The expression of fibroblast growth factor receptors during early bovine conceptus development and pharmacological analysis of their actions on trophoblast growth in vitro

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Abstract

The overall aim of this work was to examine the expression profiles for fibroblast growth factor receptors (FGFRs) and describe their biological importance during bovine pre- and peri-implantation conceptus development. FGFR1 and FGFR2 mRNAs were detected at 1-, 2-, 8-cell, morula and blastocyst stages whereas FGFR3 and FGFR4 mRNAs were detected after the 8-cell stage but not earlier. The abundance of FGFR1, FGFR3, and FGFR4 mRNAs increased at the morula and blastocyst stages. Immunofluorescence microscopy detected FGFR2 and FGFR4 exclusively in trophoblast cells whereas FGFR1 and FGFR3 were detected in both trophoblast cells and inner cell mass in blastocysts. Neither transcripts for FGF10 nor its receptor (FGFR2b) were temporally related to interferon τ (IFNT) transcript profile during peri- and postimplantation bovine conceptus development. A series of studies used a chemical inhibitor of FGFR kinase function (PD173074) to examine FGFR activation requirements during bovine embryo development. Exposing embryos to the inhibitor (1 mM) beginning on day 5 post-fertilization did not alter the percentage of embryos that developed into blastocysts or blastocyst cell numbers. The inhibitor did not alter the abundance of CDX2 mRNA but decreased (P < 0.05) the relative abundance of IFNT mRNA in blastocysts. Exposing blastocysts to the inhibitor from days 8 to 11 post-fertilization reduced (P < 0.05) the percentage of blastocysts that formed outgrowths after transfer to Matrigel-coated plates. In conclusion, each FGFR was detected in bovine embryos, and FGFR activation is needed to maximize IFNT expression and permit outgrowth formation.

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Introduction

Fibroblast growth factors (FGFs) encompass a large class of autocrine, paracrine, endocrine, and intracrine factors. More than 20 genes encode FGFs in mammals, including bovids (Elsik et al. 2009, Zimin et al. 2009, Oulion et al. 2012). The FGFs interact with a group of tyrosine kinase receptors encoded by four genes in mammals (termed FGFR1/2/3/4). Alternative splicing events observed in several species generate an array of receptor isotypes with disparate ligand binding and signaling capabilities (Powers et al. 2000, Bottcher & Niehrs 2005). One predominant splicing event occurs within the third IgG-like extracellular domain in three of these receptors (FGFR1/2/3), and the excision of one or two exons generates receptor subtypes that recognize different FGFs in various mammals, including ruminants (Johnson et al. 1991, Chen et al. 2000, Elsik et al. 2009, Zimin et al. 2009).

Several actions for FGFs have been described during embryogenesis. FGFs are required for the emergence and proliferation of extraembryonic endoderm in mice, cattle, and humans (Kuijk et al. 2008, Cockburn & Rossant 2010, Yamanaka et al. 2010, Yang et al. 2011a). Maintenance of trophoblast lineage is also controlled by FGFs. One of the first described activities in this regard was the necessity for FGF4 to maintain trophoblast stem cells in an undifferentiated state in mice (Feldman et al. 1995, Tanaka et al. 1998). Not all mammals show this need for FGF4 or another FGF to stimulate trophoblast stem cell proliferation and block differentiation. Trophoblast cells from the rhesus monkey and common vole (Microtus rossiaeneridionalis) proliferate without differentiation in the absence of exogenous factors, including
FGFs (Vandevoort et al. 2007, Grigor’eva et al. 2009). Likewise, trophoblast cells derived from ungulates (cattle, sheep, and pigs) proliferate in the absence of FGFs (Talbot et al. 2000, Shimada et al. 2001, La Bonnardiere et al. 2002, Dunlap et al. 2006, Michael et al. 2006). Although exogenous FGFs are not required for trophoblast cell proliferation and survival, FGF activities exist in trophoblast cells. Several FGFs stimulate interferon-τ (IFNT) production. Trophoblast cell-derived IFNT is the maternal recognition of pregnancy hormone in ruminants (Ealy & Yang 2009). Several FGFs (e.g. FGF1, FGF2, FGF7, and FGF10) stimulate IFNT mRNA and protein production in bovine trophoblast cell lines and blastocysts through a PKC-δ-dependent mechanism (Michael et al. 2006, Cooke et al. 2009). FGF2 and FGF10 also stimulate migration of bovine and ovine trophoblast cells (Yang et al. 2011b). Collectively, these observations indicate that FGF signals influence trophoblast cell behavior as these cells rapidly proliferate, prepare for adhesion to the uterine lining, and produce IFNT.

The need for FGF signals during the initial stages of bovine embryo development is not clear. An FGF that has been examined recently is FGF2, or basic FGF. Transcripts for FGF2 and several of the FGFR subtypes that react with FGF2 have been identified in bovine embryos during early development (Daniels et al. 2000, Cooke et al. 2009). The uterine endometrium also produces FGF2 throughout the estrous cycle and early pregnancy in sheep and cattle, and immunoreactive FGF2 is detected in uterine flushes during diestrus (Michael et al. 2006, Ocon-Grove et al. 2008). In some studies, providing FGF2 during bovine embryo culture improved subsequent blastocyst formation (Larson et al. 1992, Neira et al. 2010). However, previous studies from this laboratory failed to detect positive effects of FGF2 on blastocyst formation unless large amounts of recombinant protein were provided (500–1000 ng/ml; Fields et al. 2011). The overall aim of this work was to resolve whether embryo-derived FGFs and FGFR signaling is essential during early embryo development in cattle. Specific objectives were to describe the expression of FGFRs during early and pre-/peri-implantation conceptus development in cattle and define early developmental events that require FGFR activity.

Figure 1 The relative abundance of FGFR1/2/3/4 mRNAs throughout early bovine conceptus development. Total cellular RNA was extracted from IVP bovine embryos collected either 24 h after fertilization (1-cell; presumptive zygotes) or at the 2-cell, 8-cell, morula or blastocyst stages (n = 5 replicate studies, ten embryos/time point/replicate). Quantitative RT-PCR was completed to determine the relative abundance of FGFR1/2/3/4 (A, B, C and D respectively). Data are presented relative to the abundance of the reference transcript (HIST2H2AC). Different superscripts depict differences in mRNA abundance within each panel (P<0.05).
Results

Expression profiles for FGFR1/2/3/4 during pre-implantation conceptus development

The ontogeny of FGFR expression at the initial stages of bovine embryo development has not been examined previously. Therefore, a study was completed to describe the expression profiles for each FGFR throughout early embryogenesis (Fig. 1). Abundance of HIST2H2AC mRNA was consistent from the 1-cell to blastocyst stage and was used as the endogenous control. Transcripts for FGFR1 were detected throughout early development, but the relative abundance of FGFR1 mRNA was greater ($P<0.05$) in morulae and blastocysts than 1- and 2-cell embryos (Fig. 1A). Similar levels of FGFR2 mRNA were detected throughout early development with the exception of 8-cell embryos, which contained less ($P<0.05$) mRNA than other stages (Fig. 1B). Transcripts for FGFR3 and FGFR4 were not detected in 1- and 2-cell embryos but were detected in 8-cell embryos (Fig. 1C and D). The relative abundance of FGFR3 and FGFR4 mRNA increased ($P<0.05$) in morulae and blastocysts.

Immunolocalization of FGFRs in bovine blastocysts

Immunofluorescence was used to examine the localization of each FGFR protein from in vitro-produced (IVP) embryos. Immunoreactive FGFR1, FGFR2, FGFR3, and FGFR4 could not be detected in 1-, 2-, or 8-cell embryos (data not shown), but immunoreactivity for each receptor was detected at the blastocyst stage (Fig. 2). Morulae were not examined. FGFR1 and FGFR3 proteins were detected in both the inner cell mass and the trophoblast cells whereas FGFR2 and FGFR4 localized to the trophoblast cells.

Expression profiles of selected transcripts during peri- and postimplantation periods

The association between FGF10 and IFNT was of interest because both are produced by the peri-implantation conceptus around the time when IFNT production is maximal, suggesting that FGF10 may assist in the control of peak IFNT production (Cooke et al. 2009). Also, the FGF10 receptor, termed FGFR2b (or keratinocyte growth factor receptor, a spliced variant of the FGFR2 subfamily), is produced by trophectoderm (TE) during the peri-implantation period in sheep (Chen et al. 2000), but its expression throughout early embryo development remained unexplored.

A biphasic pattern of IFNT mRNA abundance was detected throughout pre-, peri-, and post-attachment periods of bovine conceptus development (Fig. 3A). IFNT mRNA was detected in blastocysts, and mRNA abundance increased ($P<0.05$) in ovoid conceptuses collected at day 14 of pregnancy. The relative abundance...
of IFNT mRNA increased ($P<0.05$) further in elongating conceptuses collected at day 14. Maximal amounts of IFNT mRNA were detected in elongated conceptuses collected at day 17. IFNT mRNA was detected at day 21 and in TE at day 28, albeit at very low levels. No IFNT mRNA was detected in extraembryonic mesoderm (ExM) at day 28.

Both FGF10 and FGFR2b mRNAs were detected throughout all the stages of conceptus development. However, neither expression pattern mirrored that of IFNT. The transcript abundance for FGF10 was low in blastocysts and increased ($P<0.05$) in conceptuses collected at day 14 (Fig. 3B). The relative abundance of FGF10 mRNA was unchanged in ovoid and elongating conceptuses collected at day 14 and conceptuses collected at day 17 and was increased ($P<0.05$) at days 21 and 28. FGF10 mRNA was only detected in ExM at day 28. Two studies examined the expression profiles for FGFR2b. The first study examined FGFR2b mRNA abundance during the initial stages of bovine conceptus development (Fig. 3C). FGFR2b mRNA was detected in 1-, 2-, and 8-cell embryos, and its relative abundance increased ($P<0.05$) at morula and blastocyst stages. mRNA collected at subsequent stages of bovine conceptus development (Fig. 3D) determined that the relative abundance of FGFR2b mRNA remained unchanged between the blastocyst stage and day 17. An increase ($P<0.05$) in FGFR2b mRNA abundance was detected at day 21 and in TE at day 28. No FGFR2b mRNA was detected in ExM.

**FGFR signaling requirements for embryo development and IFNT expression**

The requirement of FGFR signaling during pre-implantation embryo development was examined by blocking FGFR activation with PD173074, a cell-permeable FGFR kinase inhibitor used by this laboratory and others to examine FGFR-mediated events in bovine tissues (Yang et al. 2011a, Kuijk et al. 2012, Zhang & Ealy 2012). This compound competitively inhibits ATP binding to receptor kinase regions of FGFRs (Mohammadi et al. 1998, Skaper et al. 2000). Studies have not been completed to verify the specificity of this compound to bovine FGFRs, but this specificity is probable given the high degree of sequence conservation in FGFR tyrosine kinase regions of cattle and humans (>98% amino acid sequence identity with corresponding human FGFR kinase domains). A pilot study verified that PD173074 inhibited FGF2-dependent responses in a bovine trophoblast cell line (CT1). Cells were exposed to 10 ng/ml FGF2 and various concentrations of PD173074. In non-PD173074-treated controls, FGF2 increased ($P<0.05$) IFNT mRNA abundance by 5.0 ± 1.6-fold. No significant changes in FGF2-induced IFNT mRNA abundance were detected by providing 10 or 100 nM PD173074, but exposing cells to 1 μM PD173074 for 24 h before FGF2 treatment prevented the FGF2 effect (0.81 ± 0.15-fold effect vs control).

Exposing embryos to 1 μM PD173074 from days 5 to 8 post-IVF did not affect blastocyst development rate and...
total cell numbers in blastocysts (Fig. 4A and B). Also, no changes in mRNA abundance of CDX2, a trophoblast lineage marker, were observed (Fig. 4C). However, PD173074 treatment decreased ($P < 0.05$) IFNT mRNA abundance (Fig. 4D).

**Figure 4** Inhibition of FGFR activity with PD173074 does not affect embryo development or trophoblast gene expression during in vitro culture. Bovine embryos were generated by IVP. Embryos containing $\geq 8$ blastomeres were collected at day 5 post-IVF and incubated in medium containing 1 $\mu$M PD173074 or carrier only (control). In one study, rates of blastocyst formation were determined at day 8 (A) and blastocysts were fixed and stained to determine blastomere numbers (B) ($n = 6$ replicate studies, 28–31 embryos/treatment/replicate). In another study, blastocysts ($n = 4$ replicate studies, 10–15 blastocysts/treatment/replicate) were collected and RNA was extracted at day 8 and qRT-PCR was used to determine the abundance of CDX2 (C) and IFNT (D) transcripts relative to the abundance of the reference transcript (GAPDH). The asterisk depicts differences between PD173074-treated and control samples ($P < 0.05$).

Interruption in blastocyst outgrowth development following FGFR inhibition

The need for FGFR signals after blastocyst formation was examined by assessing the ability of blastocysts to adhere to Matrigel-coated plates and form trophoblast outgrowths (Fig. 5). Individual blastocysts were incubated in growth medium containing 1 $\mu$M PD173074 or carrier only from days 8 to 11 post-IVF, then medium was exchanged, and blastocysts were cultured without treatments thereafter. In controls, trophoblast outgrowths were evident in a low proportion of blastocysts (<3%) as early as day 10. Additional outgrowths were detected thereafter until day 15 (Fig. 5A). No additional outgrowths were formed after day 15. Exposure to PD173074 delayed ($P < 0.05$) the time when outgrowth development was first observed (day 12). The treatment also decreased ($P < 0.05$) the percentage of blastocysts that formed outgrowths at each day of examination (Fig. 5A). A large variation in outgrowth size (i.e. surface area) was evident in both treatments (Fig. 5B). A numerical reduction in outgrowth size was observed between controls and PD173074-treated blastocysts, but this outcome was not statistically significant ($P = 0.15$).

**Discussion**

The first major focus of this work was to identify stages when bovine embryos are potentially responsive to FGFs by describing the ontogeny of FGFR mRNA and protein production during bovine embryogenesis. Differences in

**Figure 5** Exposing blastocysts to PD173074 compromises subsequent outgrowth formation. Bovine embryos were generated by IVP. Blastocysts were collected at day 8 post-IVF and incubated in individual wells of a 24-well plate containing Matrigel basement membrane matrix with medium containing 1 $\mu$M PD173074 or carrier only (control; $n = 6$ replicate studies, 10–14 blastocysts/treatment/replicate). At day 11 post-IVF, medium was exchanged with medium lacking PD173074. The incidence of blastocyst outgrowth formation was examined daily until day 15 (A). Size of outgrowths was determined at day 15 (B). Asterisk (*) indicates differences in outgrowth formation due to treatment within day ($P < 0.05$).
expression profiles exist between FGFR subtypes. Of particular note is the presence of FGFR1 and FGFR2 transcripts throughout early embryonic development vs the presence of FGFR3 and FGFR4 transcripts only after embryonic genome activation (8–16-cell stage in bovine embryos; Telford et al. 1990). Transcripts found in embryos before embryonic genome activation are synthesized by the maternal genome before ovulation and stored in the cytoplasm during fertilization and the first several days of embryogenesis (Gilchrist et al. 2008). Therefore, both maternal and embryonic FGFR1 and FGFR2 transcripts exist in the bovine embryo whereas only embryonic transcripts were detected for FGFR3 and FGFR4. The loss of FGFR2 mRNA at the 8-cell stage likely reflects the use and degradation of maternal transcripts before embryonic transcription begins. Transcripts for FGFR1, FGFR3, and FGFR4 exhibited marked increases in their relative abundance at the blastocyst stage in cattle when compared with their abundance at the 8-cell stage. The pattern of FGFR2 expression was different in this regard, and no dramatic changes in overall transcript abundance occurred after embryonic genome activation.

It is unclear what roles FGFR1 and FGFR2 may be playing in early bovine embryo development. The major spliced variants for these receptors react with numerous FGFs, including FGF2 and FGF10 (Powers et al. 2000, Bottcher & Niehrs 2005), and their presence likely contributes to the embryotrophic activities for FGF2 during early bovine embryogenesis (Larson et al. 1992, Neira et al. 2010). Interestingly, the necessity for FGFR1 and FGFR2 activity is not evident until after blastocyst formation in mice. Early embryogenesis occurs normally in knock-out models of each receptor, and loss of FGFR1 function induces a postimplantation embryonic lethal phenotype characterized by defects in mesoderm and endoderm migration and cell fate determination (Deng et al. 1994, 1997, Yamaguchi et al. 1994) whereas loss of FGFR2 function generates a post-blastocyst stage embryonic lethality characterized by lack of visceral endoderm and reduced TE mass (Arman et al. 1998). Epiblast-derived FGF4 interacting with the FGFR2c-spliced variant is responsible for these activities. Variants of FGFR2 also appear important for conceptus development in ruminants. FGF2, which reacts with FGFR2c and several other FGFRs, is produced by the endometrial epithelium and peak secretion occurs around the time of conceptus elongation in sheep (Michael et al. 2006, Ocon-Grove et al. 2008). Also, the major FGFR2c-spliced variant, FGFR2b, is expressed in bovine, ovine, and porcine TE before implantation and its primary ligands (FGF7 and FGF10) are produced in endometrium before and during implantation (Chen et al. 2000, Ka et al. 2001, Cooke et al. 2009, Yang et al. 2011a). In the sheep uterus, FGF10 production from the uterine stroma is controlled by progesterone and its interaction with FGFR2b in the luminal epithelium is associated with uterine events that control early conceptus development (Chen et al. 2000, Satterfield et al. 2008). Also, immature binucleate cells contain FGFR2 transcripts, and perhaps endometrial FGF10 regulates trophoblast differentiation, migration, and hormone production (Pfarrer et al. 2006).

Not only did FGFR3 and FGFR4 mRNA profiles differ from the other FGFRs but the relative transcript abundance of these FGFRs was much greater than the other receptors. At the blastocyst stage, FGFR3 mRNA abundance was 15–300-fold greater than FGFR2 and FGFR1 respectively. Similarly, FGFR4 mRNA was 10–40-fold greater in abundance than FGFR2 and FGFR1 respectively. Neither absolute mRNA concentrations nor adjustments to the relative abundance measurements were made here; therefore, direct comparisons between these transcripts cannot be completed. However, the high expression of these transcripts suggests that these receptors may be important during early embryogenesis. It is unclear what these functions may be. FGFR3 reacts with several FGFs (FGF1 and FGF9 for FGFR3b; FGF1, FGF2, FGF4, FGF8, and FGF9 for FGFR3b; Itoh & Ornitz 2004), and it is expressed through gestation in human and rodent placenta and localizes to extraembryonic endoderm and trophoblast giant cells in mice (Rappolee et al. 1998, Antebay et al. 2005). Mutations in FGFR3 function in mice and humans are associated with skeletal dysplasia, notably achondroplasia during fetal development and uterothial tumorigenesis in adulthood (Ilyer & Milowsky 2012, Laederich & Horton 2012). No spliced variants exist for FGFR4. Numerous FGFs interact with FGFR4 (e.g. FGF1, FGF2, FGF4, FGF6, FGF8, and FGF9; Itoh & Ornitz 2004), and it is expressed throughout gestation in the mouse and human placenta (Rappolee et al. 1998, Antebay et al. 2005). In the human placenta, FGFR4 mRNA localizes to several trophoblast cell types at the decidual–trophoblast border, suggesting a role in maintaining this interface (Antebay et al. 2005). FGFR4 is also needed during neural development and is linked with various cancers in several species (Marchal et al. 2009, Ota et al. 2010, Tenhagen et al. 2012).

Detecting FGFR proteins at the blastocyst stage is consistent with the transcript profiles for FGFR3 and FGFR4 but it is not clear why immunoreactive FGFR1 and FGFR2 were not detected at earlier stages of development. Perhaps posttranscriptional events (e.g. RNA silencing) limit FGFR protein translation during early stages of embryo development. Alternatively, perhaps the sensitivity of detection was insufficient to visualize immunoreactivity at these early stages. Nonetheless, the detection of each FGFR transcript and protein at the blastocyst stage suggests that each major FGFR subtype is available to interact with FGFs at this stage. FGFR2 exhibits a trophoblast-specific localization pattern in mice, pigs, and sheep (Haffner-Krausz et al. 1999, Chen et al. 2000, Ka et al. 2007). This work demonstrated the same expression profile in cattle.
The localization patterns of other FGFRs have not been characterized previously at blastocyst stage. These outcomes indicate that FGFR4 localizes solely to the trophoblast cell in blastocysts whereas trophoblast and inner cell mass cells express FGFR1 and FGFR3.

The expression profile for FGFR2b and one of its ligands, FGF10, was examined throughout the pre-, peri-, and postimplantation stages in cattle to examine a potential linkage between this FGF ligand/receptor system and peak IFNT expression in peri-attachment conceptuses. Recent work from this laboratory detected an increase in the relative abundance of embryonic IFNT mRNA around the time of bovine conceptus elongation (Cooke et al. 2009), thereby implicating FGF10 as a potential mediator of the rapid surge in IFNT production during elongation. However, work presented herein found no correlations in expression patterns. The IFNT mRNA profile was identical to that reported by others (Kubisch et al. 1998, Ealy et al. 2001, Robinson et al. 2006). A pronounced increase in IFNT mRNA occurred with elongation, and maximal transcript abundance was evident at day 17 whereas the FGF10 mRNA profile remained unchanged around the time of conceptus elongation (days 14–17). Also, FGFR2b mRNA abundance remained unchanged during conceptus elongation. Although FGF10 and FGF2b do not appear to be principle players defining the pattern on IFNT expression, this and other FGFs certainly appear to play a role in regulating the constitutive production of IFNT in ruminant TE (Michael et al. 2006, Cooke et al. 2009). Also, work presented herein found that blocking FGFR kinase activity reduces IFNT mRNA abundance. No changes in CDX2 mRNA, a trophoblast lineage marker, were noted, indicating that the number and overall transcriptional activity of trophoblast cells is not altered with this inhibitor. Rather, this outcome supports the hypothesis that blastocyst-derived FGFs (e.g. FGF2 and FGF10) mediate constitutive IFNT production in cattle. With that said, to date, no studies have directly examined the necessity for FGFR signals during peak IFNT expression in ruminants. Work at this critical stage of pregnancy is needed before we can claim whether FGFR signaling is required for maximal IFNT production or if these signals are only being used before the peak in IFNT production at days 14–17 in cattle.

The second major focus of this work was to describe functions for FGFRs during early embryonic development. The necessity for specific FGFRs has not been described in the cow, but as described previously, loss-of-function mutation studies on mice determined that FGFR1 and FGFR2 are vital for post-blastocyst survival and FGFR3 and FGFR4 are important after implantation (Deng et al. 1994, Yamaguchi et al. 1994, Arman et al. 1998, Iyer & Milowsky 2012, Laederich & Horton 2012). Present work failed to detect any adverse developmental effects by inhibiting FGFR activity before blastocyst formation. A previous study using another FGFR kinase domain inhibitor (SU5402) observed that fewer cultured embryos formed blastocysts at day 7 but not at day 8 post-fertilization (Fields et al. 2011). Blastocyst formation at day 7 was not examined in this project, but no differences in blastocyst formation, blastomere numbers, and the expression of CDX2 were detected at day 8 post-fertilization in embryos exposed to PD173074. Therefore, FGFR inhibition seems to impact the timing of blastocyst formation but not overall blastocyst formation, embryonic numbers, and lineage commitment to trophoblast cells. It remains possible that FGFs function, at least in part, as cooperative factors during early embryonic development in cattle. One recent study found that the positive effect of FGF2 supplementation on in vitro bovine blastocyst development was enhanced when it was co-supplemented with other factors, such as insulin-like growth factors or colony-stimulating factor 2 (Neira et al. 2010). Another study failed to detect a benefit of FGF2 supplementation on in vitro bovine embryo development unless it was co-cultured with transforming growth factor β ( Larson et al. 1992).

The last study examined potential impacts of inhibiting FGFR signals after blastocyst formation. Bovine blastocysts attach to basement membrane matrices, such as Matrigel, when cultured in medium containing serum (Talbot et al. 2007, Yang et al. 2011a). Exposing blastocysts to the FGFR inhibitor delayed the onset of outgrowth formation and reduced the overall rate of outgrowth formation. The size of the outgrowths at day 15 was not statistically different between groups. This is not surprising as so few outgrowths were established after exposure to the FGFR inhibitor. Observing such small outgrowths at day 15 could reflect that FGFR inhibition compromised trophoblast cell growth potential. Alternatively, observing smaller outgrowths at day 15 could simply reflect the delayed ontogeny of outgrowth formation. FGF2 and FGF10 stimulate bovine and ovine trophoblast cell migration across trans-well inserts (Yang et al. 2011b). Therefore, FGFR signaling may mediate the chemotactic, adhesive, migratory, and/or proliferative activities required for changes in trophoblast cell morphology, adhesion and attachment, and conceptus elongation during peri-implantation development. However, it remains untested whether post-blastocyst stage development is compromised by FGFR signal interference in utero.

One aspect of the outgrowth work that remains unresolved involves pinpointing the precise timing for FGFR signal control of outgrowth formation. Temporary reductions in FGFR protein abundance have been noted by others following PD173074 treatment (Azuma et al. 2011, Lamont et al. 2011). If similar PD173074-induced reductions in FGFR expression also occur in bovine blastocysts, then these embryos may be unresponsive to endogenous FGF signals after removal of the inhibitor at day 11 post-IVF. FGF mRNA profiles and FGFR activity assessments were not examined during and after
PD173074 treatment. Therefore, it is not known whether the effects observed resulted from FGFR signal inhibition between days 8 and 11 post-IVF or if continued inhibition of FGFR activity after day 11 was responsible for the outcomes.

In summary, these observations provide new insights into the necessity for FGFR signaling during early embryo development in cattle. Transcripts for FGFR1 and FGFR2 were detected before embryonic genome activation and all four FGFR subtypes were detected after genome activation. Immunoreactive proteins for each FGFR localized to trophoblast cells at the blastocyst stage. FGFR1 and FGFR3 proteins were detected in the inner cell mass in blastocysts. Interference of FGFR signaling did not impact development to the blastocyst stage and blastomere numbers in blastocysts. However, this report identified that FGFR activity mediates IFNT expression in blastocysts and maximizes blastocyst outgrowth formation and spreading. This work implicates embryo-derived FGFs and FGFR signaling as important components for IFNT production and trophoblast cell adhesion in cattle.

Materials and Methods

In vitro embryo production

Oocyte maturation, fertilization, and embryo culture was completed as described previously (Loureiro et al. 2009, Zhang et al. 2010, Fields et al. 2011). In brief, ovaries were obtained from a local slaughterhouse (Central Beef Industries, Center Hill, FL, USA). Pools of COCs (n=10–20) were collected and matured in 50 μl drops of oocyte maturation medium (OMM; TCM199 containing Earle’s salts (Invitrogen Corp.), 25 μg/ml porcine FSH (Follitropin-V, Bioniche Life Sciences, Belleville, ON, Canada), 2 μg/ml estradiol 17β, 0.2 mM sodium pyruvate, 50 μg/ml gentamicin sulfate, 1 mM glutamine, and 1 mg/ml polyvinyl alcohol (Sigma–Aldrich)). After 21–22 h, COCs were placed in fertilization medium (TL-Fert (Caisson, Sugar City, ID, USA) containing 0.5% (w/v) essentially fatty acid-free BSA and 1 mM Na pyruvate) and incubated with Percoll gradient-purified bovine spermatozoa for 8–10 h at 38.5 °C (5% CO2 in humidified air). A pool of semen from four bulls was used for all studies. After 8 h of fertilization, cumulus cells were removed by vortexing in 1000 U/ml hyaluronidase (Sigma–Aldrich). In brief, samples were incubated with RNase–free DNase (New England Biolabs, Ipswich, MA, USA) and reverse transcribed (High-Capacity cDNA RT Kit, Applied Biosystems) and amplified with SYBR Green PCR Master Mix reagent (Applied Biosystems) and normal PCR conditions (40–50 cycles of 95 °C for 15 s and 60 °C for 1 min). Primer pairs (Table 1) were validated in previous work for adequate primer efficiency and correctness of amplicon sequence (Cooke et al. 2009, Zhang et al. 2010, Yang et al. 2011). Amplification of a single PCR product for each sample was examined by dissociation curve analysis. Also, the exclusion of reverse transcriptase was included as a negative control for each sample. Histone HIST2H2AC (HIST2H2AC) was used as the transcript reference control for studies examining changes in transcript abundance from 1-cell to blastocyst stages and GAPDH was used as the reference control for studies to day 28 of gestation. The abundance of each reference control transcript was unchanged during the specified stages of development similar to that described previously by others (Robert et al. 2011).
Table 1 Primers used for qRT-PCR.

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<tr>
<th>Gene of interest</th>
<th>Primer sequences (5’–3’)</th>
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<tr>
<td>FGF10</td>
<td>F: AGAGGACAGAAAACACGAAAGGAAA&lt;br&gt;R: GCTTACAGACCCAGTCTGCC</td>
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<td>FGFR1</td>
<td>F: TGGTACAGACCCAGTCTGCC&lt;br&gt;R: GAACTACGTCCCGAGCGCGG</td>
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<tr>
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<td>GAPDH</td>
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<td>IFNT</td>
<td>F: GATTCTTCTGAGCTGGTYG&lt;br&gt;R: GGGCGATGACAGACTTCCY</td>
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F, forward primer (sense); R, reverse primer (antisense); Y, insertion of a C or T nucleotide. Primers were validated in previous work for amplification efficiency and correctness of amplification (Cooke et al. 2009, Zhang et al. 2010, Yang et al. 2011a).

2002, Purcell et al. 2009). Data were normalized based on the ratio of transcript abundance for the gene of interest to that of the reference control (2−ΔCT gene of interest/2−ΔCT reference control).

Immunofluorescence and confocal microscopy

IVP bovine embryos were fixed with 4% (v/v) paraformaldehyde in PBS for 15 min, permeabilized for 15 min with 0.25% Triton X-100 in PBS, and blocked with PBS containing 5% (w/v) BSA. Embryos were incubated overnight at 4 °C with primary antibody (FGFR1, FGFR3, and FGFRs purchased from Abcam (Cambridge, MA, USA), FGFR2 (1:200, Abnova, Taipei City, Taiwan), FGFR3 (1:200, Abcam), and FGFR4 (1:100, ab65974, Abcam)). Each preparation was diluted in Tris-buffered saline containing 0.1% (v/v) Tween 20 (TBST) and 1% (w/v) BSA. After washing, embryos were incubated in secondary antibody (anti-mouse IgG conjugating FITC (1:1000, Abcam) or anti-rabbit IgG conjugating Alexa Fluor 555 (1:1000, Cell Signaling Technology, Danvers, MA, USA)) prepared in TBST–BSA for 1 h at room temperature. Nuclei were labeled with 4’,6-diamidino-2-phenylindole (DAPI) (1 μg/ml). Fluorescence signals were visualized with a Zeiss Pascal LSM5 Confocal Laser Scanning Microscope (Zeiss, Göttingen, Germany). Western blotting was completed with bovine cell lysates (CT1 cells and Madin-Darby bovine kidney (MDBK) cells) to verify that antibodies recognized a single protein band of the approximate molecular weight specified for each FGFR (data not shown).

FGFR inhibition during early embryo development

In one study, ≥16-cell embryos were collected at day 5 post-IVF and transferred to mSOF medium containing PD173074 (1 μM, Stemgent, Cambridge, MA, USA) or vehicle only (0.005% DMSO) (n=6 replicate studies, 28–31 embryos/treatment/replicate). At day 8, embryos were examined for their progression to the blastocyst stage, and blastocysts were stained with DAPI to determine blastomere numbers as described earlier.

In another study, pools of blastocysts derived from cultures containing or lacking PD173074 were collected and RNA was extracted as described previously (n=4 replicate studies; 10–15 blastocysts/treatment/replicate). Effects of PD173074 treatment on NANOG, GATA4, CDX2, and IFNT mRNA abundance were examined by qRT-PCR as described previously (Zhang et al. 2010, Yang et al. 2011a).

FGFR inhibition and outgrowth formation

Blastocyst outgrowth cultures were completed as described previously (Yang et al. 2011a). Blastocysts were collected at day 8 post-IVF and individual blastocysts were placed into Matrigel-coated wells (0.95 cm²) in 500 μl DMEM containing high glucose (4.5 g/l), 10% FBS, and either 1 μM PD173074 or vehicle only (0.005% DMSO) (n=6 replicate studies, 10–14 blastocysts/treatment/replicate). Cultures were incubated at 38.5 °C in a 20% oxygen environment. After 3 days’ exposure to treatment (i.e. day 11 post-IVF), medium was removed and replaced with DMEM containing 10% fetal bovine serum (FBS) but lacking treatments. Cultures were removed from the incubator daily from days 9 to 15 post-IVF to examine the onset of outgrowth formation as described previously (Yang et al. 2011a). At day 15 post-IVF, the surface area of a subset of the outgrowths was determined by tracing the outline of the outgrowths using a Nikon SMZ1500 stereomicroscope and Nikon DXM-1200F digital camera and quantifying surface area using NIS-Elements Software (Nikon, Melville, NY, USA).

Statistical analyses

Data were analyzed by ANOVA using the general linear model of the Statistical Analysis System (SAS Institute, Inc., Cary, NC, USA). Differences between individual means were compared using pairwise comparisons (Probability of Difference (PDf) Analysis in SAS). Relative abundances of selected transcripts were analyzed over time and across treatments after log-transformation. Percentage data were arcsine transformed. Data are presented as arithmetic mean ± S.E.M.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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References


Grigor’eva EV, Shevchenko AI, Mazurok NA, Elisaphenko EA, Zhelezova AI, Shilov AG, Dyban PA, Dyban AP, Noniashvili EM, Slobodyanyuk SY et al. 2009 FGF4 independent derivation of trophoblast stem cells from the common vole. PlOS ONE 4 e7161. (doi:10.1371/journal.pone.0007161)


Lamont FR, Tomlinson DC, Cooper PA, Snyder SD, Chester JD & Knowles MA 2011 Small molecule FGF receptor inhibitors block FGFR-dependent urothelial carcinoma growth in vitro and in vivo. British Journal of Cancer 104 75–82. (doi:10.1038/jjc.6606016)


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