Progesterone effects on mouse sperm kinetics in conditions of viscosity

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Abstract

The spermatozoa delivered to the female genital tract need to swim towards the oocyte through viscous secretions. Once close to the oocyte, the spermatozoa are guided by a gradient of progesterone (P₄) and other unknown chemoattractants via a process known as chemotaxis. Using polyvinylpyrrolidone to establish the conditions of viscosity, we examined the response of mouse spermatozoa to P₄. Herein, we show that in low-viscous media, P₄ induces hyperactive-like motility whereby sperm show erratic trajectories and non-progressive movement. However, an opposite response is produced in viscous medium that trajectories are linear and motility is more progressive and less erratic. Our observations provide a behaviour explanation for the chemotaxis of spermatozoa swimming under viscous conditions in a spatial gradient of the chemoattractant P₄. They also highlight the importance of using viscous solutions to mimic in vivo conditions when analysing sperm behaviour in response to any stimulus.

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Introduction

Once in the female genital tract, to reach the fertilization site, spermatozoa need to tackle several anatomical structures, each with specific and complex physicochemical properties (Suarez & Pacey 2006). Active mechanisms will guide the spermatozoa to the proximity of the oocyte (Eisenbach & Giojalas 2006). So far, three such guidance mechanisms have been proposed: temperature gradient (thermotaxis), against fluid flow (rheotaxis) and chemoattractant(s) gradient (chemotaxis). All the three mechanisms are functional in the oviduct (see Pérez-Cerezales et al. (2015) for a review).

A feature shared by the three guidance mechanisms is that spermatozoa show a hyperactivation-like motility (Armon & Eisenbach 2011, Miki & Clapham 2013, Boryshpolets et al. 2015). Hyperactivation is a property of capacitated spermatozoa that, under low-viscous conditions, involves vigorous movements produced by asymmetrical and high-amplitude waves in the flagella, resulting in erratic swimming trajectories (Suarez 2008). Currently, the model suggested for spermatozoa swimming in a spatial gradient of temperature (Boryshpolets et al. 2015) or chemoattractant (Gakamsky et al. 2009, Armon & Eisenbach 2011) is similar to that described for bacterial chemotaxis (Macnab & Koshland 1972). According to this model, a spermatozoon swimming in a gradient towards a lower temperature or lower chemoattractant concentration responds by increasing the frequency of hyperactivation-like events and turns to modify its direction of swimming. Conversely, when swimming towards a higher temperature or chemoattractant concentration, this constant stimulation causes the suppression of turns and hyperactive-like events. If no changes are perceived, the spermatozoon recovers its non-stimulated swimming mode, consisting of a straighter movement pattern with occasional hyperactivation events and turns. Although the model has been empirically validated for chemotaxis and thermotaxis, the behavioural response of spermatozoa to progesterone (P₄) as the main known chemoattractant has shown some discrepancies with the model. In studies addressing the response of a population of human spermatozoa to P₄ (1–10 nM), enhanced hyperactivation-like motility events have been observed (Uhler et al. 1992, Gakamsky et al. 2009). This argues against the proposed model as increasing concentrations of chemoattractant should have a damping effect on hyperactivation-like motility. In an elegant series of experiments, Armon et al. (2011) showed that hyperactive-like motility of human spermatozoa was effectively reduced in the sperm subpopulation swimming towards higher concentrations of P₄. However, the physiological meaning of P₄’s main effect of increasing the hyperactive-like motility of the general sperm population remains unknown.

It has been established that CatSper mediates P₄-induced Ca²⁺ influx (Lishko et al. 2011, Strünker et al. 2011) as well as hyperactivation (Carlson et al. 2003) in human spermatozoa. However, studies conducted so far have failed to detect CatSper activation by P₄ in mouse spermatozoa (Lishko et al. 2011), neither P₄-mediated chemotaxis (Chang et al. 2013). It has been shown in
mice that the initial Ca\(^{2+}\) influx provoked by P\(_4\) occurs at the base of the sperm head (Fukami et al. 2003), whereas CatSper activation initiates Ca\(^{2+}\) influx in the main stretch of the flagellum (Xia et al. 2007). Moreover, P\(_4\) elicits transient Ca\(^{2+}\) influx in spermatozoa from CatSper1-null mice (Ren et al. 2001), suggesting that P\(_4\) might activate another Ca\(^{2+}\) influx pathway besides CatSper in mammalian sperm. It has been reported that P\(_4\) fails to induce Ca\(^{2+}\) fluctuations in Plocδ-null mice (Fukami et al. 2003), suggesting PLC pathway involvement in this response, possibly via Ca\(^{2+}\) mobilization from internal Ca\(^{2+}\) stores at the base of the sperm head through the IP3 receptor (Darszon et al. 2011). Thus, given the localization of the P\(_4\) membrane receptor in mouse spermatozoa (Pietrobon et al. 2003), its activation by P\(_4\) might be the mechanism mediating the hyperactivation response, perhaps through the PLC pathway.

Interestingly, it has been noted that the behaviour of human spermatozoa in response to temperature changes varies under conditions of viscosity or low viscosity. Under viscous conditions considered closer to those of the physiological environment, the temperature-dependent hyperactive-like behaviour observed in low-viscous conditions changes to one showing a higher frequency of turns when the temperature is reduced (Boryshpolets et al. 2015). This indicates the different behaviour of spermatozoa in response to a stimulus depending on the viscosity of the surrounding medium. Indeed, hyperactivation-like motility in low-viscous conditions has been proposed to be the manifestation of a more efficient motility mode for sperm propagation under viscous conditions in hamster (Suarez et al. 1991) and mouse spermatozoa (Suarez & Dai 1992). Thus, the hyperactive-like motility shown by spermatozoa swimming in low-viscous conditions in response to P\(_4\) could also indicate a more efficient swimming mode under physiological conditions of viscosity.

This study was designed to address the inconsistencies observed in the P\(_4\) chemotaxis model by determining the behaviour of mouse spermatozoa in response to this chemoattractant under conditions simulating the viscosity of the physiological environment of the female genital tract.

Materials and methods
Reagents
All media components were purchased from Sigma–Aldrich, except where otherwise stated.

Spermatozoa
Experiments with mice were carried out in strict accordance with recommendations of the guidelines of European Community Council Directive 86/609/EEC. Every effort was made to minimize suffering. Experiments were approved by the Committee on the Ethics of Animal Experiments of the INIA (permit number CEEA 2014/025).

For sperm analysis, F1 generation males from crosses between C57BL/6j (B6) female × DBA/2j (D2) male mice were used. The animals were kept in conditions of a 14 h light:10 h darkness cycle with no food or water restrictions. At the age of 4 months, males were individually isolated for 2 weeks before killing by cervical dislocation. Spermatozoa were collected from the caudal epididymis and suspended in human tubal fluid (HTF) medium (2.04 mM CaCl\(_2\) × 2H\(_2\)O, 101.6 mM NaCl, 4.69 mM KCl, 0.37 mM KH\(_2\)PO\(_4\), 0.2 mM MgSO\(_4\) × 7H\(_2\)O, 21.4 mM sodium lactate, 0.33 mM sodium pyruvate, 2.78 mM glucose, 25 mM NaHCO\(_3\), 100 U/mL penicillin, 50 μg/mL streptomycin SO\(_4\) and 0.001% (w/v) phenol red, supplemented with 1% BSA and stabilized overnight at 37°C in a 5% CO\(_2\) atmosphere. Once collected, the sperm samples were incubated for 1 h under an atmosphere of 5% CO\(_2\) at 37°C for capacitation.

Sperm processing and motility
After capacitation, spermatozoa were diluted to a final concentration of (~2 × 10\(^6\)/mL in HTF with or without polyvinylpyrrolidone (PVP) of average MW360000 at 4, 6 or 7.5% (w/v). For each experiment with P\(_4\), 20 μL of P\(_4\) or DMSO (control) pre-diluted in HTF (1:10) were added to 100 μL of spermatozoa diluted in HTF with or without PVP. Motility was then determined within 20 s of adding P\(_4\) or DMSO by placing 20 μL of sperm suspension onto a pre-warmed slide on the stage heated to 37°C of a Nikon Eclipse 50i microscope (Nikon) fitted with a digital camera Basler A312f (Basler AG, Ahrensburg, Germany) capable of recording 25 frames/s. In each repetition, five videos of 1 s (20–60 moving spermatozoa) were recorded in different fields on the top of the drop and analysed using the Integrated Semen Analysis System (ISAS; Projectes i Serveis R+D S.L., Valencia, Spain). In each experiment, three replicates for each condition/treatment were set up. The parameters analysed were as described by Mortimer et al. (2000): straight-line velocity (VSL); time-averaged velocity of the sperm head along a straight line from its first position to its last position, expressed in μm/s; curvilinear velocity (VCL); time-averaged velocity of the sperm head along its actual curvilinear path, expressed in μm/s; average path velocity (VAP); velocity over an average path generated by a roaming average between frames, expressed in μm/s; linearity (LIN) (defined as (VSL/VCL) × 100); straightness (STR) (defined as (VSL/ VAP) × 100); wobble (WOB) (defined as (VAP/VCL) × 100); amplitude of lateral head (ALH) displacement (width of the lateral movement of the sperm head, expressed in μm) and beat-cross frequency (BCF; number of times the sperm head crosses the direction of movement per second, expressed in Hz).

Acrosome reaction
The method used was based on acrosome staining using Pisum sativum lectin conjugated with FITC (PSA–FITC) following a standard protocol as described previously (Lybaert et al. 2009) with minor modifications. Briefly, P\(_4\) at 0.015 pM,
15 pM, 15 nM, 15 μM or the same volume of DMSO (as a negative control) was added to capacitated spermatozoa and these were immediately processed for acrosome staining. In parallel, spermatozoa were diluted in HTF with or without 6% PVP (w/v) and also immediately processed for acrosome staining. As a positive control, spermatozoa were incubated for 30 min with 10 μM of the ionophore A23187 to induce the acrosome reaction. The spermatozoa were washed twice in PBS by centrifugation (1 min at 500 g) and subsequently smeared on a microscope glass slide and air-dried for 15 min. Next, the slides were immersed in absolute methanol for 30 s, air-dried and rinsed in PBS twice for 5 min before incubation with PSA–FITC (15 μg/mL in PBS) in a humid box for 30 min. Finally, the slides were washed with distilled water for 15 min and mounted with an aqueous mounting medium (Fluoromount G; EMS, Hatfield, PA, USA). Slides were examined in a fluorescence microscope and numbers of acrosome-reacted and non-acrosome-reacted spermatozoa were counted by randomly moving across different fields of the slide (counting 100 cells per slide). For each treatment, experiments were run in duplicate using two sperm samples from two different animals.

**Statistical analysis**

All statistical tests were performed using the software package SPSS Statistics 22.0 for Windows (IBM, Armonk, NY, USA). Results are provided as means ± S.E.M. Means were compared and analysed by three-way ANOVA followed by Tukey’s post hoc test. Significance was set at *P* < 0.05.

**Results**

To establish the swimming pattern of mouse spermatozoa in viscous conditions, we examined sperm kinetics at four concentrations of PVP (0 (low-viscous medium), 4, 6 and 7.5% (viscous media)). Differences in most of the variables recorded were detected between the viscous media and low-viscous medium. Velocities (VCL, VSL and VAP) decreased with increasing percentages of PVP in the medium (*P* < 0.05, three-way ANOVA; Fig. 1A). The velocity ratios LIN and STR were only affected at the highest PVP concentration (7.5%; *P* < 0.05, three-way ANOVA; Fig. 1B), indicating equally proportional reductions in the three velocities when spermatozoa swam in a medium containing 4 and 6% PVP. The other velocity ratio analysed, WOB, did not vary with PVP concentration. In addition, viscosity effects were also observed on motility patterns such that in the viscous media, the lateral displacement of the cell was reduced in an inverse manner with respect to the PVP concentration, reflected as a reduction in BCF and ALH with increasing PVP percentage (*P* < 0.05, three-way ANOVA; Fig. 1C). Accordingly, PVP led to the reduced velocity of spermatozoa and reduced side-to-side movement of the head and frequency of this movement, leading to a slower, more regular swimming trajectory (Fig. 1D). In the absence of PVP, the swimming pattern was erratic, including more irregular side-to-side head movements.

We then went on to assess the dose-dependent effect of P₄ on spermatozoa in low-viscous vs viscous medium. For these experiments, 6% PVP was selected as the highest concentration for which relationships among the three velocities defined were unaffected, determining that STR, WOB and LIN remained unchanged. This means that 6% PVP does not affect the STR of the sperm swimming trajectory and that any change in sperm behaviour could be attributed to their response to P₄. Moreover, this percentage of PVP did not induce the acrosome reaction (Fig. 2), which could in itself affect the behaviour of the spermatozoa.

In low-viscous medium, P₄ reduced the LIN of sperm movement indicated by reduced values of LIN, STR and WOB (*P* < 0.05, three-way ANOVA; Fig. 3). These effects were already observed at the lowest P₄ concentration tested (*P* < 0.05, three-way ANOVA). Such changes produced in LIN, STR and WOB (Mortimer 2000) reflect the effect of P₄ on sperm, provoking a slight increase in VCL and slight decreases in VSL and VAP (these effects alone were insignificant; three-way ANOVA; Fig. 3). In addition, 15 nM and 15 μM concentrations of P₄ led to a greater ALH (*P* < 0.05, three-way ANOVA), whereas no effects were observed on BCF. These observations indicate that the vigorous motility induced by P₄ consisted of a greater sperm head...
Moreover, P₄ and progesterone (P₄; concentration 0), DMSO (solvent) was added instead of P₄ at the same volume. Varying concentrations of P₄ were used to induce the acrosome reaction (0.015 µM, 15 pM, 15 nM and 15 µM). *P<0.05 and **P<0.02 according to three-way ANOVA. Mean ± S.E.M., n=8 determinations; each determination is the average of measurements in 52–60 spermatozoa.

Figure 2 Acrosome-reacted spermatozoa visualized with *Pisum sativum* agglutinin conjugated to FITC (PSA–FITC) under different conditions. A23187 was used as a positive control to induce the acrosome reaction. 6% PVP was used for the viscosity conditions. As a negative control for progesterone (P₄; concentration 0), DMSO (solvent) was added instead of P₄ at the same volume. Varying concentrations of P₄ were used to induce the acrosome reaction (0.015 µM, 15 pM, 15 nM and 15 µM). *P<0.05 and **P<0.02 according to three-way ANOVA. Mean ± S.E.M., n=8 determinations; each determination is the average of measurements in 52–60 spermatozoa.


displacement distance but did not affect the frequency at which the head crossed the swimming trajectory. These changes in sperm kinetics are characteristic of a hyperactive-like motility, as described by others for human spermatozoa in response to P₄ (Uhler et al. 1992, Gakamsky et al. 2009). P₄ at 15 pM, 15 nM and 15 µM induced the acrosome reaction (Fig. 2; P<0.05, three-way ANOVA), meaning that the motility changes induced by P₄ were accompanied by the release of acrosome contents.

In the viscous medium (6% PVP), contrary effects of P₄ to those produced in the low-viscous medium were observed. The P₄ dose-dependent curves between low viscosity vs viscosity were significantly different for VSL, VAP, LIN, STR, WOB and ALH (P<0.01, three-way ANOVA; Fig. 3). Thus, sperm velocities (VSL and VAP) were increased (in response to 15 nM and 15 µM P₄; P<0.05, three-way ANOVA) and movement became linear as indicated by LIN and WOB ratios with increasing P₄ concentrations (significant for 15 nM and 15 µM at P<0.05, three-way ANOVA) (Fig. 3). Moreover, P₄ led to a higher BCF (significant for 15 µM; P<0.05, three-way ANOVA), whereas ALH was unaltered. These results suggest that the P₄-induced non-progressive sperm motility (hyperactivation-like) observed in low-viscous medium is equivalent to the progressive motility observed under the more physiological conditions of the viscous media.

Discussion

Once in the female genital tract, spermatozoa have to swim in different mucus secretions whose viscosity differs according to anatomical site (Jansen 1978, Smith et al. 2009) and oestrous cycle stage (Jansen 1978, Miki & Clapham 2013). Thus, the averaged viscosity of mouse uterus and oviduct is 81 and 1.8 cP, respectively (Miki & Clapham 2013), in the range of viscosity obtained with the percentages of PVP (Hyakutake et al. 2015) used in our study. The effects of viscosity on mouse sperm kinetics observed here including a reduced velocity but more regular trajectory are in agreement with observations in hamster (Suarez et al. 1991), mice (Suarez & Dai 1992) and cattle (Hyakutake et al. 2015). The different behaviour shown by spermatozoa in low-viscous compared with viscous media (Fig. 4A) suggests a need to study those factors determining the sperm motility mode under the latter conditions. P₄ is among the most studied modulators of sperm function (Baldi et al. 2009). This hormone is secreted by cumulus cells and is a physiological sperm chemotactic (Teves et al. 2006, Oren- Benaroya et al. 2008). In agreement with earlier data for human spermatozoa (Uhler et al. 1992, Gakamsky et al. 2009), under low-viscous conditions, we also observed hyperactivation-like motility in mouse spermatozoa in response to acute P₄ treatment. Thus, P₄ reduced LIN, STR and WOB and increased lateral displacement of the head (ALH; Figs 3 and 4A). As a consequence of these motility changes, the P₄-activated spermatozoa acquired a more erratic swimming pattern, moving from side to side with higher amplitude (Fig. 4A). This motility type is known to facilitate spermatozoon progression under viscous conditions (Suarez et al. 1991, Suarez & Dai 1992). Remarkably, under viscous conditions, we observed the opposite effects of P₄ on sperm kinetics (Figs 3 and 4A). Thus, sperm LIN and BCF increased, whereas head displacement was reduced (Figs 3 and 4A). These results indicate that, contrary to what occurs in low-viscous media, the motility type induced by P₄ is one of the propagation under viscous conditions. Therefore, it seems logical that the known non-progressive motility mode induced by P₄ is actually an artefact of analysing the response under low-viscous conditions. This means that spermatozoa behave as expected for the model suggested for sperm chemotaxis when swimming in a spatial gradient of the chemotactic P₄ (Arnon & Eisenbach 2011). Thereby, when a spermatozoon swims towards a higher concentration of P₄, its motility becomes linearized, propagating the cell towards the higher concentrations (Fig. 4B). In contrast, a spermatozoon swimming
towards lower concentrations of P₄ would show a more circular trajectory until higher concentrations of P₄ linearized its motility again.

Chang et al. (2007) did not succeeded in detecting spatial chemotaxis in response to gradient of P₄ in mouse spermatozoa when using conditions of low viscosity. Our results provide an explanation to this observation, as only under viscous conditions, the spermatozoa would migrate towards a P₄ gradient in a chemotaxis spatial assay. Accordingly, the chemotaxis of human spermatozoa to P₄ was reported using a chemotaxis spatial assay under viscous conditions (3.5% PVP 25 K; Armon & Eisenbach 2011). This demonstrates the importance of using conditions closer to physiological when studying the behavioural response of spermatozoa to any kind of stimuli. Complementary studies would include analyses of the response to P₄ under other rheological conditions of the surrounding medium such as the viscoelasticity that characterize some fluids present along the female genital tract (Suarez & Dai 1992). It has been reported that at similar viscosity, the VSL and flagellar beat frequency of cattle spermatozoa are higher in non-Newtonian fluids leading to a more linear motility type (Hyakutake et al. 2015) compared with Newtonian fluids. Accordingly, it has been shown that under viscoelastic conditions, hyperactive samples of mouse spermatozoa linearize the motility whereas non-hyperactive spermatozoa behave in an opposite way (Suarez & Dai 1992).

Our results also show that at a certain concentration (higher than 15 pM), P₄ induces the acrosome reaction (Fig. 2). This observation is consistent with the dual effect of P₄ on human spermatozoa, whereby it acts as a chemoattractant at lower concentrations and induces the acrosome reaction at higher concentrations (Uhler et al. 1992). This release of acrosome contents could be related to the P₄ gradient existing within the cumulus cells (Teves et al. 2006). Thus, we propose that P₄ could linearize motility when sperm swimming up the P₄ gradient penetrate the cumulus and this could simultaneously provoke the release of acrosome contents to facilitate this penetration (Fig. 4B). This dual effect of P₄ could only apply to certain sperm populations, as recently described in human spermatozoa (Uñates et al. 2014). Accordingly, the acrosome-reacting population would eliminate the barrier formed by the cumulus cells and the chemotactic population would progress towards the oocyte by acquiring a more linear path in response to the P₄ gradient. It is also possible that both acrosome-reacting and chemotactic populations are comprised in part by the same spermatozoa. Indeed, it seems that most fertilizing spermatozoa undergo the acrosome reaction before reaching the zona pellucida of cumulus-enclosed oocytes (jin et al. 2011). This would be an interesting case of altruism between ‘sister’ cells that needs to be further explored to extend the concept of cooperation between spermatozoa of a single ejaculate. Despite no descriptions to date in the laboratory mouse, heterogeneity among different sperm populations of the same ejaculate and cooperation phenomena have been observed in the wood mouse Apodemussylvaticus (Moore et al. 2002) and species of Peromyscus (Fisher & Hoekstra 2010).

In conclusion, our findings suggest that the hyperactive-like motility induced by P₄ in mouse spermatozoa swimming in low-viscous media is a manifestation of the chemotactic behaviour of spermatozoa that propagates them towards a chemoattractant gradient in viscous conditions. In view of the dramatically different responses to P₄ produced in spermatozoa swimming in...
low-viscous or viscous media, we propose the viscosity of the medium is an essential factor to consider in any study designed to address sperm behaviour.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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