on the A model. However, this may be confusing if one considers fragmentation of a syncytium, which gives rise to multiple syncytia that belong to a single ‘clone’. Indeed, such fragmentations have recently been observed by live imaging (Nakagawa et al. 2010). Our recent literature used ‘cysts’, which are widely used for germline of Drosophila and sometimes for other animals including mammalians (Nakagawa et al. 2010). However, in Drosophila, syncytia of spermatogonia are encapsulated by somatic cells, while ‘cysts’ are used for this entire structure of spermatogonia and the surrounding somatic cells (called cyst cells). Given that mammalian spermatogonia are not encapsulated, I realize that ‘cyst’ may also be confusing. Therefore, ‘syncytia’ or ‘spermatogonial syncytia’ will
Heterogeneous composition of ‘undifferentiated’ spermatogonia in their gene expression

‘Undifferentiated’ spermatogonia, which include \( A_s \), \( A_{pr} \), and \( A_{al} \), are anatomically heterogeneous, with regards to the number of cells within individual syncytia (Fig. 2A). However, they share a very similar characteristic nuclear morphology that is distinct from that of differentiating spermatogonia, i.e. minimal heterochromatin condensation (Chiarini-Garcia & Russell 2001, 2002), suggesting that this entity of cells (\( A_s \), \( A_{pr} \), and \( A_{al} \)) shares some important features. Indeed, undifferentiated spermatogonia are characterized by unique gene expression, i.e. \( Plzf^{pos} / E-cadherin^{neg} / Kit^{neg} \). This is distinct from ‘differentiating’ spermatogonia, which are \( Plzf^{neg} / E-cadherin^{pos} / Kit^{pos} \) (Schrans-Stassen et al. 1999, Buaas et al. 2004, Costoya et al. 2004, Tokuda et al. 2009). However, expression of some genes such as \( Gfra1 \) (also known as \( GFRA1 \)), \( Nanos2 \), and \( Neurog3 \) (also known as \( Ngn3 \) or \( Neurogenin3 \)) exhibits internal heterogeneity in undifferentiated spermatogonia population (Yoshida et al. 2004, Hofmann et al. 2005b, Tokuda et al. 2007, Suzuki et al. 2009, Zheng et al. 2009, Nakagawa et al. 2010, Yoshida 2010), as shown in a simplified manner in Fig. 2B. The \( Gfra1^{pos} / Nanos2^{pos} \) (\( Neurog3^{neg} / Nanos3^{low} / E-cadherin^{pos} / Plzf^{pos} / Kit^{neg} \)) population contains a majority of the \( A_s \) and \( A_{pr} \) spermatogonia, while the \( Neurog3^{pos} \) population (\( Gfra1^{neg} / Nanos2^{neg} / Nanos3^{high} / E-cadherin^{pos} / Plzf^{pos} / Kit^{neg} \)) largely corresponds to the \( A_{al} \) spermatogonia. Interestingly, this heterogeneity is different from morphological heterogeneity, namely the number of cells within a syncytium. Importantly, \( A_s \) spermatogonia population, which has been generally considered to be uniformly the stem cells, was shown to be a heterogeneous entity: while many of them are \( Gfra1^{pos} / Nanos2^{pos} / Neurog3^{neg} \), some are \( Gfra1^{neg} / Nanos2^{neg} / Neurog3^{pos} \).

It is also likely, however, that the above-mentioned heterogeneity of gene expression is not the only heterogeneity in the \( A_{undiff} \) population. For example, interestingly, \( Id4 \) is reported to be expressed only in \( A_s \) spermatogonia (Oatley et al. 2011). Accumulation of gene expression pattern would reveal more detailed heterogeneity in this population.

Transplantation: functional assessment of spermatogenic stem cells

Experiments such as artificial cryptorchid testes and testes of vitamin A-deficient animals (Nishimune et al. 1978, van Pelt & de Rooij 1990, Hogarth & Griswold 2010) had functionally indicated that ‘undifferentiated’ spermatogonia population is responsible for the stem cell activity: in these situations, spermatogenesis arrests and only undifferentiated spermatogonia were observed, which is enough to reestablish complete...
spermatogenesis after testes are relocated back into scrotum or dietary vitamin A is restored.

Then, post-transplantation repopulation of spermatogenesis established by Brinster and colleagues opened the door for the functional and quantitative assay of spermatogenic stem cells (Brinster & Avarbock 1994, Brinster & Zimmermann 1994). Like hematopoiesis, self-renewing, and differentiating, abilities of a single stem cell became assayable by direct measurement of the post-transplantation colony formation. Using this experimental system, it was rediscovered that a vast majority of the stem cell activity resides in Kit<sup>pos</sup> ‘undifferentiated’ spermatogonia (Shinohara et al. 2000, Ohbo et al. 2003). Sorted fractions of Kit<sup>pos</sup> ‘differentiating’ spermatogonia (A<sub>1</sub> to A<sub>4</sub>, In and B) also showed some low activity of post-transplantation colony formation (Shinohara et al. 2000, Ohbo et al. 2003, Barroca et al. 2009). The origin of this activity is interesting, while the purity of the fractions warrants careful evaluation.

In undifferentiated spermatogonia, transplantable stem cell activity was detected both in Gfra1<sup>pos</sup> and in Gfra1<sup>neg</sup> (likely to be Neurog3<sup>pos</sup>) fractions (Hofmann et al. 2005a, Nakagawa et al. 2007, Grisanti et al. 2009). It is noteworthy that in vitro culture that retains the stem cell activity, another big breakthrough in this field, has been developed taking advantage of this transplantation-based stem cell assay (Kanatsu-Shinohara et al. 2003, Kubota et al. 2004).

On transplantation, the donor testes are dissociated into single cell suspensions, making it impossible to link the morphological classification of spermatogonia (i.e. A<sub>u</sub>, A<sub>pu</sub>, or A<sub>al</sub>) to their stem cell activity. In addition, the host germ cells need to be depleted before transplantation. Therefore, while transplantation reveals the ability of self-renewal, it may not reflect the stem cell functionality in the steady-state spermatogenesis that undergoes in undisturbed testes. Indeed, the same donor cells give rise to much larger number of colonies when transplanted into young hosts, indicating that detection of stem cell potential is context-dependent (Shinohara et al. 2001). Therefore, while it is no doubt that transplantation and in vitro cultures have provided important implications for the stem cell behavior in vivo, methods that enable to analyze the stem cell function under normal tissue architecture have also been warranted.

Pulse chase and live imaging: stem cell analysis in undisturbed testes

In principle, two types of experiments are capable of analyzing particular cells’ behavior in undisturbed tissues or organs. First is the ‘pulse-chase’ experiment, in which irreversible and inheritable label is introduced to the cells of interest followed by detection of labeled cells after a certain period of time (Fig. 3). If stem cells are pulse labeled, the label is expected to persist in both the stem and the differentiating descendants for prolonged period (Fig. 3B). On the other hand, if transit amplifying cells are induced, the label will be observed in differentiating progeny only transiently (Fig. 3C). Several kinds of genetic labeling technology have been developed mainly in mice and fruitflies (Fox et al. 2008, Buckingham & Meilhac 2011). In mice, tamoxifen-inducible Cre recombinase–LoxP system is the most widely used system, which recombines the reporter transgene integrated in the genome for permanent labeling on administration of tamoxifen. Potentially, unexpected effects of tamoxifen (or its active metabolite 4-OH tamoxifen) could disturb the steady state: we have evaluated that the dose used in our experiments does not affect the steady state spermatogenesis (Nakagawa et al. 2007 and data not shown).

Secondly, a more direct method of cell fate analysis is live-imaging. GFP and other fluorescent proteins have enabled visualization of the cells of interest in living organisms when expressed under an appropriate promoter. Thus, filming of living cells’ action in living animals became a realistic challenge. In 2007, our group established an in vivo live-imaging system for GFP-labeled spermatogonia in a testis of anesthetized mice (Yoshida et al. 2007). Live imaging is quite informative: entire pedigree of a cell lineage, morphological change and movements, and cell death are only available from this technique. However, prolonged cell fate analyses (for months and sometimes for years) are necessary for the stem cell assessment, which are available only from pulse-chase experiments. So, combination of these complementary strategies is very useful. By these experiments, a number of properties of mouse undifferentiated spermatogonia have been uncovered in recent years (Nakagawa et al. 2007, 2010, Yoshida et al. 2007, Sada et al. 2009).

Stem cells supporting the steady state

To dissect the mouse spermatogenic stem cell system, pulse-labeling experiments have been applied for Nanos2<sup>pos</sup> and Neurog<sub>3</sub><sup>pos</sup> subpopulations of undifferentiated spermatogonia, using transgenic mice expressing tamoxifen-inducible Cre under the control of Nanos2 and Neurog3 regulatory sequences (Nakagawa et al. 2007, 2010, Sada et al. 2009). Many of the labeled Nanos2<sup>pos</sup> spermatogonia persisted in the tissue and continued to produce differentiating progeny for several months, indicating the central role of Nanos2<sup>pos</sup> (also Gfra1<sup>pos</sup>) spermatogonia in maintenance of steady state (Sada et al. 2009). On the other hand, a vast majority of the labeled Neurog3<sup>pos</sup> spermatogonia proliferated and differentiated but did not stay in the tissue (Nakagawa et al. 2007, 2010). It is also supported that Nanos2<sup>neg/pos</sup> Gfra1<sup>neg</sup>/Neurog3<sup>neg</sup> spermatogonia give rise to Neurog3<sup>pos</sup> (also Nanos2<sup>neg</sup>/Gfra1<sup>neg</sup>/Nanos3<sup>high</sup>)
spermatogonia, and that a bulk of Neurog3pos spermatogonia differentiates into Kitpos differentiating spermatogonia (Sada et al. 2009, Nakagawa et al. 2010). Altogether, it appears that Gfra1pos/Nanos2pos spermatogonia mainly self-renew their population and give rise to Neurog3pos spermatogonia, which in turn mostly act as transient amplifying cells and differentiate into being Kitpos (Fig. 2C).

Given that the Gfra1pos/Nanos2pos and the Neurog3pos populations largely correspond to A4/Apr and Aal spermatogonia, respectively, this behavioral hierarchy in steady state seems to parallel the morphological hierarchy that appeared in the classic ‘As model’ (compare Fig. 2A and C). It is noteworthy that live imaging of Neurog3pos spermatogonia directly observed the extension of spermatogonial syncytia for the first time (e.g. A4 → Aal-4 → Aal-8), which had long been believed to occur based on the snapshot analyses (Nakagawa et al. 2007, Yoshida et al. 2007).

However, some observations in undisturbed testis were not supposed by the A4 model (Fig. 2D). First, contrary to the idea that all the A4 are equivalent and act as stem cells, Neurog3pos subset of A4 spermatogonia preferentially differentiated. Secondly, it is shown that a portion of Neurog3pos cells sometimes ‘revert’ into being Gfra1pos and act as self-renewing stem cells (Nakagawa et al. 2007, 2010). Moreover, syncytia of Neurog3pos spermatogonia occasionally fragmented into single cells and shorter syncytia (Nakagawa et al. 2010). However, it is an important question but still to be determined whether the resultant A4 cells after fragmentation of syncytial spermatogonia do replenish the stem cell pool or not.

**Regeneration is supported by different population from that acting in steady state**

It is generally considered that a single population of stem cells both supports the cell turnover in steady state and contributes to regeneration after tissue injury or transplantation. However, the lack of adequate experimental systems has prevented researchers from determining whether these cells, both can be designated as ‘stem cells’, are truly the same. Nakagawa et al. (2007, 2010) analyzed the behavior of pulse-labeled Neurog3pos spermatogonia population under three different conditions, i.e. steady state, regeneration after insult, and colony formation after transplantation (an extreme case of regeneration). The results suggest that Neurog3pos spermatogonia (mostly found in syncytia), which differentiate without self-renewal in steady state with a high probability, shift...
their mode to self-renewal in response to tissue insult or transplantation. This accompanied increased ‘reversion’ of Neurog3\textsuperscript{pos} spermatogonia into being Gfra1\textsuperscript{pos} (reversion in terms of gene expression): increased fragmentation of syncytia (reversion in terms of morphology) was also expected to occur at a high frequency, but this is yet to be examined experimentally. On the other hand, behavior of Gfra1\textsuperscript{pos} spermatogonia under regeneration or after transplantation is still to be elucidated. Grisanti et al. (2009) showed that transplantable stem cell activity of Gfra1\textsuperscript{pos} fraction is not as high as Gfra1\textsuperscript{neg} (probably Neurog3\textsuperscript{pos}) fraction, suggesting that, unlike steady state, contribution of Gfra1\textsuperscript{pos} spermatogonia in post-transplantation colony formation would not be so crucial. Systematic analyses of the Gfra1\textsuperscript{pos} population after transplantation and on regeneration will be warranted to address this important issue.

These data demonstrate that a unique population of ‘stem cells’ does not play a role in every aspect of stem cell function; rather, different cell populations are recruited into stem cell pools in different contexts. In regeneration, behaviors of the stem cell compartment become largely different from what was proposed by the ‘A, model’ and reminiscent of the ‘clone fragmentation model’, which were based on the stem cell behavior during regeneration (Erickson (1976, 1981) and Meistrich & van Beek (1993) for review). Therefore, the classical ‘A, model’ and ‘clone-fragmentation model’ appear to represent different aspects of a reversible and flexible stem cell system. Similar reversion, namely fragmentation of syncytial spermatogonia into germline stem cells, has been observed in Drosophila spermatogenesis and oogenesis (Brawley & Matunis 2004, Kai & Spradling 2004).

**Hierarchy and reversibility of the stem cell compartments**

The above-mentioned evidence clearly indicates that the mouse spermatogenic stem cell compartment includes heterogeneous cell types with different degrees of stem cell potential (i.e. probability of self-renewal vs differentiation), rather than a small population of a single kind of stem cells. Interestingly, these different cell populations are recruited to the stem cell pool in different situations, which may be beneficial for tissue homeostasis in that the stem cell pool can be recovered quickly when necessary.

The spermatogenic stem cell system involves at least two types of heterogeneity: morphology (number of connected cells within a syncytium) and gene expression (Gfra1\textsuperscript{pos}/Nanos2\textsuperscript{pos} and Neurog3\textsuperscript{pos} populations). These two features are related to each other – the longer the syncytia are, the higher the proportion of Neurog3\textsuperscript{pos} cells is – but they are not perfectly matched (Fig. 2A). Of note, both differentiation-related traits can be reversible. Then, which feature is more closely related to the regulation of cellular function, morphology, or gene expression? Data suggest that the gene expression shows a more close relationship with differentiation: in steady state, Neurog3\textsuperscript{pos} spermatogonia are clearly directed for differentiation regardless of their morphology (even A, spermatogonia that are Neurog3\textsuperscript{pos} are destined for differentiation), and Gfra1\textsuperscript{pos} cells barely become differentiating spermatogonia directly (Nakagawa et al. 2010). Neurog3\textsuperscript{pos} spermatogonia represent a unique property: while they are destined for differentiation, they clearly retain their potential of self-renewal. This is consistent with the idea of ‘potential stem cells’ that was conceptually proposed by Potten & Loeffler (1990).

On the other hand, it is no doubt that syncytium formation, which is a characteristic for germ cells widely in animal kingdom (Pepling et al. 1999), is an important property associated with differentiation. The relationship between morphology and gene expression in the stem cell self-renewal and differentiation is one of the important questions for the complete understanding of the stem cell system. With this regard, elucidation of the live behaviors of Gfra1\textsuperscript{pos} population will be very much warranted.

**Stem cell replacement in steady state**

From a classical viewpoint, every single stem cell is supposed to strictly repeat asymmetric division to form one self-renewing and one differentiating daughter, and it is carefully protected in tissue as schematically shown in Fig. 4A. The Drosophila gonads have been representing such a typical stem cell system and have provided a theoretical basis widely for stem cell research (Fuller & Spradling 2007, Losick et al. 2011, Spradling et al. 2011). On the other hand, investigation has been warranted for other systems including mammalian spermatogenesis.

A study of pulse chase of individual mouse spermaticgenic stem cell-derived cohorts over 1 year gave unexpected results (Nakagawa et al. 2007). If you assume the above-mentioned scenario of repeated asymmetric division, all the individual stem cell lineages are expected to persist over time and their size should be constant (Fig. 4A). In reality, however, a majority of stem cell lineages that had persisted over, for example, 3 months became extinct in the following months (Fig. 4B), while the surviving lineages expanded (Fig. 4C). This indicates that stem cells regularly lose their function (perhaps due to differentiation of all the descendants) and are replenished by stem cells that were born from neighboring stem cell cohorts (Figs 3D and 4D). Therefore, although spermatogenesis is a highly stable process, its stem cells are continuously turning over with each other. Mathematical analyses suggested...
that stem cells have an average longevity of <2 weeks and that such a frequent replacement occurs stochastically (Klein et al. 2010). The stochastic features of stem cell replacement obtained from this macroscopic lineage tracing may suggest that the behavior of individual stem cells is equipotent regardless of whether they are ‘bona fide’, ‘copied’, or ‘reverted’ stem cells. However, this does not mean that individual stem cells are completely the same and behave independently from their microscopic history and local situations. It is one of the most interesting and the biggest challenges to link such macroscopic scale of population dynamics of stem cells with microscopic scale of behaviors of individual spermatogonia.

**Flexibility and stochasticity may be common features of stem cells**

The above-mentioned findings may shift our view for the mouse spermatogenic stem cells. ‘Stem cells’ may not be a rare entity of specialized cells that always undergoes asymmetric division, but may be a population of cells showing flexible and stochastic features. Similar characteristics are recently emerging from other stem cell systems, too. The behavior of pulse-labeled stem cells in mammalian skin and intestinal epithelia revealed essentially the same class of mathematical signatures, suggesting that stochasticity commonly underlies the stem cell systems (Clayton et al. 2007, Doupe et al. 2010, Lopez-Garcia et al. 2010, Snippert et al. 2010, Klein & Simons 2011, Simons & Clevers 2011).

Interestingly, similar flexible stem cell behavior is also suggested in *Drosophila* germline stem cells, which was classically considered as a ‘stereotypic’ system. *Drosophila* spermatogenic stem cells turn over with age, which was shown by pulse-chase experiment (Wallenfang et al. 2006). In addition, recent success of long-term live imaging of testis organ culture visualized interesting behaviors of stem cell daughters (Sheng & Matunis 2011): although a stem cell always divided in a geographically asymmetric manner, namely perpendicular to the niche (hub), as had been observed (Yamashita & Fuller 2005, Losick et al. 2011, Spradling et al. 2011), the subsequent fates of daughter cells were not always asymmetric. According to Sheng and Matunis (2011), ~7 and ~13% of stem cell divisions produced daughter pairs that both self-renew (symmetric renewal) and daughter pairs that both differentiate (symmetric differentiation) respectively. The probabilities of symmetric renewal and symmetric differentiation were increased and decreased, respectively, and the reversion from syncytia also takes place when regeneration was induced. These altogether result in the recovery of stem cell pool after tissue insult. It may be an issue of semantics whether this stem cell replacement is an exceptional event or whether it represents regular turnover. The observed frequency looks high enough to support the latter, although the possibility of

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**Figure 4** Stem cell replacement in steady-state spermatogenesis. (A) Conceptual illustration of stem cell fate based on the assumption that stem cells always undergo strict asymmetric division and never replace with each other. Lineage of initial stem cells should be stable over time (five stem cell lineages are shown in different colors). (B and C) Chase of labeled single stem cell-derived patches in seminiferous tubules: many of the stem cell lineages become extinct (B) while others expand (C) over months. Reproduced from Klein AM, Nakagawa T, Ichikawa R, Yoshida S & Simons BD 2010 Mouse germ line stem cells undergo rapid and stochastic turnover. Cell Stem Cell 7 214–224. Reproduced with permission from Elsevier. Copyright 2010 Elsevier. (D) Scheme for stem cell replacement that explains the actual pulse-chase data shown in (B) and (C).
an artifact caused by ex vivo cultivation needs to be assessed carefully. I think that mathematical analysis of pulse-labeled stem cell cohorts in vivo is warranted in this system as well.

Remaining questions: spatio-temporal regulation of the stem cells

Stem cells’ behavior is regulated properly in the seminiferous tubules. Stem cell niche, which is generally defined as the microenvironment that regulates stem cell function, is considered as an indispensable element of the stem cell system (Morrison & Spradling 2008). Mouse seminiferous tubules do not represent clearly specialized niche structure cells unlike Drosophila testes (hub cells) and ovaries (cap cells), or mouse intestine (base of the crypts). This is why the study of the mouse spermatogenic stem cell niche has been based on the localization of Aundiff spermatogonia. Thanks to precise and insightful observations of testis sections, Chiarini-Garcia et al. discovered that these Aundiff spermatogonia preferentially localize to the area adjacent to the interstitium between seminiferous tubules (Chiarini-Garcia et al. 2001, 2003). Subsequently, three-dimensional reconstruction of serial sections of the tubules confirmed this biased localization of Aundiff spermatogonia to the blood vessels and accompanying interstitial cells (Yoshida et al. 2007). In addition, live imaging visualized that when Neurog3PH spermatogonia transit into A1 spermatogonia they depart this presumptive vasculature-associated niche region and spread to all over the basal compartment (Yoshida et al. 2007). These region-related distributions and movements of spermatogonia clearly indicate that seminiferous tubules, although seemingly uniform, comprised multiple functional compartments in a manner related to the vasculature pattern. In addition, alteration of vasculature pattern accompanies the reorganization of spermatogonia, suggesting that the blood vessels (and/or their associated cells) determine the niche region. Given that blood vessels and interstitial cells do not have direct contact with spermatogonia, some molecular mechanisms should link the vasculature/interstitium with the niche microenvironment that is proximal to the stem cells (Yoshida 2011). One of such molecules may be Csfr1, which are expressed in interstitial cells and a subset of peritubular cells and increase the colony-formation activity in cultured spermatogonia (Oatley et al. 2009). However, the molecular and cellular nature of this specialized microenvironment remains to be mostly elucidated.

Another unsolved question is related to the seminiferous epithelial cycle: differentiation process from A1 spermatogonia to mature sperm, which takes place 35 days, progresses in a periodical manner with the interval of 8.6 days. Sertoli cells also change their functions cyclically so that they can support the different stages of germ cells properly (Oakberg 1956, Russell et al. 1990). The stem cell behaviors seem to be also related to this cyclic phenomenon (de Rooij 1998), suggesting that some cycle-related environmental cues regulate the undifferentiated spermatogonia. Of note, expression of GDNF, the ligand for Gfra1, shows an oscillating pattern along with the seminiferous epithelial cycle (Johnston et al. 2011, Sato et al. 2011, Grasso et al. 2012). It is also suggested that local retinoic acid concentration oscillates along with the cycle and induce transition from ‘undifferentiated’ to A1 differentiating spermatogonia at stages VII and VIII (Sugimoto et al. 2012). In addition, retinoic acid also acts on Sertoli cells so that germ cell differentiation and Sertoli cells are well coordinated. Elucidation of the role of these and other cycle-related environmental factors on the stem cell regulation remains as an open question.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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