BVDV alters uterine prostaglandin production during pregnancy recognition in cows

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Abstract

Embryonic mortality in cows is at least in part caused by failure of pregnancy recognition (PR). Evidence has shown that bovine viral diarrhoea virus (BVDV) infection can disrupt pregnancy. Prostaglandins (PG) play important roles in many reproductive processes, such as implantation. The aim of this study was to investigate the effect of BVDV infection on uterine PG production and PR using an in vitro PR model. Bovine uterine endometrial cells isolated from ten BVDV-free cows were cultured and treated with 0 or 100 ng/mL interferon-τ (IFNT) in the absence or presence of non-cytopathic BVDV (ncpBVDV). PGFα2 and PGEα2 concentrations in the spent medium were measured using radioimmunoassays, and in the treated cells expression of the genes associated with PG production and signalling was quantified using qPCR. The results showed that the IFNT challenge significantly stimulated PTGS1 and PTGER3 mRNA expression and PGEα2 production; however, these stimulatory effects were neutralised in the presence of ncpBVDV infection. ncpBVDV infection significantly increased PTGS1 and mPGES1 mRNA expression and decreased AKR1B1 expression, leading to increased PGEα2 and decreased PGFα2 concentrations and an increased PGEα2:PGFα2 ratio. The other tested genes, including PG, ESR1, OXTR, PTGS2, PTGER2 and PTGFR, were not significantly altered by IFNT, ncpBVDV or their combination. Our study suggests that BVDV infection may impair PR by (1) inhibiting the effect of IFNT on uterine PG production and (2) inducing an endocrine switch of PG production from PGFα2 to PGEα2 to decrease uterine immunity, thereby predisposing the animals to uterine disease.

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Introduction

Poor reproductive performance, of which early embryonic mortality is a major component, causes major economic loss to the dairy industry (De Vries 2006). Embryonic mortality rates in cattle can be as high as 40%, with 70–80% of losses occurring before day 16 of gestation (Diskin et al. 2011). Following successful ovulation and subsequent fertilisation, production of interferon-τ (IFNT) by the trophectoderm begins at around day 8 of gestation and increases dramatically during conceptus elongation (Kimura et al. 2004). A sufficient threshold level of IFNT must be reached by day 16 to ensure pregnancy recognition (PR) and prevention of luteolysis (Forde et al. 2011, Lonergan & Forde 2014). IFNT acts in a paracrine manner on the uterine endometrium to develop a receptive environment. This involves changes in the uterine epithelium that are tightly regulated by steroid hormones, type I interferons, cytokines, prostaglandins (PGs) and growth factors and their receptors (Forde et al. 2011, Dorniak et al. 2012, Spencer et al. 2013, Lonergan & Forde 2014). Many reproductive processes, such as luteolysis, PR and implantation, involve inflammation and associated up-regulation of inflammatory mediators (e.g. PGs) and recruitment of immune cells to the uterine endometrium (Jabbour et al. 2009). PGEα2 (a vasodilator) is luteotrophic for maintaining progesterone secretion by the corpus luteum, whereas PGFα2α (a vasoconstrictor) is luteolytic (Weems et al. 2006). In PG production pathways, two series PGs, such as PGEα2 and PGFα2α′ are produced from arachidonic acid (AA). PG-endoperoxide synthase isozymes (PTGS1 and PTGS2) catalyse AA into PGH2, and PGH2 is converted into PGFs by the action of PGE synthase (PGES), and into PGFs by the action of PGF synthase (PGFS) (Wathtes et al. 2007). There are many isoforms for both PGES and PGFS. Previous studies have shown that mPTGES1 and AKR1B1 are the predominant
isoforms for PGE\(_2\) and PGE\(_{2\alpha}\) production, respectively, in bovine endometrium (Fortier et al. 2008).

During PR, IFNT inhibits the up-regulation of oxytocin (OXT) receptors (OXTR) in the uterine epithelium, thereby preventing the pulsatile release of PGE\(_{2\alpha}\) which is necessary for luteolysis (Wathes & Lamming 1995, Mann et al. 1999). In contrast, basal release of PGs, such as PGE\(_2\) and PGE\(_{2\alpha}\), from both the conceptus and the maternal uterine endometrium rises during early pregnancy (Zarco et al. 1988, Dorniak et al. 2013). This increased PG production may facilitate the effects of IFNT on the expression of endometrial type I IFN-stimulated genes (ISGs), a vital component of PR (Spencer et al. 2013).

Bovine viral diarrhoea virus (BVDV), a single-stranded RNA virus, causes widespread infection in the cattle worldwide. The reproductive system, including the uterine endometrium, is a major site for infection, maintenance and spreading of the virus (Grooms 2004, Lanyon et al. 2014). BVDV infection can have a significant impact on all stages of pregnancy, including causing early embryonic death, such that reproductive losses are one of the most important consequences of the disease (Grooms 2004). Our recent studies demonstrated that infection of bovine endometrial cells in vitro with non-cytopathic BVDV (ncpBVDV) caused many significant changes in pathways associated with innate immunity, particularly those associated with type I interferon signalling (Oguejiofor et al. 2015a).

ncpBVDV thus inhibited the expression of many ISGs that are also involved in PR, such as IFITs, BST2, MX2, OAS1, USP18 and RSAD2 (Forde et al. 2011). Some of these genes are also regulated by PGs (Spencer et al. 2013). In monocytes, BVDV infection stimulated the production of PGE\(_2\) (Welsh et al. 1995). When bovine endometrial cells were also exposed to bacterial lipopolysaccharide (LPS), ncpBVDV increased the expression of PLA2G4F and PTGES (mPGES1) (Oguejiofor et al. 2015a). PLA2G4F belongs to the family of PLA2 enzymes that are essential for the initial release of AA in PG biosynthesis (Tithof et al. 2007), whereas PTGES subsequently catalyses the isomerisation of PGH\(_2\) to PGE\(_2\) (Samuelsson et al. 2007).

These results suggested that ncpBVDV infection in cows may (1) interfere with the normal PR signals in response to IFNT and (2) increase endometrial PGE\(_2\) production following bacterial infection, potentially compromising the innate immune response. To explore these possibilities further, this study mimicked PR by stimulating bovine endometrial cells with IFNT in the presence or absence of ncpBVDV infection and investigated the effects of these treatments on PG signalling pathways.

**Materials and methods**

All reagents were purchased from Sigma or BDH Merck Ltd (Leics, UK) unless otherwise stated. All culture media used included 50,000 units/L penicillin and 50 mg/L streptomycin. All culture media and serum used were certified BVDV free.

**Animal, cell isolation and culture**

Fresh and apparently healthy uteri from cows in the early luteal phase of the oestrous cycle were collected at the local abattoir. Uterine endometrial cells (a mixture of primary epithelial and stromal cells) were isolated and cultured following the methods described previously (Cheng et al. 2013, Oguejiofor et al. 2015b). Briefly, under sterile conditions, strips of intercaruncular endometrium were separated and put into serum-free Dulbecco’s Modified Eagle’s Medium/Nutrient Mixture F-12 Ham (DMEM/F12 medium) (Sigma) and chopped into 1 mm\(^2\) cubes. They were digested for 90 min at 37°C in a medium containing 100 mg bovine serum albumin (BSA; Sigma), 50 mg trypsin III (Worthington, Lakewood, NJ, USA) and 50 mg collagenase A (Roche) per 100 mL of Hank’s balanced salt solution (HBSS; Sigma). After filtration through a 100 µm mesh into 50 mL Falcon vials, the suspension was re-suspended with HBSS containing 10% foetal bovine serum (FBS; PAA, Somerset, UK) and 3 µg/mL trypsin inhibitor (Sigma) and centrifuged at 100 g at 10°C for 10 min. After two repetitions of the above washing procedures, the cells were suspended in the culture medium (DMEM/F12 medium with 10% FBS) and plated in 24-well IWKI microplates (Sitecth DIV; Asahi Techno Glass, Funabashi, Japan) at 2 mL per well containing 0.5 × 10\(^6\) cells (day 1). Culture medium was changed every 48 h to allow the cells to grow. The composition of the cell population was confirmed using immunocytochemical staining validated in our laboratory (Oguejiofor et al. 2015b). After culturing for 8 days (day 8), epithelial cells comprised about 90% and stromal cells about 10% of the population. Contamination of immune cells was negligible (<0.001%).

**BVDV test for experimental samples**

The experiment was designed to use BVDV-free bovine endometrial samples and FBS. The examination was carried out using a PCR method reported previously (Vilcek et al. 1994, Pinheiro de Oliveira et al. 2013). The primer pair for the PCR was as follows: forward (ATGCCCTAGTAGGACTAGCA; position 108–128) and reverse (TCAACTCCATGTGCCATGTAC; position 395–375) with an expected product size of 288 bp, which are the highly conserved 5’ non-coding/non-structural coding regions of the pestivirus BVDV genome strain NADL (Vilcek et al. 1994). Total RNAs in the uterine endometrial tissue and FBS were extracted using a Qiagen RNA Lipid MiNi kit (Qiagen) following the supplied protocol. RNA was reverse transcribed into cDNA using a cDNA synthesis kit supplied by PCRBiosystems (London, UK), and PCR was performed using the G-Storm thermal cycler (G-Storm Ltd, Somerset, UK) and Qiagen Multiple PCR kit (Qiagen). The testing system also included a BVDV-positive control prepared using the pT7Blue-2 blunt vector, linearised (Novagen, Cambridge, MA, USA), reverse transcription-negative controls and a housekeeping gene β-actin (ACTB, see Table 1 for its primers) to verify that PCR reagents and reaction volumes were free of genomic DNA contamination. The above testing was carried out on the day of collection of uterus in parallel with the cell isolation and culture. The endometrium initially tested negative for BVDV was used for further experiments.
Propagation of ncpBVDV

The ncpBVDV (Pe515nc strain) was acquired from the BVDV Research Group, Royal Veterinary College, UK. This strain was isolated from a cow diagnosed with mucosal disease and virologically cloned as non-cytopathogenic virus. To provide adequate infectious BVDV for the present experiments, the virus stock was propagated to achieve a 50% tissue culture infective dose (TCID\textsubscript{50}) of 5 × 10\textsuperscript{5} per mL following the method used in our group (Oguejiofor \textit{et al.} 2015\textit{a}).

Experimental protocols

The experiments of ncpBVDV infection and IFNT challenge were carried out in the endometrial cells from ten cows confirmed BVDV negative tested using the above method. Cells from each cow were collected as a batch and grown in two 24-well plates as described previously (day 1). On day 4 of the culture when the cells grew to approximately 70% of confluence, FBS in the culture medium was reduced to 5% (maintenance medium, MM) to prevent overgrowth of the cells. The cells from each cow were divided into four treatment groups: control (CONT), IFNT, ncpBVDV and IFNT+ncpBVDV. Wells treated with the virus were maintained in separate plates from those treated without the virus to prevent cross-contamination. For the cells designated as BVDV infected, 0.25 mL MM containing Pe515nc BVDV at a multiplicity of infection (MOI) of 0.1 was added for 3 h to allow for virus infection. The same procedures, but with 0.25 mL MM only, were carried out for the cells designated as non-infected controls. The volume in all wells was made up to 1 mL with MM and the medium was changed after 2 days. IFNT treatment was carried 4 days after infection (day 8). For the wells specified for IFNT treatment, the medium was replaced with 1 mL MM containing 100 ng IFNT (recombinant ovine IFNT; Cell Sciences, Canton, MA, USA) and incubated for 24 h. The other wells were changed and incubated with 1 mL MM. The spent medium was harvested (on day 9) and stored at −20°C for PG quantification, and the treated cells were used for total RNA extraction.

Assessment of BVDV cell infection and cell viability

Bovine endometrial cell infection with ncpBVDV was confirmed using both the PCR method with the extracted RNA as described above and an indirect enzyme (alkaline phosphatase) immunostaining procedure as described previously (Oguejiofor \textit{et al.} 2015\textit{a}). The cell viability after exposure to the infection and treatment was assessed using an MTS reduction assay method as described previously (Oguejiofor \textit{et al.} 2015\textit{b}).

Quantification of PGs using radioimmunoassay (RIA)

The concentrations of PGE\textsubscript{2} and PGE\textsubscript{2a} in the spent medium were measured with charcoal–dextran-coated RIA methods described previously (Cheng \textit{et al.} 2001). The PG antisera were a kind gift from Dr N L Poyser (University of Edinburgh, Edinburgh, UK). The cross-reactivities for PGE\textsubscript{2} antiserum were 23, 100 and 15% with PGE\textsubscript{1}, PGF\textsubscript{2a} and PGE\textsubscript{3}, respectively, and those for PGF\textsubscript{2a} antiserum were 34, 100 and 25% with PGE\textsubscript{1}, PGF\textsubscript{2a} and PGE\textsubscript{3a} respectively. The standards for PGE\textsubscript{2} and PGF\textsubscript{2a} were purchased from Sigma, and the tritiated tracers of PGE\textsubscript{2} ([5,6,8,11,12,14,15(16)-3H]-PGE\textsubscript{2}) and PGF\textsubscript{2a} ([5,6,8,9,11,12,14,15(11)-3H]-PGF\textsubscript{2a}) were supplied by PerkinElmer. The samples were diluted (×10–200) in the RIA buffer. This allowed the sample PG concentrations to fall within the analytical ranges of the RIAs (0.02–5.0 ng/mL for PGE\textsubscript{2} and 0.01–2.5 ng/mL for PGF\textsubscript{2a}), and the interruption from the medium contents with the RIAs was minimised, making extraction unnecessary. The limits of detection were 2 pg/tube and 1 pg/tube for PGE\textsubscript{2} and PGF\textsubscript{2a} respectively. The intra-assay and inter-assay coefficients of variation were 3.5 and 6.3% for PGE\textsubscript{2} (n = 6) and 4.1 and 9.6% for PGF\textsubscript{2a} respectively (n = 6).

RNA extraction

After collection of spent medium, the cells from each treatment group (six wells) were pooled and the total RNA extraction was carried out using RNeasy Mini Kits (Qiagen) following the supplier’s protocol. The concentrations and purity of RNA were measured using a NanoDrop ND-1000 spectrophotometer (NanoDrop Technologies Inc., Wilmington, DE, USA).

Primer design and PCR

DNA sequences for all primers were obtained from GenBank at NCBI (http://www.ncbi.nlm.nih.gov/Database/index.html) and the primers were designed using a “Primer 3” web-based programme (http://frodo.wi.mit.edu/primer3). Their alignment specificity and quality were checked using the Blast tool at NCBI (http://www.ncbi.nlm.nih.gov/blast/). The detailed information of the primers is given in Table 1. The primers were made by Eurofins MWG Operon (Ebersberg, Germany). RNA (1 μg) was reverse transcribed into cDNA using a cDNA synthesis kit supplied by PCRBiosystems following the supplier’s protocol. The resulting cDNA (20 μL) was diluted in nucleoside-free water up to 100 μL.

PCR for the tested genes was performed using the G-Storm thermal cycler and QiaGen Multiple PCR kit (Qiagen), and the DNA products for each gene were used for (1) verifying the primer specificity using electrophoresis on a 2% (w/v) agarose gel and (2) preparation of the DNA standards for each gene in the following quantitative PCR (qPCR) procedures.

qPCR analysis for gene expression

The concentrations of all the ten target genes (PTGS1, PTGS2, mPGES1, AKR1B1, PTGER2, PTGER3, PTGFR, OXTR, PGR and ESR1) and four reference genes (GAPDH, RPL19, ACTB and 18sRNA) were quantified using qPCR via an absolute quantification approach following the method described previously (Cheng \textit{et al.} 2013). The DNA amplified from cDNA used for standards in the qPCR assay was purified using a QIAquick PCR purification kit (Qiagen), and their quality and concentrations were determined with the NanoDrop

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ND-1000 spectrophotometer. Eight standards were prepared from 1 to $1 \times 10^{-7}$ ng/mL. Annealing and amplicon-specific melting temperatures of the primers were determined using a gradient function of the qPCR machine (CFX96 Real-Time System DNA; Bio-Rad Laboratories) with eight identical reactions containing 2 ng of DNA standard, 10 µl Sygreen Mix (PCRBiosystems), 0.8 µl of each 10 µM forward and reverse primer, and nucleate-free water added up to 20 µl. The optimised annealing temperatures are given in Table 1.

To quantify the expression levels of each gene, the qPCR assay contained a standard curve, no template control (NTC) and a sample cDNA in duplicate with the same final volume on CFX96 Real-Time Systems (Bio-Rad). Each well contained 5 µl cDNA standard or samples, 10 µl Sygreen Mix (PCRBiosystems), 0.8 µl of 10 µM forward primer, 0.8 µl reverse primer and 3.4 µl nucleate-free water following the protocol supplied by PCRBiosystems and using the optimised annealing temperatures shown in Table 1. This included an initial Taq activation for 2 min followed by 38 cycles of denaturation (95°C), annealing (the annealing temperatures are given in Table 1) and extension (63°C). An amplicon-specific fluorescence acquisition reading based on the melting temperature obtained in the above gradient test was applied to avoid any noise from smaller non-specific products such as dimers before the product acquisition. The results were analysed using the CFX Manager Software package (Bio-Rad).

The limit of quantification was $1 \times 10^{-6}$ to $1 \times 10^{-7}$ ng/mL for all tested genes. In order to minimise variation, RT for all samples was performed in one assay. For each gene, the standards and all samples were prepared under the same conditions in a single plate with the same master mix of reagents.

### Table 1 Oligonucleotide primer sequence information.

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**Effect of ncpBVDV, IFNT and their combination on PG production by uterine endometrial cells**

The results showing the effect of ncpBVDV, IFNT and their combination on PG production by uterine endometrial cells are given in Fig. 1. ANOVA with repeated measurements showed that the differences of PGE₂ concentrations between treatment groups were significant (P<0.018). Fisher's LSD multiple comparisons illustrated that both IFNT challenge and ncpBVDV infection alone significantly stimulated PGE₂ production (P<0.05), whereas the stimulatory effect was neutralised when IFNT was combined with ncpBVDV (P<0.05) (Fig. 1A).

Compared with CONT, IFNT challenge appeared to decrease PGF₂α production; however, statistical significance was not achieved (P>0.05). In the cells infected with ncpBVDV, concentrations of PGF₂α were significantly lower than in the CONT cells (P<0.05). When the ncpBVDV-infected cells were challenged with IFNT, PGF₂α concentrations were significantly lower than in the CONT cells (P<0.05), and the concentrations were slightly lower than in the cells infected with ncpBVDV alone, although the difference was not statistically significant (P>0.05) (Fig. 1B).

Compared with the CONT cells, in the ncpBVDV-infected cells, the ratios of PGE₂ to PGF₂α significantly increased (P<0.05). Both IFNT and IFNT+ncpBVDV treatments appeared to increase the ratio of PGE₂:PGF₂α produced; however, the differences were not statistically significant due to large variations between animals (P>0.05) (Fig. 1C).

**Effect of ncpBVDV, IFNT and their combination on the expression of selected reference genes in uterine endometrial cells**

We selected 18SrRNA, ACTB, RPL19 and GAPDH as potential reference genes for qPCR. The results showed, however, that the expression of 18SrRNA, ACTB and RPL19 was significantly altered by IFNT, ncpBVDV or their combination (P<0.05–0.01). Only GAPDH expression was not affected by any of the above treatments (P>0.05) (Fig. 2A, B, C and D). Subsequent gene expression measurements have therefore been presented as absolute values given as femto gram per micro gram reverse-transcribed total RNA.

**Effect of ncpBVDV, IFNT and their combination on expression of PGR, ESR1 and OXTR in uterine endometrial cells**

In the bovine endometrial cells, expression of OXTR mRNA was relatively higher (1879±419 fg/μg RNA), whereas ESR1 mRNA expression was very low (0.003±0.0008 fg/μg RNA). The effects of all tested treatments on the expression of PGR, ESR1 and OXTR mRNA were moderate and no statistical significance was achieved as tested using ANOVA with randomised block design (Fig. 3A, B and C).

**Effect of ncpBVDV, IFNT and their combination on the expression of PTGS1, PTGS2, mPGES1 and AKR1B1 in uterine endometrial cells**

In the cells infected with ncpBVDV, expression of PTGS1 mRNA was significantly higher than in the CONT
cells ($P<0.05$). IFNT challenge significantly induced PTGS1 mRNA expression ($P<0.01$); however, in the presence of ncpBVDV infection, this stimulatory effect was completely neutralised ($P<0.05$) (Fig. 4A). In the cells infected with ncpBVDV, PTGS2 mRNA expression appeared to be lower than other groups; however, statistical significance was not achieved ($P>0.05$) (Fig. 4B). Figure 4C demonstrated that ncpBVDV infection stimulated mPGES1 mRNA expression by up to three-fold ($P=0.0003$) compared with the CONT cells, whereas neither IFNT nor IFNT+ncpBVDV challenge affected its expression ($P>0.05$). As shown in Fig. 4D, neither IFNT nor ncpBVDV alone altered AKR1B1 mRNA expression significantly ($P>0.05$), whereas their combination led to a significant decrease in its expression ($P<0.05$).

**Effect of ncpBVDV, IFNT and their combination on the expression of PTGER2, PTGER3 and PTGFR in uterine endometrial cells**

As shown in Fig. 5A and C, the expression of PTGER2 and PTGFR mRNA was not significantly changed by treatment with IFNT, ncpBVDV or their combination ($P>0.05$). Compared with the CONT, treatment with ncpBVDV alone only moderately reduced PTGER3

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*Figure 2* Effect of non-cytopathic bovine viral diarrhea virus (ncpBVDV), interferon-τ (IFNT) and their combination on the reference gene expression of (A) 18SrRNA, (B) ACTB, (C) RPL19 and (D) GAPDH by uterine endometrial cells isolated from BVDV-free cyclic cows. When cells were grown to approximately 70% confluence after 4 days of culture, ncpBVDV was inoculated at 0 or a multiplicity of infection of 0.1. Four days following inoculation, the cells were challenged with IFNT at 0 or 100 ng/mL for 24 h. Total RNA was extracted from the treated cells and the gene expression was quantified using an absolute qPCR approach. There were three replicates per treatment for each cow and ten cows per treatment. CONT: control without ncpBVDV and IFNT. The columns labelled with different letters were significantly different at $P<0.05–0.01$ (a>b).

*Figure 3* Effect of non-cytopathic bovine viral diarrhea virus (ncpBVDV), interferon-τ (IFNT) and their combination on (A) PGR, (B) ESR1 and (C) OXTR expression by uterine endometrial cells isolated from BVDV-free cyclic cows. When cells were grown to approximately 70% confluence after 4 days of culture, ncpBVDV was inoculated at 0 or a multiplicity of infection of 0.1. Four days following inoculation, the cells were challenged with IFNT at 0 or 100 ng/mL for 24 h. Total RNA was extracted from the treated cells and the gene expression was quantified using an absolute qPCR approach. There were three replicates per treatment for each cow and ten cows per treatment. CONT: control without ncpBVDV and IFNT.
mRNA expression \((P > 0.05)\). In the cells stimulated with IFNT, PTGER3 mRNA expression was up-regulated by up to 2.5-fold \((P < 0.01)\), whereas in the presence of ncpBVDV infection, the stimulatory effect of IFNT on PTGER3 mRNA expression was intensively inhibited \((P < 0.01)\) and the expression was even lower than that in the CONT cells \((P < 0.01)\) (Fig. 5B).

**Discussion**

PR in cows is initiated by IFNT release from the conceptus acting on the uterine endometrium leading to maintenance of the corpus luteum. Regulation of the synthesis and release of PGE_2 and PGF_2α are crucial to this process (Bazer 2013, Lonergan & Forde 2014). Failure of PR is a significant risk factor for embryonic mortality (Diskin et al. 2011, Forde et al. 2011). BVDV infection can cause early embryonic death (Grooms 2004); however, little evidence is currently available on the mechanisms involved. In this study, we investigated the effect of BVDV infection on PG production and signalling pathways in bovine endometrium. We have demonstrated that (1) ncpBVDV infection induced an endocrine switch of PG production and signalling from PGF_{2α} to PGE_2 and (2) IFNT stimulates uterine PGE_2 production and its signalling pathway; however, this stimulatory effect is abolished by ncpBVDV infection.

In addition to PR, IFNT possesses antiviral, anti-proliferative and immunosuppressive activities (Pontzer et al. 1988, 1991, Kohara et al. 2012). It was previously reported that the replication of ncpBVDV was completely suppressed by bovine IFNT treatment in cultured bovine muscular cells (Kohara et al. 2012). In our study, after IFNT challenge for 24h, ncpBVDV RNA was well detected in all infected endometrial cells using a PCR method, whereas it was negative in the ncpBVDV-free cells. Using a qPCR method, we confirmed that there was no significant difference in the ncpBVDV mRNA expression between the ncpBVDV and IFNT+ncpBVDV groups. Kohara et al. (2012) carried out infection of their cells after IFNT treatment, whereas in our study, infection was carried out before challenge. The different cell types used with their different structures and cellular signalling systems may also be important in this response. Our data support previous findings that the uterine endometrium is one of the major sites for infection, maintenance and spreading of BVDV in the cattle population (Grooms 2004, Lanyon et al. 2014).

In the non-pregnant uterine endometrium, PGF_{2α} is predominantly produced by uterine epithelial cells and PGE_2 by stromal cells (Danet-Desnoyers et al. 1994, Charpigny et al. 1999). In this study, the endometrial cells produced significant amounts of both PGE_2 (4.8 ± 0.5 ng/mL) and PGF_{2α} (13.3 ± 2.2 ng/mL). Together with the immunocytochemical staining results, this confirmed the presence of both epithelial and stromal cell populations. The paracrine interactions between these two cell types are important for the investigation of overall response of the endometrium to IFNT challenge and ncpBVDV infection.

In ruminants, although IFNT inhibits the pulsatile release of PGE_{2α} by the uterine epithelium, the basal release of PGE_2 and PGF_{2α}, from both the conceptus and the endometrium increases during early pregnancy (Zarco et al. 1988, Dorniak et al. 2013). PGs secreted by both epithelial and stromal cells co-ordinate with the IFNT to regulate uterine endometrial functions. This is important to develop a receptive environment for conceptus development, thereby promoting elongation.

**Figure 4** Effect of non-cytopathic bovine viral diarrhoea virus (ncpBVDV), interferon-τ (IFNT) and their combination on mRNA expression of (A) PTGS1, (B) PTGS2, (C) mPGES1 and (D) AKR1B1 by uterine endometrial cells isolated from BVDV-free cyclic cows. When cells were grown to approximately 70% confluence after 4 days of culture, ncpBVDV was inoculated at 0 or a multiplicity of infection of 0.1. Four days following inoculation, the cells were challenged with IFNT at 0 or 100 ng/mL for 24 h. Total RNA was extracted from the treated cells and the gene expression was quantified using an absolute qPCR approach. There were three replicates per treatment for each cow and ten cows per treatment. CONT: control without ncpBVDV and IFNT. The columns labelled with different letters were significantly different at \(P < 0.05\) – 0.01 (a > b > c).
production was significantly up-regulated, whereas the PGF$_{2\alpha}$ production was not altered. This agreed with the previous finding that IFNT does not affect the expression of PTGS2, the rate-limiting enzyme in PG synthesis, but does stimulate PGE$_2$ production by cells of the bovine uterus (Dorniak et al. 2013). We also found that IFNT challenge increased PTG51 and PTGER3 gene expression by up to 2.5-fold. Such an increase in both PGE$_2$ and its receptor indicates that IFNT favours the PGE$_2$ signalling pathway that would benefit maintenance of the corpus luteum as PGE$_2$ is a luteotrophin (Pratt et al. 1977, Dorniak et al. 2013). In the presence of ncpBVDV infection, however, the stimulatory effects of IFNT on PGE$_2$ production and its receptor PTGER3 expression were neutralised, and the PGE$_2$ concentrations were even slightly lower than in the CONT cells. When the ncpBVDV-infected cells were treated with IFNT, basal PGF$_{2\alpha}$ production and AKR1B1 mRNA expression were also lower than the CONT. AKR1B1 is a predominant isoform for PGF$_{2\alpha}$ production (Fortier et al. 2008). Therefore, decreased or disrupted basal PG production may contribute to failure of pregnancy establishment and maintenance following ncpBVDV infection.

These results also suggest that IFNT treatment may act on uterine epithelial and stromal compartments differentially as PGE$_2$ is produced by stromal cells and PGF$_{2\alpha}$ by epithelial cells. This supports previous studies in ewes that have shown cell-specific effects of IFNT for many ISGs crucial for developing a receptive endometrium and implantation (Bazer 2013, Dorniak et al. 2013). In support of this, intra-uterine infusion of meloxicam, a PTGS inhibitor, prevented uterine PG production and led to failure of conceptus elongation in ewes (Diskin et al. 2011). In cows, successful development to term and delivery of a live calf could be predicted by the expression of PTGS2 in day 7 blastocysts (Bazer 2013). In this study, when the endometrial cells isolated from cyclic cows were given IFNT for 24 h, their PGE$_2$ and implantation (Bazer 2013, Dorniak et al. 2013). In support of this, intra-uterine infusion of meloxicam, a PTGS inhibitor, prevented uterine PG production and led to failure of conceptus elongation in ewes (Diskin et al. 2011). In cows, successful development to term and delivery of a live calf could be predicted by the expression of PTGS2 in day 7 blastocysts (Bazer 2013). In this study, when the endometrial cells isolated from cyclic cows were given IFNT for 24 h, their PGE$_2$
In summary, IFNT challenge stimulated uterine PTGS1 and PTGER3 mRNA expression and PGE₂ production; however, these stimulatory effects were neutralised in the presence of ncpBVDV infection. The interruption of IFNT-induced PG production and signalling by BVDV infection may directly cause failure of PR. ncpBVDV infection stimulated PTGS1 and mPGES1 mRNA expression and moderately suppressed AKR1B1 expression, leading to increased PGE₂ and decreased PGF₂α concentrations and an increase in PGE₂:PGF₂α ratios in bovine uterine endometrium. This endocrine switch of PG production from PGF₂α to PGE₂ may decrease uterine immunity to predispose the animals to uterine diseases, indirectly leading to failure of PR. Our results thus suggest a mechanism whereby ncpBVDV infection in cows may cause early embryonic death and reduced fertility.

Declaration of interest
The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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