Bovine embryo-oviduct interaction in vitro reveals an early cross talk mediated by BMP signaling

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Abstract

Signaling components of bone morphogenetic proteins (BMPs) are expressed in an anatomic and temporally regulated fashion in bovine oviduct. However, a local response of this signaling to the presence of the embryo has yet to be elucidated. The aim of the present study was to evaluate if early embryo-oviduct interaction induces changes in the gene expression of BMP signaling components. For this purpose, we used an in vitro co-culture system to investigate the local interaction between bovine oviductal epithelial cells (BOEC) from the isthmus region with early embryos during two developmental periods: before (from the 2-cell to 8-cell stage) or during (from the 8-cell to 16-cell stage) the main phase of embryonic genome activation (EGA). Exposure to embryos, irrespective of the period, significantly reduced the expression levels of this signaling in the oviductal cells while increases them in the early embryo. This suggests that BMP signaling pathway could be involved in an early cross talk between the bovine embryo and the oviduct during the first stages of development.


Introduction

Maternal-embryo communication during the preimplantation period plays a critical role for the establishment and maintenance of pregnancy. Any disturbance in this reciprocal cross talk can lead to pregnancy failure and can also have long-term consequences in the developmental potential and health of the offspring (Wolf et al. 2003, Fazeli 2008, Rizos et al. 2016).

In mammals, the first maternal site that makes contact with the embryo is the oviduct. In the particular case of cattle, this contact is established during the first four days after fertilization (Maillo et al. 2016). Within this window of time, important developmental events occur in the embryo, including the first cleavage divisions (Lonergan et al. 1999) and the epigenetic reprogramming and activation of the embryonic genome (Memili & First 2000, Graf et al. 2014). To provide an optimal environment and to ensure the correct synchronization of these events, the oviduct needs to initiate a molecular dialog with the developing embryo. Studies in mice, pigs and horses have provided evidence for this initial cross talk, demonstrating that the presence of the embryo(s) produces transcriptional changes in the oviductal cells (Lee et al. 2002, Almiñana et al. 2012, Smits et al. 2016). In cattle, although the presence of a single embryo in vivo has no detectable effect on the oviductal transcriptome, the transfer of 50 embryos into the oviduct of heifers produces changes in the gene expression pattern of the oviductal cells, suggesting that the effect of the embryo, particularly in mono-ovulatory species, seems to be more local than was expected (Maillo et al. 2015).

It is known that epithelial cells lining the lumen of the mammalian oviduct can synthesize and secrete a wide range of proteins, including diverse growth factors and cytokines that can mediate a local interaction with the
embryo through different signaling pathways (Buhi et al. 2000, Lee & Yeung 2006, Aviles et al. 2010). Among the candidate signaling pathways that could actively participate in this local embryo-maternal cross talk in the oviduct, we focused our attention on the signaling mediated by bone morphogenetic proteins (BMPs). BMPs are a subfamily of growth factors that belongs to the TGF-β superfamily (Bragdon et al. 2011). These factors exert their effects as ligand dimers by binding to tetrameric complexes of type I (BMPR1A, BMPR1B or ACVR1A, also known as ALK3, ALK6 and ALK2 respectively) and type II (BMPR2, ACVR2A or ACVR2B) serine/threonine kinase receptors leading to signal transduction through SMAD-dependent pathways which regulates transcriptional responses (Beyer et al. 2013). Several BMPs are expressed in the maternal tract (ovary, uterus and placenta) and play important roles as autocrine and paracrine regulators of ovarian follicular development, blastocyst implantation in the uterus, and morphogenesis and organogenesis during embryo development (Shimasaki et al. 2004, Kishigami & Mishina 2005, Jones et al. 2006, Knight & Glister 2006). In addition, different components of the BMP signaling pathway are expressed by the embryo in a developmentally regulated fashion during the preimplantation development (Roelen et al. 1997, Zhang et al. 2007, Lee et al. 2014). Particularly in mice, knockout experiments and inhibition assays demonstrated that during the preimplantation period BMP signaling is critical for cell cleavage and the normal development of extra-embryonic lineages (Graham et al. 2014, de Mochel et al. 2015).

Previous studies have revealed that different members of the BMP family are expressed in the epithelial cells of the bovine oviduct in an anatomically and temporally regulated fashion during the estrous cycle (Garcia et al. 2014). In particular, BMP5 exhibits differential expression in the isthmus region and high transcriptional levels during the periovulatory phase. The presence of this factor in the oviductal fluid suggests that BMP ligands produced by the oviduct could have a paracrine action on the bovine embryo (Garcia et al. 2014). In fact, the addition of BMP5 to the culture medium during the first two days after fertilization produces an increase in the blastocyst rate and the relative mRNA abundance of pluripotency-related genes (Garcia et al. 2015). Interestingly, mRNA expression of BMP receptors is greater in early stage embryos (from 2-cell to 8-cell stage), suggesting that the embryo could be target of BMPs during its transit through the oviduct (Garcia et al. 2015). Moreover, in a recent study by RNA sequencing, Maillo and coworkers (Maillo et al. 2015) observed that the presence of multiple embryos induces changes in the expression levels of several genes in the bovine oviduct in vivo, included among them are various genes associated with the BMP signaling pathway. However, until now the participation of this signaling pathway in an initial cross talk between the embryo and the oviduct during the preimplantation period has not been explored in depth.

Considering this evidence, BMP signaling could be a candidate pathway for embryo-maternal communication during the preimplantation period. Within this context, the objective of this study was to evaluate whether early embryo-oviduct interaction induces changes in the gene expression levels of BMP signaling components both in the oviducal cells and in the preimplantation embryos. For this purpose, an in vitro co-culture system allowing a local and temporal interaction between early bovine embryos and isthmus epithelial cells was established.

Materials and methods

Chemicals

All reagents were purchased from Sigma Chemical Química S.A Company unless otherwise stated.

Bovine oviduct epithelial cell isolation and in vitro culture

Bovine oviducts at the early luteal phase (corresponding to Days 3–5 of the estrous cycle) were obtained from slaughtered heifers and selected based on ovarian morphology according to Ireland and coworkers (Ireland et al. 1980). The oviducts were sealed in a plastic bag and transported to the laboratory on ice within 2 h of slaughter. The surrounding fat and connective tissues were carefully removed and each oviduct was washed three times in sterile PBS (pH 7.4). The isthmus region was separated and the oviducal mucosa was collected by squeezing and was washed 2 times with PBS by centrifugation at 300 g for 10 min. The cell pellet was then resuspended in 1 mL of synthetic oviductal fluid (SOF) supplemented with 10% (v/v) of fetal calf serum (FCS) and was passed 10 times through a 25 G syringe needle to obtain a single cell suspension. Bovine oviduct epithelial cells (BOEC) were counted in a hemocytometer, diluted to a final concentration of 2 × 10⁶ cells/mL and then cultured in 500 µL of SOF + 10% FCS in four-well plates at 38.5°C, 5% CO₂, 5% O₂, 90% N₂, and saturated humidity until confluence. Half of the media (SOF + 10% FCS) was replaced every 48 h. On Day 5 of culture, the medium was changed to SOF with 5% FCS; and 24 h later, half of the media was renewed and the cells were used for co-culture with embryos.

Immunocytochemical analysis with antibodies anti-bovine-pancadherin (C1821), anti-bovine-pancytokeratin (C2931) and anti-bovine-vimentin (V2258), following the protocol described by Lopera-Vásquez and coworkers (Lopera-Vásquez et al. 2016), confirmed the epithelial nature of the cultured cells showing positive staining for the epithelial markers cadherin and cytokeratin, and negative staining for the fibroblast marker vimentin (Fig. 1A).
**Oocyte collection and in vitro maturation**

Immature cumulus-oocyte complexes (COCs) were recovered by aspirating follicles (2–8 mm in diameter) from ovaries of heifers slaughtered at local abattoir and *in vitro* maturation was performed as described by Lopera-Vásquez and coworkers (Lopera-Vásquez et al. 2015). Only oocytes with a compact, non-atretic cumulus of at least three layers and a homogeneous ooplasm (Class 1 and 2 COCs) were selected and matured for 24 h in 500 µL of maturation medium (TCM 199 (M4530) supplemented with 10% (v/v) FCS and 10 ng/mL epidermal growth factor (E4127)) in four-well dishes (50 COCs per well) at 38.5°C under an atmosphere of 5% CO$_2$, with maximum humidity.

**In vitro fertilization**

Frozen semen from a single Asturian Valley bull (ASEAVA, Asturias, Spain), previously tested for IVF, was thawed at 37°C in a water bath for 1 min and selected on a gradient of Bovipure (Nidacon Laboratories AB, Göthenborg, Sweden) following the protocol described by Lopera-Vázquez and coworkers (Lopera-Vázquez et al. 2016). Sperm concentration was determined and adjusted to a final concentration of $1 \times 10^6$ sperm cells/mL. Gametes were coincubated for 18–22 h in 500 µL of fertilization medium (Tyrode’s medium with 25 mM bicarbonate, 22 mM Na-lactate, 1 mM Na-pyruvate and 6 mg/mL fatty acid-free bovine serum albumin (BSA) supplemented with 10 µg/mL heparin sodium salt (Calbiochem)) in a four-well dish, in groups of 50 COCs per well under an atmosphere of 5% CO$_2$, with maximum humidity at 38.5°C.

**In vitro embryo culture and co-culture conditions**

After the fertilization period, presumptive zygotes were completely denuded of cumulus cells by vortexing and groups of 50 were initially cultured in 500 µL of SOF supplemented with 4.2 mM sodium lactate (L4263), 0.73 mM sodium pyruvate (P4562), 30 µL/mL basal medium eagle (BME) amino acids (B6766), 10 µL/mL minimum essential medium (MEM) amino acids (M7145), 1 µg/mL phenol red (P0290) and 3 mg/mL bovine serum albumin (BSA; A9647) in a four-well dish, under an atmosphere of 5% CO$_2$, 5% O$_2$ and 90% N$_2$ at 38.5°C. At 33 and 54 h post-insemination (hpi); those embryos that reach the 2-cell stage and the 8-cell stage respectively were selected and randomly cultured in groups of 30 embryos in SOF supplemented with 3 mg/mL of BSA or in SOF supplemented with 5% (v/v) of FCS in the presence or absence of a BOEC monolayer, depending on the experimental group to which they were assigned (see ‘Experimental design’ described below for more details). In order to limit and establish a local area of contact between the embryos and the BOEC monolayer, a nontoxic woven polyester mesh (Sefar Petex; Sefar, Bury, Lancashire, UK) was used with a grid size of 41 × 41 openings, covering an area of 121 mm$^2$ that fitted perfectly inside the well of a NuncTM 4-well dish. First of all, one mesh square containing delimited marked areas of 6 × 5 openings was placed outside at the bottom of the wells and was used as a guide to localize the co-culture area (Fig. 1B). Secondly, another mesh square of the same size was washed once with 70% ethanol, thrice with PBS and twice with SOF media, and then was introduced into the well so that the external and internal mesh were overlapped. Then, 30 embryos/well were

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**Figure 1 In vitro co-culture system.**

(A) Immunofluorescence analysis of BOEC monolayers showing positive staining for the epithelial markers, cadherin and cytokeratin, and negative staining for the fibroblast marker, vimentin. Cell nuclei were counterstained with Hoechst stain (blue). (B) Representative image of the polyester meshes employed to establish a local area of contact between the embryos and the BOEC monolayer. Thirty embryos/well were transferred to the mesh and placed in a 6 × 5 openings grid covering an area of 2.1 mm$^2$ over the monolayer. Mesh dimensions: 170 µm opening and 96 µm filament diameter.
transferred to the mesh and placed in a 6 × 5 grid, covering an area of 2.1 mm² over the monolayer (Fig. 1B). Preferably, the central grid was selected. Following culture, embryos were transferred and cultured in 25 μL droplets of SOF with BSA until Day 9 pi.

**RNA isolation and reverse transcription**

Five independent samples of BOEC cultured with and without embryo under each experimental condition, and also three independent groups of 10 embryos per stage (8-cell, 16-cell and blastocyst stage) and per experimental group were collected and processed for poly(A) RNA extraction. Poly(A) RNA was extracted using the Dynabeads mRNA Direct Extraction Kit (Dynal Biotech, Oslo, Norway) following the manufacturer’s instructions and with minor modifications as it was described previously by Bermejo-Alvarez and coworkers (Bermejo-Alvarez et al. 2008). After 10 min of incubation in lysis buffer with Dynabeads, poly(A) RNA attached to the Dynabeads was extracted with a magnet and washed twice in washing buffer A and washing buffer B. RNA was then eluted with Tris–HCl. Immediately after extraction, the reverse transcription (RT) reaction was performed as recommended by the manufacturer (Epicentre Technologies Corp, Madison, WI, USA). Briefly, oligo-dT (0.2 µM) and random primers (0.5 µM) were added to each RNA sample and were heated for 5 min at 70°C to denature the secondary RNA structure. Next, the tubes were incubated at 25°C for 10 min to promote the annealing of the primers. Then, the RNA was reverse-transcribed for 60 min at 37°C in a final volume of 50 μL containing 0.375 mM dNTPs (Biotools, Madrid, Spain), 6.25 U RNasin RNase inhibitor (Promega), 10× MMLV-RT buffer with 8 mM dithiothreitol and 5 U MMLV high performance reverse transcriptase (Epicentre Technologies Corp, Madison, WI, USA), followed by incubation at 85°C for 5 min to inactivate the RT enzyme.

**Gene expression analysis**

The mRNA expression levels of the selected genes were determined by real-time quantitative reverse transcription polymerase chain reaction (qRT-PCR) using specific primers designed with Primer-BLAST (http://www.ncbi.nlm.nih.gov/tools/primer-blast/) to span exon-exon boundaries when possible. Primers (Table 1) were previously validated for adequate primer efficiency; and specificity of their PCR products was confirmed by electrophoresis on a 2% agarose gel. All target genes showed efficiencies between 97 and 100% and correlation coefficients close to 1.0.

All PCR reactions were performed in a final volume of 20 μL, containing 0.25 mM of forward and reverse primers, 10 μL of GoTaq qPCR Master Mix (Promega) and 2 μL of each cDNA sample derived from BOEC (≈60 ng/μL), 8-cell embryos (≈4 ng/μL), 16-cell embryos (≈20 ng/μL) or blastocysts (≈60 ng/μL), using a Rotorgene 6000 Real Time Cycler (Corbett Research, Sydney, Australia) and SYBR Green as double-stranded DNA-specific fluorescent dye. For each experimental group, five different cDNA samples from BOEC and three different cDNA samples from each embryonic stage were used in two repetitions for all genes of interest. The PCR program consisted of an initial denaturalization step at 94°C for 2 min, followed by 35 cycles of denaturalization at 94°C for 10 s, annealing at 56°C for 30 s, extension at 72°C for 15 s and 10 s of fluorescence acquisition defined for each primer. At the end of each PCR run, melt curve analyses were performed for all products was confirmed by electrophoresis on a 2% agarose gel. All target genes showed efficiencies between 97 and 100% and correlation coefficients close to 1.0.

**Table 1** Set of primers used for qRT-PCR assays.

<table>
<thead>
<tr>
<th>Gene</th>
<th>Primer sequences (5′–3′)*</th>
<th>Amplicon length (bp)</th>
<th>GenBank accession number</th>
</tr>
</thead>
</table>
| BMP5  | Forward: GATGTGCTTGCTTTGCTT  
   Reverse: CCTGATGACCCGAGCTTTA | 271 | NM_001305016.1 |
| BMP7  | Forward: AACCATTGCGTGCAGAGCCG  
   Reverse: AGCCCGGACCAGGCAAACC | 250 | NM_001305016.1 |
| BMP1R1 | Forward: TTTGGGAATGGCTCGTCTGT  
   Reverse: AGCACAAATGGCCGGAACAC | 142 | NM_001305016.1 |
| BMP1R1B | Forward: TTTGGGAATGGCTGGCTAGGA  
   Reverse: GCCGAGCGCTTTTGATACAC | 132 | NM_001305016.1 |
| BMP2  | Forward: TCCTTCCGCCACACATGCTCT  
   Reverse: GTTCAGTGGAATGACCCAGG | 158 | NM_001304285.1 |
| SMAD1 | Forward: TGAGAACCCGCGCTCTGAGACG  
   Reverse: TCCCTGCAGGTTGTATTCTCGTCT | 99 | NM_001206015.1 |
| SMAD5 | Forward: TTTGCTCGGTCTGCCTACGTC  
   Reverse: TCCCTGCAGGCTATGCTGCTGCTC | 119 | NM_001206015.1 |
| SMAD6 | Forward: GGAGAAATCTGCTCTCAGGAGC  
   Reverse: CCGGCGGCTTAACCAACG | 242 | NM_001206145.1 |
| ID2   | Forward: CGACTGCACATCTGCTTCT  
   Reverse: AGACCCGAGTTGGATTTGCTGTG | 145 | NM_001304231.2 |
| H2AFZ | Forward: AGGAGCACTAGCAGCATGACGTTG  
   Reverse: CACAAGACAGGATATGCTGCTC | 209 | NM_174809.2 |
| ACTB  | Forward: GAGAAGCTGCTGACTGTCG  
   Reverse: CCAGACAGCCCGCTGTGGG | 263 | NM_001206145.1 |

*All the primers were designed with NCBI Primer-BLAST online tool of the National Centre for Biotechnology Information (http://www.ncbi.nlm.nih.gov/tools/primer-blast/).
genes to ensure single product amplification and exclude the possible interference of dimers.

Relative expression levels were quantified by the comparative cycle threshold (\(\Delta\Delta CT\)) method (Schmittgen & Livak 2008). Values were normalized using two housekeeping genes (\(H2AFZ\) and \(ACTB\)) that were tested in previous studies (Maillo et al. 2015). Fluorescence was acquired in each cycle to determine the threshold cycle during the log-linear phase of the reaction at which fluorescence increased above background for each sample. According to the comparative CT method, the \(\Delta CT\) value was determined by subtracting the mean CT value of the two housekeeping genes (\(H2AFZ\) and \(ACTB\)) for each sample from each gene CT value of the sample. Calculation of \(\Delta\Delta CT\) involved using the highest sample \(\Delta CT\) value (i.e. the sample with the lowest target expression) as an arbitrary constant to subtract from all other \(\Delta CT\) sample values. Fold changes in the relative gene expression of the target were determined using the formula \(2^{-\Delta\Delta CT}\) (Livak & Schmittgen 2001). All the study were carried out following the Minimum Information for Publication of Quantitative Real-Time PCR Experiments recommendations (Bustin et al. 2009).

**Experimental design**

**Experiment 1: Effect of BOEC co-culture on early embryo development in vitro**

In the first experiment, an in vitro co-culture system was established to provide a local and temporal interaction between a BOEC monolayer and early bovine embryos during two developmental periods: (a) Experimental Group 1 (G1): from 2-cell to 8-cell stage, before the main phase of embryonic genome activation (EGA); and (b) Experimental Group 2 (G2): from 8-cell to 16-cell stage, during the main phase of EGA (Fig. 2). To determine if this in vitro co-culture system had an effect on early embryonic development, cleavage rate and blastocyst yield were evaluated.

For this purpose, isthmus epithelial cells obtained from a pool of three post-ovulatory stage oviducts \((n=21, 3\) oviducts per experimental replicate) were cultured in a monolayer as described previously. On Day 6 of culture, the BOEC monolayer was cultured in the absence or presence of in vitro-produced embryos from 2- to 8-cell stage (G1 BOEC; 33–54 hpi) or from 8- to 16-cell stage (G2 BOEC; 54-98 hpi) in SOF medium supplemented with 5% FCS, under an atmosphere of 5% CO\(_2\), 5% O\(_2\), and 90% N\(_2\) at 38.5°C. As mentioned above, 30 embryos/well were placed in a 6 x 5 grid over the monolayer. In addition, as control without BOEC, groups of 30 embryos in both developmental stages were cultured either in SOF + 5% FCS (G1 FCS and G2 FCS) or in SOF + 3 mg/mL BSA (G1 BSA and G2 BSA). At 54 hpi (G1 BOEC/FCS/BSA) or 98 hpi (G2 BOEC/FCS/BSA), embryos that reached the 8- or 16-cell stage respectively were transferred to SOF + BSA and cultured until Day 9, maintaining the different experimental groups separately (Fig. 2).

Considering that during the experiment it was necessary to preselect the embryos at different stages of development, the developmental parameters were calculated as follows: (I) developmental rate at 33 hpi: percentage of presumptive zygotes that developed to the 2-cell stage at 33 hpi; (II) developmental rate at 54 hpi: percentage of selected 2-cell embryos that developed to the 8-cell stage at 54 hpi; (III) developmental rate at 98 hpi: percentage of selected 8-cell embryos that developed to the 16-cell stage at 98 hpi; and...
(IV) blastocyst yield: percentage of selected 8-cell embryos or 16-cell embryos that continued in culture and developed to the blastocyst stage at Days 7, 8 and 9 after insemination.

Seven experimental replicates were performed under the same assay conditions and a total of 2550 presumptive zygotes were in vitro cultured: 1192 were used for experimental groups 1 (G1 BOEC/FCS/BSA) and 1358 for groups 2 (G2 BOEC/FCS/BSA).

Experiment 2: Effect of early embryo-oviduct interaction in vitro on mRNA expression of BMP signaling components in oviductal epithelial cells

To evaluate whether in vitro interaction with early embryos induces changes in the gene expression levels of BMP signaling components in the oviductal cells, an expression analysis was performed by qRT-PCR in 5 samples of BOEC cultured with or without embryos before (G1 BOEC) or during EGA (G2 BOEC), obtained from 5 different experimental replicates of Experiment 1.

After the co-culture period, the polyester mesh and the embryos were removed and the BOEC monolayer was quickly washed twice with PBS. The cell area directly beneath the embryos was recovered from G1 BOEC at 54 hpi (BOEC G1+, n = 5) and from G2 BOEC at 98 hpi (BOEC G2+, n = 5) by mechanical scraping with a micropipette tip. At the same time, an area of the same size but from a different well of BOEC, cultured without embryos, was collected as control (BOEC G1− and BOEC G2−, n = 5). The BOEC samples collected were immediately snap-frozen in liquid nitrogen and kept at −80°C until RNA extraction.

The mRNA expression levels of two BMP ligands (BMP5/7), three BMP receptors (BMPR1A/1B/2), two signaling proteins (SMAD1/5), one inhibitor (SMAD6) and one target gene (ID2) were analyzed by qRT-PCR according to the procedures described above.

Experiment 3: Effect of early embryo-oviduct interaction in vitro on mRNA expression of BMP signaling components in preimplantation embryos

The objective of the third experiment was to evaluate if the in vitro interaction with the oviductal cells induces changes in the expression levels of BMP signaling components in the embryos at specific developmental stages. For this purpose, gene expression analysis by qRT-PCR was performed in three independent groups of 10 embryos per stage (8-cell, 16-cell and blastocyst stage) obtained from each experimental group that was cultured with or without BOEC, before or during EGA (corresponding to groups 1 (G1 BOEC/BSA/FCS) and groups 2 (G2 BOEC/BSA/FCS) respectively) in Experiment 1.

After the co-culture period, at 54 hpi (G1 BOEC/BSA/FCS) or 98 hpi (G2 BOEC/BSA/FCS), embryos that reached the 8- or 16-cell stage respectively were removed, quickly washed in PBS; and 10 embryos per stage and experimental group were immediately snap-frozen in liquid nitrogen and kept at −80°C until RNA extraction. The remaining embryos were transferred to 25 µl droplets of SOF + BSA under mineral oil and cultured until Day 9pi. At Day 7–8pi, pools of 10 expanding blastocysts per treatment were collected, snap-frozen and stored at −80°C until use for gene expression analysis.

The mRNA expression levels of two BMP ligands (BMP5/7), three BMP receptors (BMPR1A/1B/2), two signaling proteins (SMAD1/5), one inhibitor (SMAD6) and one target gene (ID2) were analyzed by qRT-PCR according to the procedures above.

Statistical analysis

Data were analyzed using SigmaStat 3.5 and SigmaPlot 10.0 statistical software (Systat Software, Richmond, CA, USA). Cleavage rate, blastocyst yield and relative mRNA abundance levels were analyzed using one-way analysis of variance (ANOVA), followed by Holm–Sidak post hoc test, when applicable, to determine statistical differences between the experimental groups. Probability values less than 0.05 (P < 0.05) were considered statistically significant.

Results

Experiment 1: Effect of BOEC co-culture on early embryo development in vitro

For all the experimental groups, only embryos that reached the 2-cell stage at 33 hpi were selected for the study. As shown in Fig. 3A cleavage rates at 33 hpi were similar for G1 and G2 (G1: 66.8 ± 2.0% vs G2: 67.6 ± 1.2%; P > 0.05). In general, most of the embryos reached the 2-cell stage at 33 hpi (G1: 53.9 ± 3.8% and G2: 58.8 ± 3.5%) and a low percentage of embryos were at a more advanced stage (G1: 12.8 ± 3.5% and G2: 8.8 ± 2.7%).

At 54 hpi in the G1 BOEC group, a lower proportion of embryos reached the 8-cell stage than control groups (G1 BOEC: 57.1 ± 4.9% vs G1 BSA: 78.6 ± 4.0% and G1 FCS: 79.0 ± 7.4% P < 0.05) (Fig. 3B). As a consequence, a significantly higher proportion of embryos with a delayed development (<8 cells) was observed in G1 BOEC than control groups (G1 BOEC: 42.9 ± 4.9% vs G1 BSA: 21.4 ± 4.0% and G1 FCS: 21.0 ± 7.4%, P < 0.05) (Fig. 3B).

Similarly, at 98 hpi, the G2 BOEC group showed a lower but not significant proportion of embryos reaching the 16-cell stage than control groups (G2 BOEC: 52.4 ± 8.4%, G2 BSA: 66.7 ± 5.9% and G2 FCS: 65.2 ± 6.8%) (Fig. 3C). In this case, a significantly higher proportion of embryos with a delayed development (<16 cells) was observed in the G2 BOEC group in comparison with the control groups (G2 BOEC: 47.6 ± 8.4% vs G2 BSA: 33.3 ± 5.9% and G2 FCS: 34.8 ± 6.8%, P < 0.05) (Fig. 3C).

Despite this initial delay observed in the developmental kinetics of the embryos, no significant differences were observed in the blastocyst rate between the control groups (G1 BSA/FCS and G2 BSA/FCS) and the groups that were co-cultured with BOEC before or during EGA (G1 BOEC or G2 BOEC) (Fig. 3D and E).
of BMP5, the expression of its mRNA was not detected in the oviductal cells cultured in vitro. In this case, to confirm that BMP5 primers work efficiently, samples of bovine oviduct epithelial cells freshly obtained from the isthmus region (BOEC in vivo) were used as positive control (data not shown).

**Experiment 3: Effect of early embryo-oviduct interaction in vitro on mRNA expression of BMP signaling components in preimplantation embryos**

Gene expression analysis in early stage embryos revealed a different transcriptional response to the interaction with BOEC, depending on the embryonic stage in which they interacted. At the 8-cell stage, embryos from G1 BOEC group showed a significant increase in the relative abundance of SMAD1 mRNA, compared to the levels detected in embryos derived from control groups (G1 BSA and G1 FCS) (Fig. 5A). Moreover, at the 16-cell stage, embryos from G2 BOEC group showed a significant increase in the relative abundance of BMPR1B, BMPR2 and ID2 mRNA compared to controls (G2 BSA and G2 FCS) (Fig. 5B). Despite these transcriptional changes detected in early stage embryos, in general, no differences were observed in the mRNA expression levels of BMP signaling components between the blastocysts obtained from the different experimental groups cultured in the presence or absence of BOEC (Fig. 6).

**Discussion**

The in vitro culture of oviductal cells has provided a suitable model for the study of the mechanisms involved in the interaction of gametes and embryos with the mammalian oviduct. Recent advances in in vitro modeling of the oviductal epithelium of different species, including bovine, porcine and human, have enabled the development of new culture techniques that better mimic the morphological and physiological characteristics of the oviduct (Gualtieri et al. 2012, Chen et al. 2013, Lawson et al. 2013, Simintiras et al. 2017). However, considering the increasing complexity of studies aimed at understanding the regulatory mechanisms that control embryo-maternal communication at a molecular level, it is necessary to establish novel strategies for in vitro culture that allow a more controlled interaction, in terms of area and contact time, between the embryo and the oviductal cells.

The main objective of the present study was to evaluate if the interaction of the embryo with the oviduct can induce changes in the gene expression of BMP signaling components. Considering the limitation of carrying out this study in vivo, an in vitro co-culture system was established to study the interaction between isthmus epithelial cells and in vitro-produced bovine embryos at developmental stages during which, under
physiological conditions, the embryo makes contact with the oviduct. To provide a local interaction, the area of contact between the embryo and the monolayer of oviductal cells was controlled and delimited with a polyester mesh. Moreover, a temporal interaction was established by analyzing two periods of embryonic development: before (from the 2-cell to 8-cell stage) and during (from the 8-cell to 16-cell stage) the main phase of embryonic genome activation (EGA). These two time windows were established taking into account that bovine embryonic genome is gradually activated, starting with minor activation, presumably initiated at the 2-cell stage and followed by a major activation at the 8- to 16-cell stage (Gad et al. 2012, Graf et al. 2014).

Embryos derived from the different experimental groups that were co-cultured with BOEC showed an early delay in developmental kinetics compared to embryos cultured without oviductal cells. However, this initial delay had no impact on the final production rate of blastocysts. A negative effect of the polyester mesh
was ruled out as control embryos cultured without BOEC showed normal development in the presence of the mesh. In concordance with this fact, Matoba and coworkers (Matoba et al. 2010) showed that the same type of mesh has no effect on embryonic development in vitro.

Other studies reported that the in vitro co-culture of BOEC with bovine embryos during the first 4 days (Rief et al. 2002, Cordova et al. 2014) or the entire 8 days of embryo development (Schmaltz-Panneau et al. 2014, 2015) can improve the early cleavage rate and/or the development to the blastocyst stage. Undoubtedly, the time of co-culture and the culture conditions are variables that have to be considered before evaluating and comparing these results. In particular, in the present study, the time of co-culture evaluated was shorter than that used in those studies. This allows to infer that an extended co-culture time between the embryo and the oviductal cells seems to be necessary to improve embryo development rate. In fact, other authors have shown that the embryotrophic effect of human oviductal cells cultured with mouse embryos is enhanced when the time of co-culture is longer and it lasts for at least four days (Xu et al. 2001).

On the other hand, gene expression analysis showed that the interaction with the embryos, before and during EGA, induced a significant decrease in the transcriptional levels of BMP signaling components in oviductal cells cultured in vitro. In particular, this reduction was observed in the mRNA levels of one ligand (BMP7), two receptors (BMPR1B and BMPR2), one signaling mediator (SMAD1) and two BMP target genes (SMAD6 and ID2). As can be seen, the effect is observed in all the genes encoding factors required for the signal transduction, suggesting a possible decrease in the activity of BMP pathway in the oviduct when the embryo is present.

It has been reported that BMP7 has a critical role in the oviduct, by acting as an inductor of apoptosis during tissue remodeling (Monroe et al. 2000). In fact, estrogen exerts a protective role by reducing the expression of this factor in the oviductal epithelium (Monroe et al. 2002, Kusumegi et al. 2004). Considering this, it would be interesting to explore if the downregulation of BMP7 expression, induced by the presence of the embryo, could be associated with a protective effect in order to maintain the integrity of the oviduct as the embryo passes.

In reference to BMP receptors, previous studies demonstrated the expression of BMPR1B and BMPR2 in the bovine embryo, showing high abundance of their transcripts at the early embryonic stages (from 2-cell stage to 8-cell stage) (Garcia et al. 2015). This temporal gene expression pattern of the receptors suggests that the embryo could respond to BMP ligands during the first developmental stages during transit through the oviduct. Based on the present study, the mRNA expression of the same receptors was detected in the oviductal cells and the presence of the embryo downregulated their expression. In this sense, the embryo could be sending...
specific signals to reduce the expression levels of these receptors in the oviduct to avoid competence. If so, BMP ligands present in the oviductal microenvironment could act preferably on the embryo. However, further studies are required to confirm this possibility.

Downregulation of mRNA expression of SMAD1 and the target genes ID2 and SMAD6 is also indicative of a suppressed BMP pathway as a response of the oviduct to the embryo. This coincides with a previous study performed in vivo in which the transfer of 50 embryos to the bovine oviduct produced changes in the gene expression pattern of the oviductal cells; SMAD6 was among the genes downregulated by the embryo presence (Maillot et al. 2015). This gene is induced via SMAD1/5 pathway, and at protein level, it acts as an inhibitor self-regulated by the BMP signaling pathway (Ishida et al. 2000, Li 2015). A recent study in mice demonstrated that SMAD signaling is required for structural integrity of the female reproductive tract and uterine function during early pregnancy (Rodriguez et al. 2016). However, until now, little is known about its role in the oviduct, in particular during the early stages of embryo development.

Similarly to other studies (Cordova et al. 2014, Schmaltz-Panneau et al. 2014, Maillot et al. 2015) these results confirm that bovine embryos can have a direct effect on the transcriptome of the oviductal cells. However, it is important to note that this study, in contrast to others, demonstrates that a short period of interaction with the early embryo is sufficient to induce specific transcriptional changes in the oviductal cells. Furthermore, a similar transcriptional response was observed irrespective of the embryonic stage, suggesting that oviductal cells do not show a temporal sensitivity to the different stages of embryo development, at least in the conditions assayed and for the genes analyzed.

On the other hand, gene expression analysis of the embryos revealed significant changes in the expression profile of BMP genes depending on whether they interact with oviductal cells before or during EGA. Embryos that interacted with BOEC before the main phase of EGA (G1 BOEC) showed higher levels of transcripts associated with BMP signaling. Particularly, a significant increase in the relative abundance of SMAD1 mRNA was observed at the 8-cell stage, compared to control groups cultured without BOEC. SMAD1 is a key component of the BMP signaling pathway that allows the transduction of the signal from membrane receptors to the nucleus (Bragdon et al. 2011). Depending on cell response, SMAD1 or SMAD5 can be activated (Li 2015). Thus, the significant increase in the mRNA levels for SMAD1 in the embryos suggests that the interaction with oviductal cells may induce the activation of BMP signaling in the embryos through the SMAD1 pathway. In mouse embryos, Smad1 coordinates the growth of extra-embryonic structures necessary to support embryo development within the uterine environment. As a consequence, the failure of Smad1 signal induces defects in extra-embryonic tissues and knockout embryos die due to problems in placentation (Tremblay et al. 2001, Arnold et al. 2006, Li 2015). Transcriptional analysis of mouse and bovine embryos reveals high abundance of SMAD1 mRNA at early stages before the main phase of EGA and a significant reduction after EGA up to the blastocyst stage (Wang et al. 2004, Lee et al. 2014) suggesting that this factor is necessary during the early stages in which the embryo transits through the oviduct. In this sense, our results suggest that early embryo-oviduct interaction could have a critical role in maintaining high transcriptional levels of this factor before EGA, during the early stages in which the embryo interacts with the oviduct.

In the case of the embryos that interacted with BOEC during EGA (G2 BOEC), a significant increase in the relative abundance of BMPR1B, BMPR2 and ID2 mRNA was detected at the 16-cell stage. This suggests that contact with the oviductal cells might induce the activation of the BMP signaling pathway in the embryo during the early stages of development. However, the precise mechanisms that control this upregulation are still unknown. It is well known that both BMP receptors, BMPR1B and BMPR2, play a critical role in preimplantation development and female fertility (Soyun et al. 2001, Nagashima et al. 2013, Graham et al. 2014, de Mochel et al. 2015). In addition, their transcript levels in the embryo are specifically elevated at early stages before EGA (Lee et al. 2014, Garcia et al. 2015).

In the case of ID2, it is a target gene of BMP signaling and it acts as a transcription factor required for maintenance and differentiation of trophoblast cells (Janatpour et al. 2000, Roberts et al. 2004, Schiffmacher & Keefer 2013). In agreement with our results, transcriptome analysis by microarray in in vivo-produced bovine embryos reported that ID2 increases its transcription during the activation of the bovine embryonic genome (Kues et al. 2008). The fact that the transcriptional levels of BMP receptors and the ID2 gene were higher during the early stages and stimulated by the interaction with the oviduct allows us to infer that BMP signaling pathway could be mediating an active cross talk between the embryo and the mother during the early stages of development.

Coincident with other studies (Wang et al. 2004, Zhang et al. 2007, Kues et al. 2008, Lee et al. 2014), our results show that during early embryogenesis, the transcript abundance of BMP signaling components is differentially regulated and that early maternal-embryo interaction can have a critical impact on the transcriptional regulation of this signaling. Moreover, the low levels detected at the blastocyst stage and the fact that at this stage there were no differences in the mRNA expression levels of BMP signaling components supports the hypothesis that BMP signaling is more important during the early stages of development. In fact, the inhibition of this signaling at the beginning
of embryo culture produces a significant reduction in the cleavage and developmental rates of mouse (de Mochel et al. 2015) and bovine embryos (La Rosa et al. 2011), suggesting that a correct balance of BMP signaling, during early stages, is needed for a proper preimplantation development. Thus, considering our results, the interaction with the oviduct could play a critical role to regulate this balance.

In conclusion, this study provides a new strategy for the study of the maternal-embryonic dialog and gives evidence about possible signaling pathways involved in the communication between the embryo and the bovine oviduct during the preimplantation period. The data presented demonstrate that local embryo-oviduct interaction in vitro induces changes in the transcriptional levels of BMP signaling, causing a bidirectional response that reduces the expression levels of this signaling in the oviductal cells while increasing them in the embryo at the early stages. These findings suggest that BMP signaling pathway could be involved in an early cross talk between the bovine embryo and the oviduct during first stages of development. Increased understanding of this intrinsic signaling for maternal-embryo communication is important for future improvements in ART and further understanding of the signaling pathways that potentially contribute to infertility in cattle and humans.

Declaration of interest
The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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References


www.reproduction-online.org
small inhibitor. Characterization of bone morphogenetic protein-
response element in the mouse Smad3 promoter. Journal of Biological
Chemistry 275: 6075–6079. (doi:10.1074/jbc.275.6075)
Janatpour MJ, McMarter MT, Ghenbacev O, Zhou Y, Dong J, Cross JC,
Israel MA & Fisher SJ 2000 lid-2 regulates critical aspects of human
cytotrophoblast differentiation, invasion and migration. Development
127 549–558.
Jones RL, Stoikos C, Findlay JK & Salamonsen LA 2006 TGF-beta
superfamily expression and actions in the endometrium and placentas. 
Kishigami S & Mishina Y 2005 BMP signaling and early embryonic
cytogfr.2005.04.002)
Knight PG & Glister C 2006 TGF-beta superfamily members and ovarian
1.01074)
Kues WA, Sudheer S, Herrmann D, Carnwath JW, Havlicek V,
expression profiling reveals distinct clusters of transcriptional regulation
during bovine preimplantation development in vivo. PNAS 105
9768–9773. (doi:10.1073/pnas.0805160105)
Kusumegi T, Tanaka J, Kawano M, Yonemoto J, Tohyama C & Sone H 2004
BMP7/ActRIIIB regulates estrogen-dependent apoptosis: new biomarkers
for environmental estrogens. Journal of Biochemical and Molecular
Toxicology 18 1–11. (doi:10.1002/jbt.20004)
La Rosa I, Camargo L, Pereira MM, Fernandez-Martín R, Paz DA &
Salamone DF 2011 Effects of bone morphogenetic protein 4 (BMP4) and
inhibin-a on ovine uterine gene expression during bovine preimplantation embryos. 
Reproductive Biology and Endocrinology 9 18. (doi:10.1186/1477-
7827-9-18)
Lawrenson K, Notarioudi M, Lee N, Benjamin E, Jacobs IJ, Jones C &
Gayther SA 2011 In vitro three-dimensional modeling of fallopian tube
secretory epithelial cells. BMC Cell Biology 14 43. (doi:10.1186/1471-
2121-14-43)
(doi:10.1002/hum.20467)
Lee KF, Yao YQ, Kwok KL, Xu JS & Yeung WS 2002 Early developing
embryos affect the gene expression patterns in the mouse oviduct. Biochemical and Biophysical Research Communications 292 564–570.
(doi:10.1006/bbrc.2002.6676)
Lee KB, Folger JK, Rauput SK & Smith GW 2014 Temporal regulation of
mRNAs for select bone morphogenetic proteins (BMP), BMP receptors and their associated SMAD proteins during bovine early embryonic
development: effects of oxogenous BMP2 on embryo developmental progression. Reproductive Biology and Endocrinology 12 67.
(doi:10.1186/1477-7827-12-67)
Li Q 2015 Inhibitory SMADs: potential regulators of ovarian function. 
Livak KJ & Schmittgen TD 2001 Analysis of relative gene expression data
Lonerong P, Khair H, Plumi F, Rieger D, Humblot P & Boland MM 1999
Effect of time interval from insemination to first cleavage on the
expression of BMP genes during bovine early embryonic development. 
Lopera Vásquez R, Hamdi M, Maillo V, López-Martín S, Gutierrez-Adán A,
Roberts RM, Ezashi T & Das P 2004 Trophoblast gene expression:
transcription factors in the specification of early trophoblast. Reproductive Biology and Endocrinology 2 47. (doi:10.1186/1477-
7827-2-47)
Rodriguez A, Tri purani SK, Burton JC, Clementi C, Larina I & Pangas SA
2016 SMAD signaling is required for structural integrity of the female reproductive tract and uterine function during early pregnancy in mice. 1.
Biology of Reproduction 95 44. (doi:10.1093/biolre.116.139477)
Roelen BA, Goumans MJ, van Rooijen MA & Mummery CL 1997
Shimasaki S, Moore RK, Otsuka F & Erickson GF 2004 The bone
Simperitas CA, Fröhlich T, Sathyapalaan T, Arnold C, Ulbrich SE, Leeise HJ &
Reproduction 153 23–33. (doi:10.1530/REP-16-0508)
Smits K, De Coninck DI, Van Nieuwerburgh F, Govaere J, Van Poucke M,
Peelman L, Deforce D & Van Soom A 2016 The equine embryo influences
(doi:10.1016/j.anireprod.2014.06.022)
Smidt-Beekman B, Lodewijckx A, Vervuurt-Vekemans K, De Vocht A,
Sperandeo V, Zebekos A, Perecca C & Mermillod M 2015 Bovine oviduct
ovoid cells differentiates partly in culture, while maintaining their ability to early embryo development rate and quality. Reproduction in Domestic Animals 50 719–729. 
(doi:10.1111/rdaj.12556)
(doi:10.1172/JCI65710)
Salamone DF 2013 In vitro three-dimensional modeling of fallopian tube
Schmittz-Panneau B, Cordova A, Dhorone-Pollet S, Hennequet-Antier C,
Uzbekova S, Martinot E, Doret S, Martin P, Mermillod P & Locatelli Y
2012 Id-2 regulates critical aspects of human preimplantation embryos
a tool for studying oocyte developmental competence. Reproduction
92 137–143. (doi:10.1530/REP-11-01079)
Schmittz-Panneau B, Cordova A, Dhorone-Pollet S, Hennequet-Antier C,
Uzbekova S, Martinot E, Doret S, Martin P, Mermillod P & Locatelli Y
2013 BMP2 regulates early embryonic development and pregnancy
Shimasaki S, Moore RK, Otsuka F & Erickson GF 2004 The bone
Simperitas CA, Fröhlich T, Sathyapalaan T, Arnold C, Ulbrich SE, Leeise HJ &
Reproduction 153 23–33. (doi:10.1530/REP-16-0508)
Smits K, De Coninck DI, Van Nieuwerburgh F, Govaere J, Van Poucke M,
Peelman L, Deforce D & Van Soom A 2016 The equine embryo influences
Soyun EY, LaPol PST, Voon BS, Chen JY-C, Lu JK & Lyons KM 2003
The TIF111 BMP receptor EngR has a role in female reproductive function. 
PNAS 98 7994–7999. (doi:10.1073/pnas.112400798)
Tremlay KD, Dunn NR & Robertson EJ 2001 Mouse embryos lacking
Sma1 signals display defects in extra-embryonic tissues and germ cell
wolf e, arnold gj, bauersachs s, beier hm, blum h, einspanier r, frohlich t, herrler a, hiendleder s, kolle s et al. 2003 embryo-maternal communication in bovine – strategies for deciphering a complex cross-talk. reproduction in domestic animals 38 276–289. (doi:10.1046/j.1439-0531.2003.00435.x)


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