Oxidative stress and ageing of the post-ovulatory oocyte

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Abstract

With extended periods of time following ovulation, the metaphase II stage oocyte experiences deterioration in quality referred to as post-ovulatory oocyte ageing. Post-ovulatory ageing occurs both in vivo and in vitro and has been associated with reduced fertilization rates, poor embryo quality, post-implantation errors and abnormalities in the offspring. Although the physiological consequences of post-ovulatory oocyte ageing have largely been established, the molecular mechanisms controlling this process are not well defined. This review analyses the relationships between biochemical changes exhibited by the ageing oocyte and the symptoms associated with the ageing phenotype. We also discuss molecular events that are potentially involved in orchestrating post-ovulatory ageing with a particular focus on the role of oxidative stress. We propose that oxidative stress may act as the initiator for a cascade of events that create the aged oocyte phenotype. Specifically, oxidative stress has the capacity to cause a decline in levels of critical cell cycle factors such as maturation-promoting factor, impair calcium homoeostasis, induce mitochondrial dysfunction and directly damage multiple intracellular components of the oocyte such as lipids, proteins and DNA. Finally, this review addresses current strategies for delaying post-ovulatory oocyte ageing with a particular focus on the potential use of compounds such as caffeine or selected antioxidants in the development of more refined media for the preservation of oocyte integrity during IVF procedures.

Introduction

Upon release from the ovary, the prophase I oocyte undergoes a process of maturation involving resumption of meiosis, breakdown of the germinal vesicle and extrusion of the first polar body (reviewed by Sun et al. (2009)). Following maturation, the oocyte once again enters meiotic arrest, now at metaphase II (MII). The MII stage oocyte awaits fertilization by the spermatozoon either in the oviduct of the female reproductive tract or, in an assisted reproduction setting, in in vitro culture media. An optimal window exists in which fertilization of this MII stage oocyte should occur; in mammalian species, this is generally within 10 h of ovulation; however, successful fertilization can still occur in the mouse for up to 15 h (Marston & Chang 1964, Tarin et al. 1999). In the event that no fertilization occurs within this time frame, oocytes experience a process of degradation referred to as ‘post-ovulatory ageing’. It should be noted that post-ovulatory ageing is a process distinct from that of ‘ovarian ageing’ (reviewed by Bentov & Casper (2013)), which occurs within the ovary of females towards the end of reproductive life, as they approach the climacteric. All references to oocyte ageing within this review refer to the deterioration of MII oocytes that occurs following their release from the ovarian follicle both in vivo and in vitro.


The cellular deterioration associated with post-ovulatory ageing can strongly influence the outcome of fertilization in vivo and in vitro. This is not surprising when considering that the oocyte provides factors to the embryo that orchestrate the early events of embryogenesis (Schultz & Heyner 1992, Minami et al. 2007), remodel the genome (Torres-Padilla et al. 2006) and repair DNA damage in both male and female nuclei.
Reproduction that fertilization could involve an synchrony between intercourse and release of the and, as a result, no mechanisms in place to ensure primates, however, there are no visual signs of ovulation synchronized with oocyte release. In humans and certain spermatozoa to the female reproductive tract is relatively result of such mechanisms is that the delivery of oestrus, or the act of mating itself triggers ovulation. The ovulation, as determined by the onset of behavioural post-ovulatory ageing is not uncommon. In the majority (Genesca et al. 1992). Unfortunately, the occurrence of mammalian post-ovulatory ageing is not uncommon. In the majority of mammalian species, mating occurs only at the time of ovulation, as determined by the onset of behavioural oestrus, or the act of mating itself triggers ovulation. The result of such mechanisms is that the delivery of spermatozoa to the female reproductive tract is relatively synchronized with oocyte release. In humans and certain primates, however, there are no visual signs of ovulation and, as a result, no mechanisms in place to ensure synchrony between intercourse and release of the oocyte. This, in turn, results in an increased likelihood that fertilization could involve an in vivo aged oocyte and freshly ejaculated spermatozoa. In an in vitro setting, oocytes employed in assisted reproduction technologies (ART) are often unavoidably subjected to extended periods of culture prior to fertilization. One circumstance in which this may occur is when ‘rescue ICSI’ is performed on failed-to-fertilize oocytes. In light of such considerations, it would clearly be beneficial to establish some control over the process of post-ovulatory oocyte ageing, particularly in an in vitro setting, where the demand for ART continues to increase exponentially (Wang et al. 2011).

This review explores the negative consequences associated with post-ovulatory oocyte ageing, discusses the poorly understood mechanisms that control this process and considers the current means by which post-ovulatory ageing can be prevented or delayed. Specifically, in light of recent research, we revisit the proposal that oxidative stress may act as the ‘trigger’ for a cascade of other events associated with oocyte ageing and hence might be an attractive approach for deterring this degenerative process in post-ovulatory oocytes.

### Table 1 Aberrations to oocyte biology associated with mammalian post-ovulatory ageing.

<table>
<thead>
<tr>
<th>Organelle</th>
<th>Consequences associated with ageing</th>
<th>In vivo/in vitro</th>
<th>Reference</th>
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<tbody>
<tr>
<td>Zona pellucida</td>
<td>Zona hardening</td>
<td>Both</td>
<td>Longo (1981), Dodson et al. (1989) and Xu et al. (1997)</td>
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<tr>
<td>Cortical granules</td>
<td>Partial exocytosis</td>
<td>Both</td>
<td>Szollosi (1971), Dodson et al. (1989), Ducibella et al. (1990), Xu et al. (1997) and Liu et al. (2009a, 2009b)</td>
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<tr>
<td>Spindle ER</td>
<td>Elongation, dispersal or disruption</td>
<td>In vitro</td>
<td>Takahashi et al. (2003)</td>
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<tr>
<td>Plasma membrane</td>
<td>Lipid peroxidation</td>
<td>Both</td>
<td>Takahashi et al. (2000)</td>
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<td>Mitochondria</td>
<td>Increased generation of ROS</td>
<td>Both</td>
<td>Zhang et al. (2011)</td>
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<td>DNA</td>
<td>Loss of membrane potential</td>
<td>In vitro</td>
<td>Zhang et al. (2011)</td>
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<td>Decreased ATP production</td>
<td>In vitro</td>
<td>Chi et al. (1988)</td>
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<td></td>
<td>Epigenetic changes</td>
<td>Both</td>
<td>Liang et al. (2008)</td>
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<td></td>
<td>Misaligned chromosomes</td>
<td>Both</td>
<td>Wakayama et al. (2004)</td>
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<td></td>
<td>Premature centromere separation</td>
<td>Both</td>
<td>Spielmann et al. (1985) and Mailhes et al. (1998)</td>
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IP₃, inositol 1,4,5-trisphosphate; ROS, reactive oxygen species.

### Clinical implications of oocyte ageing

The clinical implications of oocyte ageing include a decreased capacity for fertilization both in vivo (Marston & Chang 1964) and in vitro (Ben-Rafael et al. 1986, Badenas et al. 1989), production of poor-quality embryos (Yanagida et al. 1998, Lord et al. 2013), an increased likelihood of early pregnancy loss (Wilcox et al. 1998) and abnormalities in offspring (Tarin et al. 1999). The attributes of aged oocytes responsible for these adverse outcomes are discussed below.

### Decreased fertilization rate

A decrease in fertilization rate has been associated with aged oocytes in many species including the humans (Lash & Whittaker 1974), mice (Marston & Chang 1964, Liu et al. 2009b), cows (Chian et al. 1992) and pigs (Kikuchi et al. 1989 and Xu et al. 1989). This phenomenon can be attributed to multiple biochemical and functional alterations to the oocyte that accumulate with post-ovulatory age. First, aged murine oocytes are known to experience premature cortical granule exocytosis (Szollosi 1971, Dodson et al. 1989, Ducibella et al. 1990, Xu et al. 1997) and zona hardening (Longo 1981, Dodson et al. 1989, Xu et al. 1997); changes that would obviously impose limitations on the ability of the zona pellucida to interact with a spermatozoon and initiate the acrosome reaction. Additionally, as levels of lipid peroxidation are known to be elevated within post-ovulatory aged oocytes (Takahashi et al. 2003; Table 1), the fluidity of the oolemma may be decreased, reducing the likelihood of sperm–oolemma fusion and thereby fertilization.

In the event that sperm–zona interaction and sperm–oolemma fusion progress unimpared, the abilities of the sperm–oolemma fusion progression to be elevated within post-ovulatory aged oocytes.
post-ovulatory aged oocyte to exhibit normal activation and initiate embryo development following fertilization and initiate embryo development following fertilization are nonetheless compromised. Upon fertilization, inositol 1,4,5-trisphosphate (IP$_3$) receptors located on the oocyte's endoplasmic reticulum (ER) initiate calcium (Ca$^{2+}$) oscillations in response to IP$_3$ produced by the sperm-derived factor PLC$\_1$ (Yoon et al. 2008, Kuroda 2010). However, post-ovulatory aged oocytes experience a decline in IP$_3$ receptor/channel functionality (Zhang et al. 2011), as well as suppressed ER Ca$^{2+}$-ATPase activity, and exhibit a consequential depletion of Ca$^{2+}$ stores that reside within the ER (Takahashi et al. 2000). As a result of this impaired Ca$^{2+}$ homoeostasis, aged oocytes exhibit abnormal Ca$^{2+}$ oscillations at fertilization that have significantly higher frequency and lower amplitude than those of fertilized fresh oocytes (Igarashi et al. 1997, Takahashi et al. 2003). These abnormal Ca$^{2+}$ oscillations are purportedly associated with the onset of apoptosis following fertilization, rather than entry into a developmental pathway (Gordo et al. 2000, 2002).

**Poor embryo quality**

While the fertilization potential of post-ovulatory aged oocytes is obviously compromised, circumstances exist in which fertilization does still occur, particularly in clinical practice when ICSI is used to bypass the physiological mechanisms that would normally prevent the association of defective gametes. Not surprisingly, embryos originating from post-ovulatory aged oocytes are generally of poor quality and exhibit delayed or impaired development during subsequent embryogenesis (Yanagida et al. 1998, Lord et al. 2013).

The poor quality of these embryos is highlighted when we assess their capacity to implant in the endometrium and carry a pregnancy to term. With extensive periods of post-ovulatory ageing (>24 h in vitro), ICSI-generated embryos reportedly lose their capability for uterine implantation following embryo transfer (Chen et al. 1995). Despite this, oocytes that are subjected to shorter periods of post-ovulatory ageing can still produce embryos capable of implantation, but are prone to post-implantation errors. A clinical study assessing the effects of in vivo post-ovulatory oocyte ageing found a significant correlation between early pregnancy loss and the likelihood that delayed fertilization had occurred (Wilcox et al. 1998). These findings have also been reported in other mammalian species such as the mouse (Marston & Chang 1964).

The decline in embryo quality associated with post-ovulatory aged oocytes is presumably attributed to multiple factors. First, cytoplasmic ageing is likely to greatly hinder the capacity of the oocyte to support and orchestrate embryo development, particularly when changes in both the protein and mRNA composition of the oocyte are demonstrable with prolonged ovulation to fertilization latency (Tarin et al. 2000, Hamatani et al. 2004). Additionally, aged oocytes are likely to display a corresponding increase in the proportion of non-viable embryos following fertilization as a result of chromosomal abnormalities. Post-ovulatory aged oocytes purportedly exhibit significantly higher instances of chromosome misalignment, potentially as a consequence of meiotic spindles that are elongated, dispersed or disrupted (Wakayama et al. 2004). These characteristics increase the likelihood for erroneous chromosome separation and hence aneuploidy (Mailhes et al. 1998). Further to this, in certain species such as the bovine, advancing oocyte age is also associated with a significantly increased susceptibility to polyspermy (Chian et al. 1992), which again results in the generation of embryos that are essentially non-viable.

**Abnormalities in offspring**

Despite the obstacles associated with fertilization of post-ovulatory aged oocytes and subsequent embryo development and implantation, birth of live offspring originating from these gametes does still occur. It was originally thought that aged oocytes that manage to support embryo development to the blastocyst stage were no less capable of producing healthy offspring than their fresh oocyte counterparts (Wilcox et al. 1998). However, more recent research has demonstrated a significant decline in live birth rates and an elevated instance of abnormalities in offspring (Tarin et al. 1999).

Tarin et al. (1999, 2002) demonstrated that offspring originating from in vivo aged oocytes exhibited growth retardation, delayed development, heightened emotional state, decreased reproductive fitness and importantly decreased longevity. Although the specific factors within the post-ovulatory aged oocyte that so prominently affect offspring integrity have not been determined, it has been hypothesized that these abnormalities may stem from the transference of a subpopulation of dysfunctional mitochondria (Tarin et al. 2002). Impaired mitochondrial function has indeed been linked with diseases such as schizophrenia (Whatley et al. 1996), Alzheimer’s disease (Hutchin & Cortopassi 1995, Budd & Nicholls 1998) and a decreased lifespan (Sont & Vandenbroucke 1993).

An alternative explanation for the abnormalities observed in the offspring of in vivo and in vitro aged oocytes may lay with an altered epigenetic profile, which would essentially affect gene expression in the embryo. Both the male and female genomes experience significant demethylation events following fertilization (Mayer et al. 2000) and post-ovulatory oocyte ageing has been shown to significantly alter the methylation pattern of imprinted genes in both the oocyte and the developing placenta (Liang et al. 2008, 2011).
Factors affecting oocyte ageing and apoptosis

Post-ovulatory oocyte ageing is clearly a prominent issue affecting fecundity; however, the molecular mechanisms that control this process are not well elucidated. It is currently undetermined as to whether a single event acts to trigger a cascade of other factors, or if several biochemical and functional changes occur separately to create an ‘aged’, degenerating MII oocyte. Below we assess factors that are known to influence the onset of post-ovulatory ageing and specifically highlight the potential role of oxidative stress as the ‘trigger’ for ageing and apoptosis of the oocyte both in vitro and in vivo.

Cumulus cells

In the presence of cumulus cells, MII stage oocytes purportedly ‘age’ more rapidly. In vivo aged oocytes and in vitro aged oocytes enclosed within the cumulus cell complex experience increased rates of spontaneous activation and fragmentation (Miao et al. 2005, Wu et al. 2011), accelerated decline of MPF/MAPK (Miao et al. 2005) and decreased levels of blastocyst formation (Wu et al. 2011) when compared with denuded oocytes aged in vitro. Qiao et al. (2007) reported that the addition of cumulus cells to culture medium containing denuded oocytes caused an acceleration of post-ovulatory ageing on par with that of oocytes enclosed in a cumulus–oocyte complex, while other research groups have demonstrated that blocking gap junctions within the cumulus–oocyte complex does not prevent accelerated ageing (Wu et al. 2011). Following these observations, it has been hypothesized that cumulus cells secrete soluble/paracrine factor(s) that promote post-ovulatory ageing, potentially an event that coincides with the entry of the cumulus cells into apoptosis (Wu et al. 2011). Additionally, the accelerated depletion of the oocyte metabolite pyruvate from culture medium by cumulus cells (Downs et al. 2002) appears to directly affect oocyte metabolism and hence post-ovulatory ageing, potentially by causing downstream inhibition of protein synthesis within the oocyte and upsetting redox equilibrium (Liu et al. 2009b, Li et al. 2011).

An alternative explanation for cumulus cell-associated acceleration of post-ovulatory ageing has been detailed in publications by Perez et al. (2005) and Kujjo & Perez (2012). These researchers propose that the bioactive sphingolipid, ceramide, generated by the cumulus cells may be responsible for mitochondrial dysfunction and subsequent ageing and apoptosis in post-ovulatory oocytes. This is attributed to an increased sensitivity of aged oocytes to cytosolic ceramide spikes and the ability of ceramide to act as a pro-apoptotic factor upstream of Bax.

Although the presence of cumulus cells clearly exacerbates degeneration in ageing post-ovulatory oocytes, additional mechanisms must be at play in controlling this process, as oocytes that have been denuded of their cumulus cells also experience ageing and apoptosis (Miao et al. 2005, Lord et al. 2013).

MPF and MAPK

The critical cell cycle factors MPF and MAPK control meiotic resumption in germinal vesicle stage oocytes and then act to maintain cell cycle arrest once oocytes have reached the MII phase. A gradual decline in the concentration of these factors within the MII oocyte has been detected with post-ovulatory age and has been associated with increased levels of parthenogenetic activation and fragmentation (Kikuchi et al. 2000). The mechanism by which MPF specifically becomes degraded with post-ovulatory age has been studied extensively – particularly as a potential target for reversing the ageing process (to be discussed later in this review). MPF comprises two molecules: the catalytic subunit p34^{cdc2} and the regulatory subunit cyclin B. In the active form of MPF, these two subunits are coupled and T-161 of the catalytic subunit is phosphorylated. Following fertilization, MPF is inactivated by the dephosphorylation of T-161 and degradation of the decoupled cyclin B. Interestingly, inactivation of MPF occurs via an alternate pathway in the ageing post-ovulatory oocyte. Under these circumstances, the inactive compound ‘pre-MPF’ is formed, in which the two subunits are still bound but phosphorylation of T-14 and/or T-15 of p34^{cdc2} occurs (Kikuchi et al. 2002). The accumulation of pre-MPF is clearly a factor controlling at least some aspects of post-ovulatory ageing as acceleration of pre-MPF formation using the tyrosine phosphatase inhibitor, vanadate, has been shown to increase susceptibility to parthenogenetic activation and fragmentation, while maintenance of active MPF levels within the oocyte using caffeine decreases levels of parthenogenesis and fragmentation (Kikuchi et al. 2002), and extends the window for fertilization (Ono et al. 2011).

Mitochondrial dysfunction

Mitochondrial dysfunction appears to be another potent mediator of the post-ovulatory oocyte ageing process. Operational mitochondria are crucial to normal oocyte function, with these organelles representing the primary source of ATP production within both oocytes and early embryos (Dumollard et al. 2007). The functionality of the mitochondria does, however, become compromised with extended periods of time following ovulation, a factor that is thought to heavily influence oocyte quality. Diminished mitochondrial integrity in the in vitro aged oocyte has been demonstrated by a loss of mitochondrial membrane potential (Zhang et al. 2011) and a decline in levels of ATP production (Chi et al. 1988).

As damage to the mitochondria is known to cause an increased production of reactive oxygen species (ROS)
and the release of pro-apoptotic factors such as cytochrome c (Liu et al. 2009a), which recruit members of apoptotic machinery such as caspases (Takai et al. 2007) and endonucleases (Fujino et al. 1996), it is likely that mitochondrial dysfunction is the link between post-ovulatory oocyte ageing and apoptosis. Additionally, loss of mitochondrial integrity can result in aberrant protein synthesis and inactivation or loss of mitochondrial DNA (mtDNA; reviewed by Shigenaga et al. (1994)).

The pronounced control that mitochondrial function has over post-ovulatory ageing has been demonstrated in studies utilizing microinjection of isolated mitochondria into in vitro aged oocytes. During a 24-h culture period, MII oocytes whose mitochondrial pool was increased by 5% via microinjection showed a significant decline in levels of programmed cell death when compared with control oocytes (Perez et al. 2000).

Oxidative stress: the initiator of post-ovulatory ageing?

It has been demonstrated by several research groups that ROS – particularly hydrogen peroxide (H$_2$O$_2$), superoxide anion (O$_{2}$•−) and peroxynitrite (ONOO$^{-}$) – accumulate in oocytes with increasing amounts of time following ovulation both in vitro and in vivo (Takahashi et al. 2003, Tatone et al. 2011, Lord et al. 2013). ROS production within the MII oocyte likely occurs as a by-product of oxidative phosphorylation; however, environmental factors that oocytes are subjected to in vitro (exposure to light, lack of antioxidant rich follicular and tubal fluids and increased oxygen tension) are also thought to facilitate ROS production (Mastroianni & Jones 1965, Mass et al. 1976, Goto et al. 1993, Guerin et al. 2001). While the oocyte does offer some intracellular defence against oxidative attack in the form of the antioxidant glutathione, these resources are depleted with post-ovulatory age (Boerjan & de Boer 1990, Yoshida et al. 1993). As a consequence of the progressive increase in ROS production and the concomitant depletion of antioxidant protection, the post-ovulatory aged oocyte experiences a state of oxidative stress. We hypothesize that this oxidative stress may act as the ‘trigger’ for a cascade of other factors that orchestrate post-ovulatory ageing, as well as directly affect multiple aspects of oocyte biochemistry and functionality. The idea that oxidative stress may be the initiator of ageing in the MII oocyte is supported by recent findings indicating that the onset of oxidative stress is a relatively early event in in vitro culture (Lord et al. 2013).

Indeed, links can be established between oxidative stress in the oocyte and the aforementioned decline in critical cell cycle factors, mitochondrial dysfunction, apoptosis, impaired Ca$^{2+}$ homoeostasis, decreased fertilization rate, poor embryo quality and abnormalities in offspring. These associations will be discussed below.

We propose that the decline in the critical cell cycle factor, MPF, which reportedly orchestrates some facets of post-ovulatory ageing, may be a symptom of oxidative stress in the MII oocyte. As mentioned previously, the accumulation of ‘pre-MPF’ occurs with increasing periods following ovulation, with phosphorylation at multiple sites on the catalytic subunit of MPF effectively inactivating this compound (Kikuchi et al. 2000, 2002). The enzymes that purportedly control these phosphorylation events are cdc25 (a tyrosine phosphatase) and Wee1/Myt1 (tyrosine kinases) (Kikuchi et al. 2002). Interestingly, ROS have previously been shown to have a capacity for both inhibition of tyrosine phosphatases (Monteiro et al. 1991, Sullivan et al. 1994), including cdc25 specifically (Brisson et al. 2007), and stimulation of tyrosine kinases (Chan et al. 1986). As a result, the induction of oxidative stress in the post-ovulatory oocyte may directly affect levels of MPF within the cell, resulting in the onset of associated downstream symptoms of oocyte ageing such as parthenogenesis and fragmentation (Kikuchi et al. 2002).

Oxidative stress that occurs with post-ovulatory age also has the potential to directly affect mitochondrial function. The DNA, proteins and lipids within the mitochondria are particularly susceptible to oxidative attack, not only because of their close proximity to the source of ROS production (the electron transport chain (ETC)) but also in the case of mtDNA, because of the absence of protective histones and mechanisms for DNA repair (reviewed by Shigenaga et al. (1994)). Oxidative stress has been linked with mtDNA damage and deletions (Sohal & Dubey 1994), loss of mitochondrial membrane potential (Liu et al. 2000), increased ROS generation by the ETC (Liu et al. 2009a) and a decline in ATP production (Melov et al. 1999). Importantly, factors such as increased ROS generation and a decline of ATP production are also known to be associated with post-ovulatory ageing (Chi et al. 1988, Lord et al. 2013), suggesting that a link between oxidative stress and age-related mitochondrial dysfunction is certainly plausible. Notably, oxidative damage to mtDNA in the oocyte could potentially be the basis for the aforementioned abnormalities associated with impaired mitochondrial function in offspring originating from aged oocytes (Tarin et al. 2002).

Oxidative stress also has the capacity to directly influence the onset of apoptosis in post-ovulatory aged oocytes. H$_2$O$_2$ treatment of MII oocytes in vitro has been shown to cause a decline in levels of the anti-apoptotic molecule Bcl-2 (Takahashi et al. 2009), induce expression of pro-apoptotic Bax and caspase-3, and precipitate both DNA fragmentation (Chaube et al. 2007) and cytochrome c release (Liu et al. 2000). Importantly, these pathologies are common to both oxidative stress and oocyte ageing, and relief from oxidative stress by way of antioxidant treatment has been shown to delay the onset of fragmentation and caspase activation associated with post-ovulatory ageing (Lord et al. 2013).
In addition to potentially being a precursor to MPF/ MAPK depletion, mitochondrial dysfunction and subsequent apoptosis, the damaging nature of oxidative stress itself is likely to be directly involved in generation of the ‘aged oocyte’ phenotype. First, oxidative stress may be accountable for a perturbation of Ca\(^{2+}\) homoeostasis in aged oocytes. Previous research has demonstrated that these two events are implicitly linked, as treatment of fresh oocytes with H\(_2\)O\(_2\) results in demonstrated that these two events are implicitly linked, as treatment of fresh oocytes with H\(_2\)O\(_2\) results in abnormal Ca\(^{2+}\) oscillations following fertilization, which exhibit similar frequency and amplitude to those which occur within the oocyte following timely fertilization (Liu et al. 2009b, Li et al. 2012). As the oocyte and early embryo preferentially utilize pyruvate over glucose to drive metabolism (Leese \\& Barton 1984), it has been proposed that pyruvate supplementation delays oocyte ageing by fuelling prolonged ATP production within oocytes and also by maintaining intracellular redox potential (Liu et al. 2009b, Li et al. 2011). Increased pyruvate concentration is particularly effective in preventing accelerated in vitro ageing of the cumulus-enclosed MII oocyte, a result likely attributed to the more rapid depletion of pyruvate from culture media in the presence of cumulus cells (Downs et al. 2002).

Alteration of culture temperature has also been shown to delay the onset of post-ovulatory ageing in vitro. Porcine oocytes supplemented with pyruvate and stored at 15 °C were able to be successfully fertilized and maintain developmental potential after 42 h in vitro, whereas oocytes exposed to the same conditions at 37 °C could only be successfully fertilized after 6 h of culture (Li et al. 2012). The mechanism behind delayed ageing at low temperatures is likely related to a down-regulation of oocyte metabolism (Chip et al. 2011), resulting in reduced levels of ROS production and, subsequently, a decreased likelihood for oxidative damage and ROS-induced apoptosis (Li et al. 2012).

**Mechanisms for preventing/delaying oocyte ageing**

The final portion of this review looks into current strategies for delaying or preventing post-ovulatory oocyte ageing and assesses their effectiveness and potential for adaptation to clinical use in the future.

**Culture conditions and media composition**

Alterations to the concentration of metabolites within oocyte culture media has been shown to prominently affect post-ovulatory ageing in vitro. Specifically, increasing the concentration of pyruvate within culture media has been shown to delay post-ovulatory ageing by maintaining cortical granule integrity, decreasing susceptibility to activating stimuli, preventing an age-associated decline in levels of MPF and glutathione, improving blastocyst formation rates following fertilization and delaying the onset of apoptosis (Liu et al. 2009b, Li et al. 2012). As the oocyte and early embryo preferentially utilize pyruvate over glucose to drive metabolism (Leese \\& Barton 1984), it has been proposed that pyruvate supplementation delays oocyte ageing by fuelling prolonged ATP production within oocytes and also by maintaining intracellular redox potential (Liu et al. 2009b, Li et al. 2011). Increased pyruvate concentration is particularly effective in preventing accelerated in vitro ageing of the cumulus-enclosed MII oocyte, a result likely attributed to the more rapid depletion of pyruvate from culture media in the presence of cumulus cells (Downs et al. 2002).

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**Maintenance of MPF levels**

Currently, one of the most thoroughly investigated methods for preventing/delaying post-ovulatory oocyte
ageing in vitro is by maintaining levels of critical cell cycle factors within the cell. Caffeine is one such compound that has been utilized in this regard, as it acts to inhibit phosphorylation of MPF to pre-MPF by Wee1/Myt1. Supplementation of oocyte culture media with caffeine has indeed been shown to maintain levels of MPF post-ovulation and to subsequently decrease levels of parthenogenesis and fragmentation (Kikuchi et al. 2002). Additionally, recent studies have established that the addition of caffeine or MG132 (a proteasome inhibitor) to oocyte culture can increase fertilization rates by ICSI and decrease fragmentation in embryos (Ono et al. 2011). Further modes of action for caffeine in relation to post-ovulatory ageing include a delay in the deterioration of Ca\(^{2+}\) release mechanisms, although the underlying pharmacology is not yet clear (Zhang et al. 2011).

While live offspring have been obtained using oocytes that were exposed to caffeine treatment (Ono et al. 2011), previously published research has implicated caffeine as an inhibitor of DNA repair mechanisms (Selby & Sancar 1990). This impairment of DNA repair has been demonstrated to directly affect oocytes, as caffeine-treated golden hamster eggs lost their capacity to repair aberrations in both chromatids and chromosomes following fertilization by spermatozoa possessing DNA damage (Genesca et al. 1992). This clearly creates concern as to the safety of utilizing caffeine as an agent for preventing post-ovulatory oocyte ageing in vitro, as un-repaired DNA damage within the embryo may result in abnormalities/birth defects in offspring.

In a similar fashion to caffeine, MG132 has also been demonstrated to maintain high MPF levels in oocytes in vitro, by preventing degradation of cyclin B. Oocytes treated with MG132 have been shown to produce embryos with improved developmental potential; however, the capacity to produce live offspring is reduced (Yu et al. 2005), potentially as a result of inhibition of necessary protein degradation (Gao et al. 2005). Again,
MG132 is clearly not a suitable compound for utilization in an ART setting.

In addition to the aforementioned impacts of caffeine or MG132 on cultured oocytes, recent research has suggested that not all facets of post-ovulatory ageing can be controlled by manipulation of cell cycle factors. It has been demonstrated that caffeine-supplemented oocytes do not experience any delay in the onset of age-associated apoptosis, nor any relief from accumulation of oxidative stress (Lord et al. 2013). If accumulation of oxidative stress is indeed an upstream event that is intricately linked with age-associated declines in critical cell cycle factors, then relief from oxidative stress by way of antioxidant supplementation should maintain MPF/MAPK levels in vitro.

Antioxidants

If oxidative stress acts as a ‘trigger’ for ageing in post-ovulatory oocytes, then it would be expected that antioxidant treatment would effectively delay this process, either via supplementation of oocyte culture medium to delay ageing in vitro or by oral administration to prevent ageing of oocytes in vivo.

In vitro studies have indeed demonstrated that antioxidant supplementation can attenuate the process of post-ovulatory oocyte ageing; however, this phenomenon is reliant on the type of antioxidant utilized. Early research by Tarin et al. (1998) demonstrated that L-ascorbic acid (vitamin C) and 6-methoxy-2,5,7,8-tetramethylchloromane-2-carboxylic acid (trolox/vitamin E) could not significantly improve fertilization rate nor prevent fragmentation associated with post-ovulatory ageing, while β-mercaptoethanol and L-cysteine in fact decreased the likelihood of aged oocytes reaching the blastocyst stage following fertilization. However, this research also established that the addition of the reducing agent dithiothreitol (DTT) to oocyte culture medium resulted in increased fertilization and blastocyst formation rates, potentially by protecting free thiol groups within these ‘aging’ oocytes from becoming oxidized. Despite the reported benefits associated with DTT, the application of this method for delaying post-ovulatory ageing in a clinical setting is limited as DTT has been associated with DNA damage within cells (Held et al. 1996, Oikawa et al. 2002).

Treatment of oocytes with the nitric oxide (NO•) donor S-nitroso-N-acetylpenicillamine has also been shown to attenuate certain signs associated with post-ovulatory ageing such as cortical granule exocytosis, zona hardening and poor embryo quality (Goud et al. 2008b). NO• has been identified in multiple studies as a free radical crucial to oocyte and embryo function (Gouge et al. 1998, Kuo et al. 2000). NO• does exhibit antioxidant characteristics – specifically a capacity to scavenge O2•− (Robak & Grygleweski 1993); however, the exact mechanism by which NO• elicits its age-attenuating effects remains elusive.

More recently, a study conducted by our laboratory (Lord et al. 2013) has identified that the potent antioxidant melatonin provides protection from post-ovulatory ageing when used to supplement in vitro culture medium. Oocytes treated with melatonin experienced a delayed onset of apoptosis, decreased levels of fragmentation, an increased optimal window for fertilization and improved embryo quality when compared with control aged oocyte counterparts. These effects are presumably a direct result of relief from oxidative stress, as melatonin-treated aged oocytes exhibited significantly reduced levels of ROS (Lord et al. 2013). Further to this, previous studies have highlighted the ability of melatonin to reverse the deleterious effects of H2O2 treatment on MI oocytes (Tamura et al. 2008) and prevent hypochlorous acid-induced abnormalities in chromosomes and microtubules (Banerjee et al. 2012). Importantly, melatonin has a lack of demonstrable toxicity (Jahnke et al. 1999), making it a primary candidate for utilization in an assisted reproduction setting.

Although the ability of antioxidants to influence post-ovulatory ageing is well characterized in vitro, the capacity for these compounds to delay in vivo ageing is poorly defined. Oral administration of antioxidants such as co-enzyme Q10, melatonin and N-acetyl-L-cysteine has been linked with improved oocyte quality, as well as increased fertilization rates and litter sizes in clinical and laboratory studies (Tamura et al. 2008, Burstein et al. 2009, Liu et al. 2012); however, these reports focus primarily on the relationship between antioxidants and ovarian ageing. To the authors’ knowledge, no in vivo studies have been conducted to establish the effects of orally administered antioxidants on post-ovulatory ageing specifically.

Although the beneficial effects of antioxidants on post-ovulatory oocyte ageing are dependent on the antioxidant utilized, these compounds may be at the forefront in terms of delaying in vitro ageing prior to IVF or ICSI, particularly if oxidative stress does act as the ‘trigger’ for downstream pathological changes. The variation in therapeutic potential between antioxidants trialled to date may be attributed to the different reactive species scavenged by each compound, their stability in in vitro culture media, their capacity to infiltrate multiple organelles within the cell and also their potency at concentrations that are considered safe for clinical use. Following the success of antioxidant supplementation in vitro, it would be beneficial to establish the capability of these compounds to delay the onset of senescence in MII oocytes in vivo.

Concluding remarks

Ageing of post-ovulatory oocytes clearly has a major impact on fertilization potential, the quality of resulting
Declarations of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the review.

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References


Dodson MG, Minhas BS, Curtiss SK, Palmer TV & Robertson JL 1989 Spontaneous zona reaction in the mouse as a limiting factor for the time in which an oocyte may be fertilized. Journal of In Vitro Fertilization and Embryo Transfer 6 101–106. (doi:10.1007/BF01130735)


Hutchin T & Cortopassi G 1995 A mitochondrial DNA clone is associated with increased risk for Alzheimer disease. PNAS 92 6892–6897. (doi:10.1073/pnas.92.15.6892)


Oxidative stress and the ageing oocyte


Yoon SY, Jellerette T, Salicioni AM, Lee HC, Yoo MS, Coward K, Parrington J, Grow D, Cibelli JB, Visconti PE et al. 2008 Human sperm-seeded blastocysts containing zeta fail to induce Ca2++ release and are unable to initiate the first step of embryo development. Journal of Clinical Investigation 118 3671–3681. (doi:10.1172/JCI36942)

