

Androgens in maternal vascular and placental function: implications for preeclampsia pathogenesis

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Abstract

Adequate maternal vascular adaptations and blood supply to the uterus and placenta are crucial for optimal oxygen and nutrient transport to growing fetuses of eutherian mammals, including humans. Multiple factors contribute to hemodynamics and structuring of placental vasculature essential for term pregnancy with minimal complications. In women, failure to achieve or sustain favorable pregnancy progression is, not surprisingly, associated with high incidence of antenatal complications, including preeclampsia, a hypertensive disorder of pregnancy. While the pathogenesis of preeclampsia in women remains unknown, a role for androgens is emerging. The relationship between androgens and maternal cardiovascular and placental function deserves particular consideration because testosterone levels in the circulation of preeclamptic women are elevated approximately two- to three-fold and are positively correlated with vascular dysfunction. Preeclampsia is also associated with elevated placental androgen receptor (AR) gene expression. Studies in animal models mimicking the pattern and level of increase of adult female testosterone levels to those found in preeclamptic pregnancies, replicate key features of preeclampsia, including gestational hypertension, endothelial dysfunction, exaggerated vasoconstriction to angiotensin II, reduced spiral artery remodeling, placental hypoxia, decreased nutrient transport and fetal growth restriction. Taken together, these data strongly implicate AR-mediated testosterone action as an important pathway contributing to clinical manifestation of preeclampsia. This review critically addresses this hypothesis, taking into consideration both clinical and preclinical data.

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Introduction

Pregnancy is characterized by major cardiovascular adaptations, including marked decreases in systemic vascular resistance and mean arterial pressure, along with increases in maternal cardiac output and total blood volume (Magness 1998, Thornburg *et al.* 2000, Chinnathambi *et al.* 2013a). Studies suggest that pregnancy-enhanced vasodilatory actions allow peripheral vessels to accommodate increases in blood flow and volume (Conrad *et al.* 1993). Consistently, maternal vascular adaptations are accompanied by blunted vascular contractility (Naden & Rosenfeld 1981, Magness & Rosenfeld 1986) and enhanced release of endothelium-derived vasodilatory factors (Kawano & Mori 1983, Magness *et al.* 1990, 1996, 2000, Conrad *et al.* 1993, Sladek *et al.* 1997, Williams *et al.* 1997, Gillham *et al.* 2003, Gokina *et al.* 2010). Failure of these vascular adaptations during pregnancy are directly related to several maternal/fetal pathologies, such as increased systemic vascular resistance, hypertension,

proteinuria, poor placental growth, decreased nutrient transport and low birth weight, all characteristics associated with a diagnosis of preeclampsia (Powe *et al.* 2011) outlined in Table 1. Despite being a leading contributor of maternal and perinatal morbidity and death worldwide, the etiology and pathogenesis of preeclampsia remain unclear (Roberts *et al.* 1991, Palei *et al.* 2013, Salam *et al.* 2015).

Treatment options for preeclampsia are limited to management of high blood pressure using antihypertensives, such as methyldopa, hydralazine, labetalol and nifedipine (ACOG 2013), as well as magnesium sulfate for prevention of eclamptic seizures (Al Khaja *et al.* 2014); however, these treatments have limited efficacy, and the only cure is the delivery of the placenta with baby, a totally undesired outcome before late preterm (≥ 34 weeks gestation). While the exact causes of preeclampsia remain unknown, a large body of evidence, supported by preclinical models of preeclampsia, indicates that abnormal placentation early

Table 1 Diagnostic criteria for preeclampsia in women (ACOG Guidelines 2013).

Parameter	Diagnostic criteria
Blood pressure	Greater than or equal to 140 mmHg systolic or greater than or equal to 90 mmHg diastolic on two occasions at least 4 h apart after 20 weeks of gestation in a woman with a previously normal blood pressure OR Greater than or equal to 160 mmHg systolic or greater than or equal to 110 mmHg diastolic, hypertension can be confirmed within a short interval (minutes) to facilitate timely antihypertensive therapy
Proteinuria	Greater than or equal to 300 mg per 24 h urine collection (or this amount extrapolated from a timed collection) OR Protein/creatinine ratio greater than or equal to 0.3* Dipstick reading of 1+ (used only if other quantitative methods not available)
OR in the absence of proteinuria, new-onset hypertension plus new onset of any of the following:	
Thrombocytopenia	Platelet count less than 100,000/microliter
Renal insufficiency	Serum creatinine concentrations greater than 1.1 mg/dL or a doubling of the serum creatinine concentration in the absence of other renal disease
Impaired liver function	Elevated blood concentrations of liver transaminases to twice normal concentration
Pulmonary edema	
Cerebral or visual symptoms	

*Each measured as mg/mL.

in pregnancy is an important initial event in the onset of preeclampsia (Roberts & Redman 1993, Myatt 2002). Such preeclamptic abnormal placentation stimulates the production of anti-angiogenic factors and cytokines, resulting in generalized vascular dysfunction and the clinical manifestation of preeclampsia. In the past several years, dysregulation of steroid hormones, specifically increases in maternal testosterone levels, has emerged as an important endocrinopathy repeatedly associated with clinical manifestations of preeclampsia. In order to develop more effective therapeutic interventions for preeclampsia, it is important to fully understand the role of testosterone in maternal vascular and placental function, as well as blood pressure control, during normal pregnancy and in preeclampsia. This review critically addresses the hypothesis of testosterone-mediated pathogenesis of preeclampsia, taking into consideration data from both clinical (human) and preclinical (animal) studies.

Testosterone levels in clinical preeclampsia

Most studies have investigated the beneficial role of sex steroid hormones, especially estradiol and progesterone, on cardiovascular function during pregnancy in women (Magness 1998). The relationship between testosterone and maternal cardiovascular function, however, is relatively understudied. PubMed search with keywords, 'testosterone, preeclampsia and women', generated 40 publications that were manually screened to identify 14 full-length papers reporting testosterone levels in both preeclampsia and control groups. Twelve of these 14 studies reported elevated plasma levels of testosterone during preeclamptic compared to normotensive (control) pregnancies (Table 2) (Acromite *et al.* 1999, Serin *et al.* 2001, Steier *et al.* 2002, Ficicioglu & Kutlu 2003,

Miller *et al.* 2003, Troisi *et al.* 2003, Atamer *et al.* 2004, Baksu *et al.* 2004, Carlsen *et al.* 2005, Gerulewicz-Vannini *et al.* 2006, Salamalekis *et al.* 2006, Ghorashi & Sheikhatan 2008, Hsu *et al.* 2009, Sharifzadeh *et al.* 2012). These studies report that during late pregnancy, plasma testosterone concentrations range between 100 and 150 ng/dL and these are 1.5- to 2.4-fold higher in preeclamptic compared to normotensive pregnant women (Fig. 1A). The reported mean unbound or 'free' testosterone level circulating in preeclamptic women is also 1.4- to 3.4-fold higher compared to normotensive pregnancies (Fig. 1B). Some studies also indicate that circulating testosterone levels correlate with the severity of preeclampsia, although this is not a universal finding (Ficicioglu & Kutlu 2003, Atamer *et al.* 2004). While there are many androgens, including the relatively bio-ineffective testosterone precursors of dehydroepiandrosterone (DHEA) and androstenedione (A₄), only circulating levels of testosterone are increased during preeclampsia (Table 2). Preeclamptic hyperandrogenic measures include elevated total testosterone, free testosterone, free androgen index (FAI, total testosterone × 100/sex hormone binding globulin) and the testosterone-to-estradiol ratio. Hyperandrogenic pregnant women with polycystic ovary syndrome (PCOS) are at increased risk for preeclampsia (de Vries *et al.* 1998, Kjerulff *et al.* 2011, Kamalanathan *et al.* 2013), and it has been proposed that overproduction of testosterone by the polycystic ovary is the causal factor engaging preeclampsia in PCOS women (Diamant *et al.* 1982, Sir-Petermann *et al.* 2002, Codner & Escobar-Morreale 2007). Obesity, and accompanying insulin resistance-induced compensatory hyperinsulinemia, is predictive of preeclampsia (Seely & Solomon 2003). Insulin stimulates androgen release, including testosterone, from theca cells of normal ovaries

Table 2 Plasma androgen levels in healthy and preeclamptic pregnant women.

Reference/Androgen	Normal vs preeclampsia	P	% Increase in preeclampsia [#]	Gestational age (weeks)
<i>Acromite et al. (1999)</i>				36–38
Total TS	154.5 vs 213.6 ng/dL	<0.01	38	
Free TS	0.3 vs 0.5 ng/dL	<0.05	67	
DHEA-S	175.5 vs 171.0 µg/dL	NS		
Estradiol	33.8 vs 36.4 g/mL	NS		
<i>Salamalekis et al. (2006)</i>				30–31
Total TS	106.3 vs 154.4 ng/dL	<0.05	45	
Free TS	0.21 vs 0.34 ng/dL	<0.05	62	
DHEA-S	76.15 vs 57.62 µg/dL	>0.05		
Androstenedione	110.5 vs 107.1 ng/dL	>0.05		
<i>Ghorashi and Sheikhvatan (2008)</i>				28–39
Free TS	0.58 vs 1.97 ng/dL	0.001	240	
<i>Serin et al. (2001)</i>				34–39
Total TS	24.3 vs 44.1 ng/dL	<0.05	81	
Free TS	0.22 vs 0.44 ng/dL	<0.05	100	
DHEA-S	90.5 vs 162.5 µg/dL	NS		
Androstenedione	210 vs 220 ng/dL	NS		
Estradiol	92.2 vs 73.5 pg/mL	NS		
<i>Carlsen et al. (2005)</i>				33
Total TS	63.4 vs 86.5 ng/dL	0.001	36	
Free TS index	0.61 vs 0.83	0.012	36	
DHEA-S	102.3 vs 121.5 µg/dL	NS		
Androstenedione	280 vs 337 ng/dL	NS		
<i>Baksu et al. (2004)</i>				34
Total TS	136 vs 257 ng/dL	0.001	89	
Free TS index	0.31 vs 0.37	0.01	19	
DHEA-S	109.1 vs 104.3 µg/dL	NS		
Estradiol	5830.1 vs 6164.2 pg/mL	NS		
<i>Steier et al. (2002)</i>				30–38
Total TS	82.1 vs 172.4 ng/dL	<0.01	110	
<i>Hsu et al. (2009)</i>				37
Total TS	34 vs 52 ng/dL	<0.01	53	
<i>Gerulewicz-Vannini et al. (2006)</i>				37
Total TS	103.7 vs 152.2 ng/dL	0.02	47	
Free TS	0.144 vs 0.594 ng/dL	0.002	312	
DHEA-S	70.0 vs 51.1 µg/dL	NS		
<i>Atamer et al. (2004)</i>				34–35
Total TS	29 vs 67 ng/dL	<0.001	131	
DHEA-S	108 vs 112 µg/dL	NS		
Androstenedione	189 vs 158 ng/dL	NS		
Estradiol	2927 vs 3572 pg/mL	NS		
<i>Troisi et al. (2003)</i>				37
Total TS	141.9 vs 214.5 ng/dL	0.0007	51	
Androstenedione	316.0 vs 506.3 ng/dL	0.004	60	
<i>Sharifzadeh et al. (2012)</i>				32–33
Total TS	206 vs 370 ng/dL	<0.01	80	
Free TS	0.074 vs 0.128	<0.01	73	
DHEA-S	51 vs 75 µg/dL	NS		
<i>Miller et al. (2003)</i>				35
Total TS	206 vs 147 ng/dL	NS		
Free TS index	2.03 vs 1.50	NS		
DHEA-S	75 vs 75 µg/dL	NS		
Estradiol	18,536 vs 9619 pg/mL	NS		
<i>Ficicioglu and Kutlu (2003)</i>				34–35
Total TS	218 vs 209 ng/dL	NS		
Free TS index	4.16 vs 5.24	NS		
DHEA-S	104 vs 77 µg/dL	<0.05		
Estradiol	21,880 vs 21,370 pg/mL	NS		

All these studies used immunoassays (ELISA and RIA) to measure TS levels. This raises concern regarding assay sensitivity and the specificity because of risk of cross-reactivity between steroids and their multiple placental metabolites. Recently, liquid chromatography tandem mass spectrometry (LC-MS/MS) has been suggested as the new 'gold standard' method for measurement of TS levels. This recommendation is more geared towards situations in which measurements of TS are below detectable levels (such as in hypogonadal men, women, children etc.) or in species for which no specific antibodies are available (such as sheep). Recent studies that compared the predictive values of TS levels measured by LC-MS/MS and immunoassay showed no significant difference between the two analytical methods (Czeloth *et al.* 2017, Mitchell 2012). The TS levels reported in the studies cited here may be appropriate for two reasons. First, the TS levels in pregnant women are within detectable range, and second, the objective is to detect relative change in preeclamptic group compared to controls.

[#] % increase is calculated as $100 \times (\text{preeclampsia} - \text{normal})/\text{normal}$.

TS, testosterone; NS, not significant.

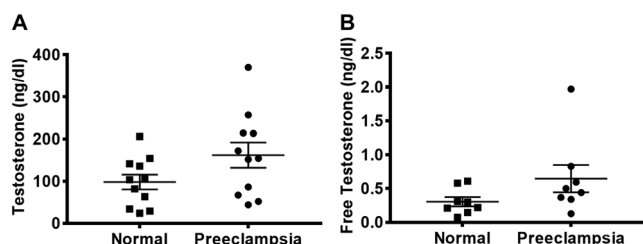


Figure 1 Mean higher total (A) and free testosterone (B) levels reported in preeclamptic patients compared normotensive controls in published studies cited in Table 1. Each point represents a single published study.

(Franks *et al.* 1999), and thus, the exaggerated hyperinsulinemia of obesity during preeclamptic gestation (Kaaja *et al.* 1995, Lorentzen *et al.* 1998) likely contributes to increased maternal testosterone levels (Andersen *et al.* 1995, Pasquali *et al.* 2000, Sutton-Tyrrell *et al.* 2010). Since both obesity, hyperinsulinemia and preeclampsia are more prevalent among hyperandrogenic pregnant women with PCOS than in pregnant women without PCOS (Lonnebotn *et al.* 2018), obesity-enhanced maternal testosterone levels may contribute to PCOS-associated preeclampsia.

Ethnicity has also been implicated in contributing hyperandrogenism-related preeclampsia. Pregnant African-American women exhibit high maternal testosterone levels (120–130%), including elevated fetal cord blood testosterone levels at term (Henderson *et al.* 1988, Potischman *et al.* 2005, Rohrmann *et al.* 2009, Agurs-Collins *et al.* 2012) and are at increased risk for developing preeclampsia (Samadi *et al.* 2001, Iavazzo & Vitoratos 2010). In addition, plasma testosterone

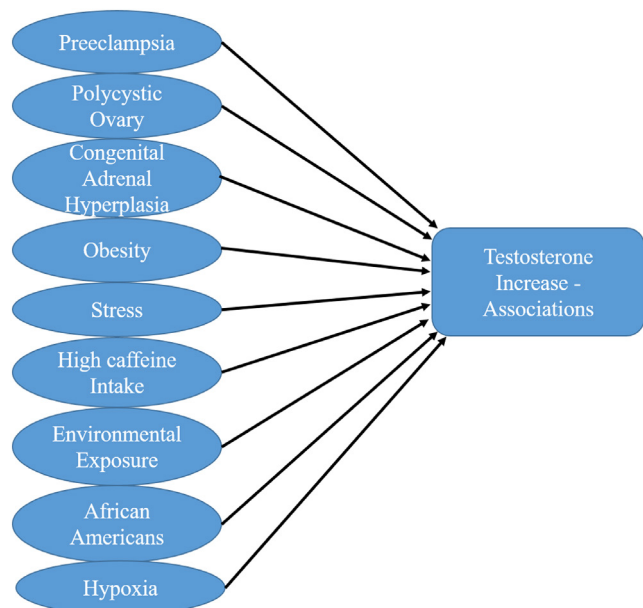


Figure 2 Possible associations for testosterone increase in females and pregnancy.

levels are increased during pregnancy in a variety of situations (Fig. 2), including classical congenital adrenal hyperplasia (Warmann *et al.* 2000, Mains *et al.* 2007), high caffeine intake (Ferrini & Barrett-Connor 1998, Svartberg *et al.* 2003) and stress (Sarkar *et al.* 2007, 2008), all of which are known risk factors for preeclampsia. Furthermore, pregnant women are inadvertently exposed to elevated testosterone levels via environmental pollutants and anabolic steroids (endocrine disruptors). High androgenic activity is reported in water from craft pulp and paper mills, as well as concentrated animal feed operations in the United States and Europe (Parks *et al.* 2001, Orlando *et al.* 2004). Reports have shown that an androgenic growth promoter used in beef cattle, trenbolone, has a half-life of greater than 260 days in animal by-products (Schiffer *et al.* 2001, Hotchkiss & Nelson 2007).

The degree of hyperandrogenism in preeclamptic women varies depending on the sex of their fetus. Preeclamptic women bearing sons exhibit higher testosterone levels than preeclamptic women bearing daughters (Steier *et al.* 2002), contributing to the notion that a male fetus and its placenta contribute additional amounts of testosterone to the maternal circulation (Sathishkumar *et al.* 2012). Such pregnant women bearing male fetuses are at increased risk for developing preeclampsia and placental dysfunction (Stark *et al.* 2006, Murji *et al.* 2012, Sykes *et al.* 2014, Li *et al.* 2018). Furthermore, daughters experiencing a preeclamptic gestation demonstrate higher circulating testosterone levels when they reach puberty (Alsnes *et al.* 2016). Such female offspring are at increased risk of developing hypertension and cardiovascular disease as adults (King *et al.* 2007, Sathishkumar *et al.* 2011c, Chinnathambi *et al.* 2012, 2013b, Vyas *et al.* 2016), and possibly preeclampsia and other pregnancy-related complications. High testosterone levels persist for at least 17 years in women with a documented history of preeclampsia (Laivuori *et al.* 1998). These studies thus provide consistent circumstantial evidence linking increased testosterone levels with preeclampsia.

The origin of the increased testosterone levels during preeclampsia remain uncertain. Studies suggest a placental contribution (Steier *et al.* 2002, Dokras *et al.* 2003). The human placenta, however, lacks the key androgen biosynthetic enzymes, 17 β -hydroxylase and 17,20-desmolase (Christensen 1974). It nevertheless expresses 3 β -hydroxysteroid dehydrogenase type 1 (HSD3B1) (Mason *et al.* 1993), endowing a ready ability to convert DHEA into A₄, as well as the estrogen-preferring 17 β -hydroxysteroid dehydrogenase type 1 (HSD17B1) (Takeyama *et al.* 1998), endowing a weak ability to synthesize testosterone from A₄. After mid-gestation, both maternal and fetal adrenals equally contribute as the major sources of C19 steroids for placental androgen biosynthesis (Turnipseed *et al.* 1976, Kowalczyk *et al.* 1998). The human fetal adrenal cortex includes a fetal

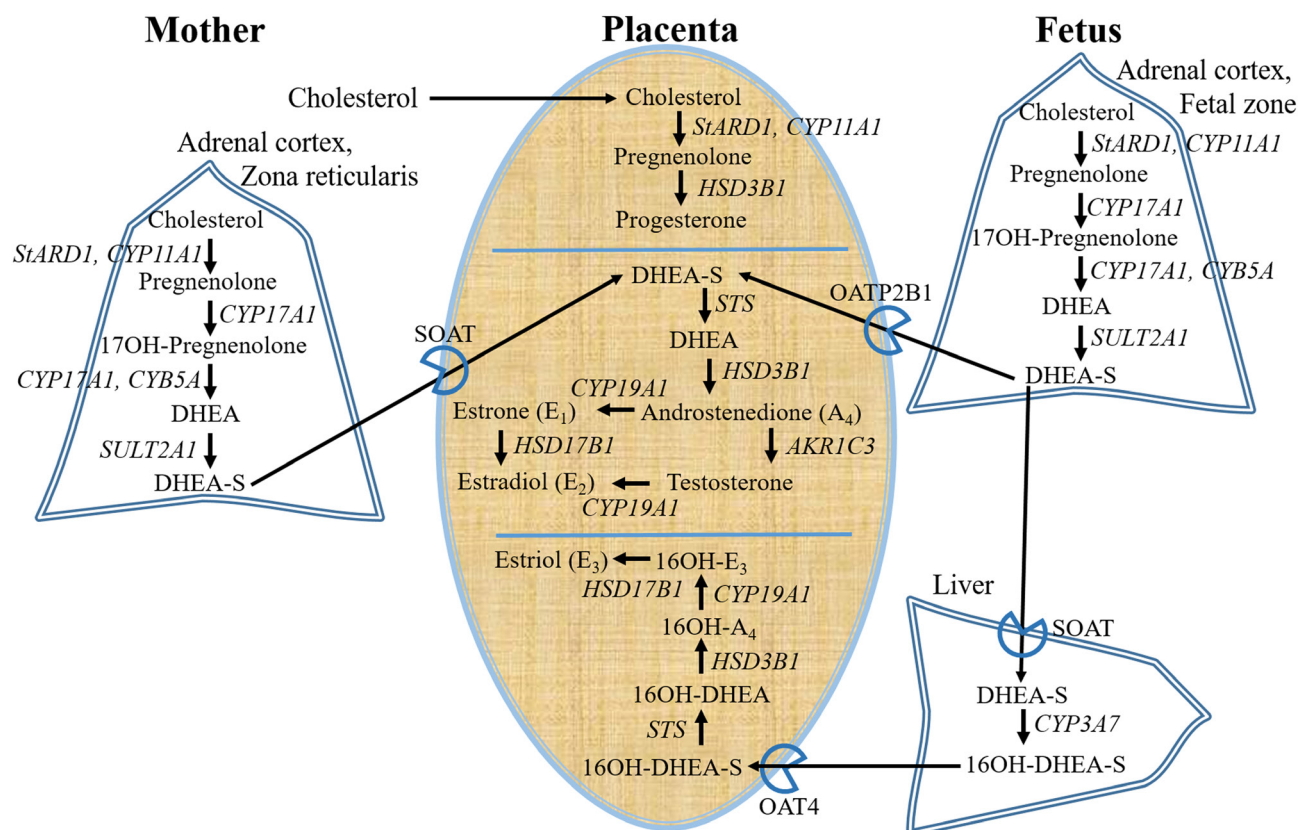


Figure 3 Pathway of biosynthesis and metabolism of testosterone, primary estrogens and progesterone during pregnancy. StARD1, steroidogenic acute regulatory protein; CYP11A1, cholesterol side-chain cleavage enzyme; HSD3B1, 3beta-hydroxysteroid dehydrogenase; CYP17A1, 17 α -hydroxylase/17,20-lyase; CYP5A, cytochrome b5; SULT2A1, sulfotransferase; STS, steroid sulfatase; AKR1C3, aldo-keto reductase type 1C3; HSD17B1, hydroxysteroid 17-beta dehydrogenase 1; CYP19A1, aromatase – cell membrane-located uptake carriers of DHEAS; SOAT, sodium dependent organic anion transporter; OATP2B1 and OAT4, organic anion transporters.

zone, expressing StARD1, CYP11A1, CYP17A1 and SULT2A1, essential for production of DHEA and DHEAS sulfate (DHEA-S), analogous to the maternal zona reticularis, the innermost zone of the maternal adrenal cortex (Fig. 3). After membrane uptake carrier transport into placental syncytiotrophoblast cells, sulfonated testosterone precursors (i.e. DHEA-S) are desulfonated by the enzyme sulfatase (STS) to yield DHEA. Placental steroidogenic enzymes (Fig. 3) then convert DHEA to A₄ (HSD3B1), and A₄ to testosterone (ARK1C3). Accompanying high levels of placental aromatase expression (Mason *et al.* 1993) ensure ready conversion of placental androgens, including testosterone, into non-androgenic, estrogenic metabolites (Gant *et al.* 1971, Buster *et al.* 1979, Dokras *et al.* 2003), including estrone, estradiol and their catechol and methoxy metabolites, some of which display placental bioactivity rivaling that of E₂ (Jobe *et al.* 2011, Landeros *et al.* 2018).

The finding that there are no statistically significant differences regarding circulating maternal levels of DHEA-S and A₄ between control and preeclamptic pregnancies implies that there is no contribution of adrenal steroids to the hyperandrogenism of

preeclampsia. Placental aromatase mRNA and protein expression, however, is decreased in the preeclamptic placenta, diminishing metabolism of A₄ and testosterone into estrogenic metabolites, and tipping the equilibrium between estrogens and androgens in favor of androgens (Sathishkumar *et al.* 2012, Perez-Sepulveda *et al.* 2015). Hepatic conjugation, and thus inactivation, of estrogens also appears diminished during preeclamptic gestations. Maternal circulating levels of unconjugated estrogens, however, remain unchanged from those in normotensive pregnant women (Rosing & Carlstrom 1984). Studies show that testosterone, alone, diminishes aromatase mRNA expression in human trophoblast cells through a miR-22-mediated mechanism (Shao *et al.* 2017). Other factors, tumor necrosis factor alpha (Lau *et al.* 2013) and lipid radicals (Mori *et al.* 2014), which are increased during preeclampsia downregulate aromatase (Milczarek *et al.* 2008, Diaz *et al.* 2009). In addition, hypoxia (which mirrors the actual conditions of the placenta in the context of preeclampsia) also downregulates placental aromatase (Jiang *et al.* 2000, Perez-Sepulveda *et al.* 2015, Yu *et al.* 2015). It would be interesting to assess whether compromised expression

of aromatase also exists in tissues and organs other than placenta during preeclamptic gestation. A recent study also indicated that overexpression of CYP11A1 mRNA (commonly referred as cholesterol side-chain cleavage enzyme that catalyzes the first steps of steroidogenesis) in human trophoblast cells induces increased testosterone production and preeclampsia-like placental dysfunction that could be reversed with flutamide, an androgen receptor antagonist (Pan *et al.* 2017). Taken together, these findings support the notion that increased testosterone during preeclamptic pregnancies may be of placental origin, although other sources cannot be excluded.

Testosterone and maternal blood pressure and uterine artery blood flow

In healthy women experiencing no complications during pregnancy, arterial pressure is stable during the early part of first trimester and then gradually decreases, reaching a nadir during the second trimester (Magness 1998, Bosio *et al.* 1999). Lack of this pregnancy-related decrease in blood pressure indicates a failure in normal cardiovascular adaptation and is considered to be a cardinal feature of preeclampsia (Ishikuro *et al.* 2012). Several independent investigators have demonstrated, through human and animal studies, the association of androgens, especially testosterone, with hypertension (Nakao *et al.* 1981, Reckelhoff *et al.* 1998, Gonzales *et al.* 2004, 2005, Park *et al.* 2004, Chen *et al.* 2007, Yanes *et al.* 2009, Makinen *et al.* 2011). Testosterone levels correlate positively with systolic blood pressure and diastolic blood pressure during and after preeclampsia (Laivuori *et al.* 1998, Serin *et al.* 2001, Carlsen & Heimstad 2012). Experimentally induced increases in maternal testosterone levels during pregnancy in rats, at concentrations that mimic testosterone levels found in human preeclamptic pregnancies, induce increases in systemic arterial pressure (Chinnathambi *et al.* 2013a, 2014b, Fornes *et al.* 2016), implying a causal role for testosterone in raising blood pressure during gestation. The exact mechanism by which testosterone mediates an increase in maternal blood pressure during pregnancy is not clear, but accumulating evidence indicates that testosterone increases vascular reactivity, activating the renin-angiotensin system and altering eicosanoid metabolism, thus favoring an increase in the thromboxane A2 to prostacyclin (PGI2) ratio and causing platelet aggregation in ways that are strikingly similar to those reported in preeclampsia (Acromite *et al.* 1999). Our unpublished observations also show that elevating rat maternal testosterone levels during pregnancy induce renal hypertrophy and proteinuria, a hallmark feature of preeclampsia (Sathishkumar *et al.* 2011a). Treatment with a selective angiotensin type 1 receptor (AT₁R) antagonist, losartan, markedly attenuated the hypertension induced by testosterone

in the pregnant rats (Chinnathambi *et al.* 2014b). These findings suggest that AT₁R activation contributes, at least in part, to the testosterone-induced increase in blood pressure in rat pregnancies.

In addition to adjustments in systemic vasculature, the uteroplacental circulation normally adapts to maintain a low vascular tone to accommodate a more than 20-fold increase in uterine blood flow near-term (Rosenfeld *et al.* 1974, Magness 1998, Osol & Mandala 2009). Studies in hyperandrogenic women with PCOS have shown that their high maternal testosterone levels are associated with increased uterine artery resistance index and reduced blood flow (Palomba *et al.* 2010, 2012). Experimentally induced increase in maternal testosterone levels in pregnant rats show significantly reduced uterine arterial blood flow by 40% (measured using transcutaneous micro ultrasound) (Gopalakrishnan *et al.* 2016). In addition, elevated testosterone decreases uterine arterial diameter and increases resistance and pulsatile index (Gopalakrishnan *et al.* 2016). These findings suggest that the mechanisms controlling blood pressure and uterine artery hemodynamics during pregnancy are perturbed by elevated maternal testosterone levels. Primary estrogens, estrone, estradiol-17 β and estriol play an important role in maintaining uterine blood flow and blunting vascular responses during pregnancy (Albrecht & Pepe 1990, Magness 1998). Jobe *et al.* (2013) elegantly demonstrated that these primary estrogens, and the majority of their catechol and methoxy metabolites, including those with demonstrable placental bioactivity (Jobe *et al.* 2011, Landeros *et al.* 2018), are reduced in preeclampsia (Jobe *et al.* 2013). The lower levels of primary estrogens, together with the reduced expression of placental aromatase could induce precursor steroid hormone accumulation, causing C19 steroids, especially testosterone, to be elevated. It is unclear if elevated testosterone acts independently or if it synergies with reduced downstream C18 estrogens to cause preeclampsia progression. Progesterone levels, however, are reported to be normal (Rosing & Carlstrom 1984, Bussen *et al.* 1998, Hertig *et al.* 2010), decreased (Acikgoz *et al.* 2013, Wan *et al.* 2018) or increased (Tamimi *et al.* 2003, Metz *et al.* 2014) in preeclampsia. Further studies will be needed to clarify whether there is hitherto unrecognized relationship between progesterone and testosterone, and if they work in concert in preeclampsia pathogenesis.

Testosterone-induced mechanisms of vascular dysfunction during pregnancy

Effects on endothelium-dependent relaxation

In humans, normal maternal vascular adaptations are accompanied by enhanced release of three major endothelium-derived vasodilatory factors including nitric oxide (NO) (Conrad *et al.* 1993, Sladek *et al.* 1997,

Williams *et al.* 1997), PGI₂ (Kawano & Mori 1983, Magness *et al.* 1990, 1996, 2000) and endothelium-derived hyperpolarizing factor (EDHF) (Gillham *et al.* 2003, Gokina *et al.* 2010). This is accompanied with concomitant pregnancy-induced increases in mRNA and protein expression of endothelial NO synthase (eNOS) (Sladek *et al.* 1997, Williams *et al.* 1997, Nelson *et al.* 2000, Magness *et al.* 2001), endothelial prostaglandin-I synthase (PGIS) (Bird *et al.* 2000, Magness *et al.* 2000) and EDHF activity (Gokina *et al.* 2010). In the systemic circulation, the principal endothelium-dependent vasodilators are NO and EDHF (Chinnathambi *et al.* 2013a). In the uterine arteries, in addition to NO and EDHF, PGI₂ also plays a role in mediating vascular relaxation (Cooke & Davidge 2003). Elevated testosterone is shown to inhibit acetylcholine-induced relaxation of rat mesenteric and uterine arteries suggesting that elevated testosterone impairs endothelium-dependent relaxation. Specifically, the NO-mediated vasodilation was significantly decreased in mesenteric and uterine arteries in a pregnant rat model of elevated maternal testosterone (Chinnathambi *et al.* 2013a, 2014a). This testosterone-induced decrease in NO-mediated arterial relaxation was found not related to decreased vascular smooth muscle sensitivity to NO, as relaxation of arterial rings to sodium nitroprusside, an exogenous NO donor, was not affected (Chinnathambi *et al.* 2013a, 2014a). These findings indicate that testosterone likely alters synthesis/release of NO. Consistently, studies in rats have shown that testosterone decreases plasma levels of NO_x (marker of NO production) with decreases in eNOS protein expression in uterine arteries (Chinnathambi *et al.* 2014a) and eNOS activity (decreased phosphorylation at excitatory Ser¹¹⁷⁷ site and increased phosphorylation at inhibitory Thr⁴⁹⁵ site) in mesenteric arteries (Chinnathambi *et al.* 2013a). The effect of testosterone in rat uterine arteries appears to be more profound than that in mesenteric arteries as in addition to decreasing NO pathway, it also compromises the EDHF- and PGI₂-mediated relaxation by decreasing expression of small conductance calcium-activated channel-3 and PGI₂ receptor, respectively (Chinnathambi *et al.* 2014a). Taken together, these results suggest that elevated testosterone during pregnancy may specifically impair the NO-mediated relaxation in systemic (mesenteric) vessels, while it compromises all three major vasodilatory pathways in reproductive (uterine) vessels.

Effects on vascular smooth muscle contractile response

Systemic and uterine vasculature are refractory to vasoconstrictions during pregnancy. In contrast, enhanced contractile responses to vasoconstrictors is a characteristic feature of preeclampsia (Naden & Rosenfeld 1981, Magness & Rosenfeld 1986, Benoit *et al.* 2007, Stanhewicz *et al.* 2017). Elevated testosterone during

pregnancy is shown to enhance contractile responses to many vasoconstrictors in endothelium-intact vessels, but in endothelium-denuded vessels, there is enhanced contractile response specific to angiotensin II in rat mesenteric (Chinnathambi *et al.* 2014b) and uterine arteries (Chinnathambi *et al.* 2014a). These enhanced responses observed in rat endothelium-denuded vessels indicate that enhanced arterial sensitivity is primarily because of increased angiotensin II-induced contractions, per se, rather than the loss of the endothelium-mediated relaxation component (Chinnathambi *et al.* 2014a). Since elevated testosterone does not alter the vasomotor response to other potent constrictors, such as K⁺ depolarization, thromboxane agonist U46619 and phenylephrine in endothelium-denuded vessels (Chinnathambi *et al.* 2014a,b), it appears that testosterone has a selective effect in enhancing vascular smooth muscle response to angiotensin II. It is possible that testosterone-mediated vascular smooth muscle dysfunction occurs at the agonist-specific receptor level rather than at common intracellular signaling pathways. Consistently, studies show that gestational elevation in testosterone levels causes selective upregulation of vasoconstrictive AT₁ receptor and downregulation of vasodilatory AT₂ receptor in mesenteric and uterine arteries implying that increased AT₁/AT₂ receptor ratio may play an underlying role in testosterone-induced exaggerated vasoconstriction to angiotensin II (Chinnathambi *et al.* 2014a,b).

Testosterone on placental development and function

The progenitor cytotrophoblast cell is the stem cell of the placenta. These cells proliferate throughout gestation, differentiating along two pathways to form either villous cytotrophoblast, which ultimately can become syncytiotrophoblasts (outer cellular layer) or extravillous cytotrophoblasts (inner cellular layer). Syncytiotrophoblast is a specialized epithelium that has several functions, including transport of gases, nutrients and waste products and synthesis of peptide and steroid hormones that regulate placental, fetal and maternal systems. Extravillous trophoblasts have a proliferative component and an invasive component. There is also a migratory extravillous trophoblast, which is neither invasive nor proliferative. AR is present in syncytiotrophoblasts and in the decidua during the first trimester of human gestation (Horie *et al.* 1992). The expression of AR in human preeclamptic placentae is considerably higher than its expression in healthy placentae from uncomplicated pregnancies (Hsu *et al.* 2009, Sathishkumar *et al.* 2012). Also, genetic polymorphisms in the AR gene are associated with increased risk of preeclampsia (Lim *et al.* 2011). Rat models show that experimentally elevated maternal testosterone levels during pregnancy induce a reduction in placental size and weight (Sathishkumar *et al.* 2011b, Sun *et al.*

2012). The reason for smaller placenta in testosterone-exposed dams is not known, but may involve increased apoptosis or decreased proliferation (Ling *et al.* 2002). Pan *et al.* (2017) revealed a critical role for testosterone in human trophoblast invasion and demonstrated that flutamide, an AR antagonist, could rescue testosterone-induced reduction in invasion (Pan *et al.* 2017). It is possible that testosterone-induced autophagy (human) (Pan *et al.* 2017), reduced invasion (human) (Pan *et al.* 2017) or advanced placental differentiation (sheep) (Veiga-Lopez *et al.* 2011), may all contribute to such alterations in placental weight/morphology.

Vasculogenesis and angiogenesis are critical processes that lead to the formation of the placental vascular network necessary for optimal uteroplacental circulation (Huppertz & Peeters 2005, Arroyo & Winn 2008). Testosterone, however, downregulates the expression of genes related to vascular development and angiogenesis (*Ccr3*, *Stra6*, *Dhcr7*, *Arid1a*, *Ptprj*, *Col1a2*, *Lef1*, *Col1a1* and *Mmp2*) in the rat placenta (Gopalakrishnan *et al.* 2016). Along with this antivasculogenetic gene expression profile, testosterone also decreases the radial and spiral artery diameters and inhibits branching angiogenesis (Gopalakrishnan *et al.* 2016). One of the important functions of the placenta is to promote nutrient transport to the fetus. Elevated testosterone is shown to decrease placental amino acid transport to rat fetuses (Sathishkumar *et al.* 2011b). This reduction in amino acid transport is related to reduced expression of the system A amino acid transporters (*slc38a2*/*Snat 2*) in the rat placenta (Sathishkumar *et al.* 2011b). Testosterone also decreases placental oxygenation with associated increase in hypoxia-inducible factor 1 α and hypoxia responsive genes, presumable due to compromised placental vascularization (Gopalakrishnan *et al.* 2016). In addition to placental compromise, the fetuses of testosterone -exposed pregnant rats also receive less oxygen and are hypoxic (Gopalakrishnan *et al.* 2016). Elevated testosterone, however, does not alter glucose transport across the rat placenta (Sathishkumar *et al.*

2011b). Thus, testosterone increases during pregnancy alter placental structure and function leading to decreases in amino acid and oxygen availability to the fetus.

Testosterone effects on fetal growth

Studies have shown that elevated maternal testosterone levels are associated with reduced birth weights in certain human populations (Sir-Petermann *et al.* 2005, Carlsen *et al.* 2006, Mehrabian & Kelishadi 2012), rats (Sun *et al.* 2012, Fornes *et al.* 2016), sheep (Manikkam *et al.* 2004, Steckler *et al.* 2005, Recabarren *et al.* 2008, Beckett *et al.* 2014) and marmoset monkeys (Smith *et al.* 2010), but not in rhesus monkeys (Abbott *et al.* 2010) and not in human populations of non-Spanish descent (Abbott *et al.* 2016). In rhesus monkeys, testosterone was experimentally increased during early-to-mid-gestation, 2 months prior to parturition, hence, it is possible that initial fetal growth restriction is masked by subsequent *in utero* catchup growth. Female infant monkeys, rats and sheep exposed to such gestational testosterone excess, however, exhibit accelerated body weight gain 2 months following parturition (Manikkam *et al.* 2004, Abbott *et al.* 2010, Sathishkumar *et al.* 2011c) and demonstrate increased abdominal adiposity and onset of type 2 diabetes and hypertension in adulthood (Chinnathambi *et al.* 2012, 2013b, Abbott *et al.* 2016). Testosterone is a lipophilic hormone and can diffuse across tissues, including placenta (Dell'Acqua *et al.* 1966, Meulenberg & Hofman 1991, Wang *et al.* 2005); however, whether fetal growth restriction induced by testosterone is the result of a direct effect on the fetus, or is secondary to decreased uterine blood flow or compromised placental function, remains to be resolved.

Conclusions

Several studies show that circulating levels of testosterone are two- to three-fold higher in preeclamptic pregnancies

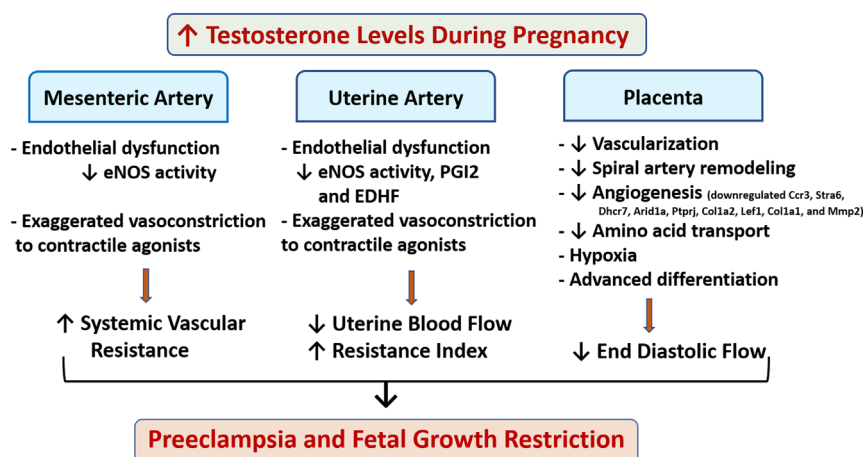


Figure 4 Unifying model depicting the central role of testosterone in preeclampsia. Increased testosterone level causes systemic, uterine and placental vascular dysfunction leading to increased blood pressure, decreased uterine artery blood flow and placental insufficiency, which may contribute to fetal growth restriction.

compared to those of healthy women experiencing uncomplicated pregnancies. Elevated testosterone in pregnant rats results in significantly increased arterial pressure and decreased uterine arterial hemodynamics. Testosterone, in pregnant rats, also causes endothelial dysfunction and exaggerated vasoconstriction to contractile agonists and dysregulates renin-angiotensin system with exaggerated vascular smooth muscle sensitivity to angiotensin II. In addition, testosterone compromises rat placenta vascularization and nutrient transport leading to placental hypoxia and fetal growth restriction. It is therefore possible that some of the vascular and placental effects observed during preeclampsia may indeed be testosterone mediated (Fig. 4). Therefore, strategies that (1) diminish excessive testosterone action in the cardiovascular and placental system and (2) identify the cause(s) of testosterone elevations during pregnancy, could have important therapeutic potential in treatment of pregnancies complicated by vascular dysfunction and fetal growth restriction.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of this review.

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References

- Abbott DH, Bruns CR, Barnett DK, Dunaif A, Goodfriend TL, Dumesic DA & Tarantal AF 2010 Experimentally induced gestational androgen excess disrupts glucoregulation in rhesus monkey dams and their female offspring. *American Journal of Physiology: Endocrinology and Metabolism* **299** E741–E751. (<https://doi.org/10.1152/ajpendo.00058.2010>)
- Abbott DH, Levine JE & Dumesic DA 2016 Translational insight into polycystic ovary syndrome (PCOS) from female monkeys with PCOS-like traits. *Current Pharmaceutical Design* **22** 5625–5633. (<https://doi.org/10.2174/1381612822666160715133437>)
- Acikgoz S, Bayar UO, Can M, Guven B, Mungan G, Dogan S & Sumbuloglu V 2013 Levels of oxidized LDL, estrogens, and progesterone in placenta tissues and serum paraoxonase activity in preeclampsia. *Mediators of Inflammation* **2013** 862982. (<https://doi.org/10.1155/2013/862982>)
- ACOG, Task Force on Hypertension in Pregnancy 2013 Hypertension in pregnancy. Report of the American College of Obstetricians and Gynecologists' Task Force on Hypertension in Pregnancy. *Obstetrics and Gynecology* **122** 1122–1131. (<https://doi.org/10.1097/01.AOG.0000437382.03963.88>)
- Acromite MT, Mantzoros CS, Leach RE, Hurwitz J & Dorey LG 1999 Androgens in preeclampsia. *American Journal of Obstetrics and Gynecology* **180** 60–63. ([https://doi.org/10.1016/S0002-9378\(99\)70150-X](https://doi.org/10.1016/S0002-9378(99)70150-X))
- Agurs-Collins T, Rohrmann S, Sutcliffe C, Bienstock JL, Monseque D, Akereyeni F, Bradwin G, Rifai N, Pollak MN & Platz EA 2012 Racial variation in umbilical cord blood sex steroid hormones and the insulin-like growth factor axis in African-American and white female neonates. *Cancer Causes and Control* **23** 445–454. (<https://doi.org/10.1007/s10552-011-9893-6>)
- Al Khaja KA, Sequeira RP, Alkhaja AK & Damanhori AH 2014 Drug treatment of hypertension in pregnancy: a critical review of adult guideline recommendations. *Journal of Hypertension* **32** 454–463. (<https://doi.org/10.1097/HJH.000000000000069>)
- Albrecht ED & Pepe GJ 1990 Placental steroid hormone biosynthesis in primate pregnancy. *Endocrine Reviews* **11** 124–150. (<https://doi.org/10.1210/edrv-11-1-124>)
- Alsnes IV, Janszky I, Asvold BO, Okland I, Forman MR & Vatten LJ 2016 Maternal preeclampsia and androgens in the offspring around puberty: a follow-up study. *PLoS ONE* **11** e0167714. (<https://doi.org/10.1371/journal.pone.0167714>)
- Andersen P, Seljeflot I, Abdelnoor M, Arnesen H, Dale PO, Lovik A & Birkeland K 1995 Increased insulin sensitivity and fibrinolytic capacity after dietary intervention in obese women with polycystic ovary syndrome. *Metabolism: Clinical and Experimental* **44** 611–616. ([https://doi.org/10.1016/0026-0495\(95\)90118-3](https://doi.org/10.1016/0026-0495(95)90118-3))
- Arroyo JA & Winn VD 2008 Vasculogenesis and angiogenesis in the IUGR placenta. *Seminars in Perinatology* **32** 172–177. (<https://doi.org/10.1053/j.semperi.2008.02.006>)
- Atamer Y, Erden AC, Demir B, Kocyigit Y & Atamer A 2004 The relationship between plasma levels of leptin and androgen in healthy and preeclamptic pregnant women. *Acta Obstetrica et Gynecologica Scandinavica* **83** 425–430. (<https://doi.org/10.1111/j.0001-6349.2004.00276.x>)
- Baksu A, Gurarslan H & Goker N 2004 Androgen levels in pre-eclamptic pregnant women. *International Journal of Gynecology and Obstetrics* **84** 247–248. ([https://doi.org/10.1016/S0020-7292\(03\)00318-7](https://doi.org/10.1016/S0020-7292(03)00318-7))
- Beckett EM, Astapova O, Steckler TL, Veiga-Lopez A & Padmanabhan V 2014 Developmental programming: impact of testosterone on placental differentiation. *Reproduction* **148** 199–209. (<https://doi.org/10.1530/REP-14-0055>)
- Benoit C, Zavec J & Wang Y 2007 Vasoreactivity of chorionic plate arteries in response to vasoconstrictors produced by preeclamptic placentas. *Placenta* **28** 498–504. (<https://doi.org/10.1016/j.placenta.2006.09.002>)
- Bird IM, Sullivan JA, Di T, Cale JM, Zhang L, Zheng J & Magness RR 2000 Pregnancy-dependent changes in cell signaling underlie changes in differential control of vasodilator production in uterine artery endothelial cells. *Endocrinology* **141** 1107–1117. (<https://doi.org/10.1210/endo.141.3.7367>)
- Bosio PM, McKenna PJ, Conroy R & O'Herlihy C 1999 Maternal central hemodynamics in hypertensive disorders of pregnancy. *Obstetrics and Gynecology* **94** 978–984. ([https://doi.org/10.1016/S0029-7844\(99\)00430-5](https://doi.org/10.1016/S0029-7844(99)00430-5))
- Bussen SS, Sutterlin MW & Steck T 1998 Plasma renin activity and aldosterone serum concentration are decreased in severe preeclampsia but not in the HELLP-syndrome. *Acta Obstetrica et Gynecologica Scandinavica* **77** 609–613. (<https://doi.org/10.1034/j.1600-0412.1998.770606.x>)
- Buster JE, Chang RJ, Preston DL, Elashoff RM, Cousins LM, Abraham GE, Hobel CJ & Marshall JR 1979 Interrelationships of circulating maternal steroid concentrations in third trimester pregnancies. II. C18 and C19 steroids: estradiol, estriol, dehydroepiandrosterone, dehydroepiandrosterone sulfate, delta 5-androstenediol, delta 4-androstenedione, testosterone, and dihydrotestosterone. *Journal of Clinical Endocrinology and Metabolism* **48** 139–142. (<https://doi.org/10.1210/jcem-48-1-139>)
- Carlsen SM & Heimstad R 2012 Androgen levels are associated with blood pressure in pregnant women after term. *Acta Obstetrica et Gynecologica Scandinavica* **91** 232–236. (<https://doi.org/10.1111/j.1600-0412.2011.01280.x>)
- Carlsen SM, Romundstad P & Jacobsen G 2005 Early second-trimester maternal hyperandrogenemia and subsequent preeclampsia: a prospective study. *Acta Obstetrica et Gynecologica Scandinavica* **84** 117–121. (<https://doi.org/10.1111/j.0001-6349.2005.00493.x>)
- Carlsen SM, Jacobsen G & Romundstad P 2006 Maternal testosterone levels during pregnancy are associated with offspring size at birth. *European Journal of Endocrinology* **155** 365–370. (<https://doi.org/10.1530/eje.1.02200>)
- Chen MJ, Yang WS, Yang JH, Chen CL, Ho HN & Yang YS 2007 Relationship between androgen levels and blood pressure in young women with polycystic ovary syndrome. *Hypertension* **49** 1442–1447. (<https://doi.org/10.1161/HYPERTENSIONAHA.106.083972>)

- Chinnathambi V, Balakrishnan M, Yallampalli C & Sathishkumar K 2012 Prenatal testosterone exposure leads to hypertension that is gonadal hormone-dependent in adult rat male and female offspring. *Biology of Reproduction* **86** 131–137. (<https://doi.org/10.1095/biolreprod.111.097550>)
- Chinnathambi V, Balakrishnan M, Ramadoss J, Yallampalli C & Sathishkumar K 2013a Testosterone alters maternal vascular adaptations: role of the endothelial NO system. *Hypertension* **61** 647–654. (<https://doi.org/10.1161/HYPERTENSIONAHA.111.00486>)
- Chinnathambi V, Yallampalli C & Sathishkumar K 2013b Prenatal testosterone induces sex-specific dysfunction in endothelium-dependent relaxation pathways in adult male and female rats. *Biology of Reproduction* **89** 97. (<https://doi.org/10.1095/biolreprod.113.111542>)
- Chinnathambi V, Blesson CS, Vincent KL, Saade GR, Hankins GD, Yallampalli C & Sathishkumar K 2014a Elevated testosterone levels during rat pregnancy cause hypersensitivity to angiotensin II and attenuation of endothelium-dependent vasodilation in uterine arteries. *Hypertension* **64** 405–414. (<https://doi.org/10.1161/HYPERTENSIONAHA.114.03283>)
- Chinnathambi V, More AS, Hankins GD, Yallampalli C & Sathishkumar K 2014b Gestational exposure to elevated testosterone levels induces hypertension via heightened vascular angiotensin II type 1 receptor signaling in rats. *Biology of Reproduction* **91** 6. (<https://doi.org/10.1095/biolreprod.114.118968>)
- Christensen A 1974 Hormone and enzyme assays in pregnancy. I. Studies on the placental and the tissue cystine-aminopeptidase activity in peripheral plasma from non-pregnant and pregnant women, and in plasma from the umbilical cord. *Acta Endocrinologica* **76** 189–200.
- Codner E & Escobar-Morreale HF 2007 Clinical review: hyperandrogenism and polycystic ovary syndrome in women with type 1 diabetes mellitus. *Journal of Clinical Endocrinology and Metabolism* **92** 1209–1216. (<https://doi.org/10.1210/jc.2006-2641>)
- Conrad KP, Joffe GM, Kruszyna H, Kruszyna R, Rochelle LG, Smith RP, Chavez JE & Mosher MD 1993 Identification of increased nitric oxide biosynthesis during pregnancy in rats. *FASEB Journal* **7** 566–571. (<https://doi.org/10.1096/fasebj.7.6.7682524>)
- Cooke CL & Davidge ST 2003 Pregnancy-induced alterations of vascular function in mouse mesenteric and uterine arteries. *Biology of Reproduction* **68** 1072–1077. (<https://doi.org/10.1095/biolreprod.102.009886>)
- Czeloth K, Bobjer J, Bronton K, Christofferson O, Frederiksen H, Holmboe S, Anderson A, Juul A & Giwercman A 2017 Comparison of testosterone measurement by the new 'gold standard' method liquid chromatography tandem mass spectrometry versus the traditionally in clinical routine used immunoassays in the prediction of symptoms of male hypogonadism. *European Urology Supplements* **16** e3018.
- de Vries MJ, Dekker GA & Schoemaker J 1998 Higher risk of preeclampsia in the polycystic ovary syndrome. A case control study. *European Journal of Obstetrics, Gynecology and Reproductive Biology* **76** 91–95. ([https://doi.org/10.1016/S0301-2115\(97\)00164-4](https://doi.org/10.1016/S0301-2115(97)00164-4))
- Dell'Acqua S, Mancuso S, Eriksson G & Diczfalussy E 1966 Metabolism of retrotestosterone and testosterone by midterm human placentas perfused in situ. *Biochimica et Biophysica Acta/General Subjects* **130** 241–248. ([https://doi.org/10.1016/0304-4165\(66\)90028-6](https://doi.org/10.1016/0304-4165(66)90028-6))
- Diamant YZ, Rimón E & Evron S 1982 High incidence of preeclamptic toxemia in patients with polycystic ovarian disease. *European Journal of Obstetrics, Gynecology and Reproductive Biology* **14** 199–204. ([https://doi.org/10.1016/0028-2243\(82\)90097-1](https://doi.org/10.1016/0028-2243(82)90097-1))
- Diaz L, Noyola-Martinez N, Barrera D, Hernandez G, Avila E, Halhali A & Larrea F 2009 Calcitriol inhibits TNF-alpha-induced inflammatory cytokines in human trophoblasts. *Journal of Reproductive Immunology* **81** 17–24. (<https://doi.org/10.1016/j.jri.2009.02.005>)
- Dokras A, Spaczynski RZ, Behrman HR & Duleba AJ 2003 Testosterone levels in pregnant women correlate with the insulin response during the glucose tolerance test. *Fertility and Sterility* **79** 492–497. ([https://doi.org/10.1016/S0015-0282\(02\)04764-7](https://doi.org/10.1016/S0015-0282(02)04764-7))
- Ferrini RL & Barrett-Connor E 1998 Sex hormones and age: a cross-sectional study of testosterone and estradiol and their bioavailable fractions in community-dwelling men. *American Journal of Epidemiology* **147** 750–754. (<https://doi.org/10.1093/oxfordjournals.aje.a009519>)
- Ficioglu C & Kutlu T 2003 The role of androgens in the aetiology and pathology of pre-eclampsia. *Journal of Obstetrics & Gynaecology* **23** 134–137.
- Fornes R, Hu M, Maliqueo M, Kokosar M, Benrick A, Carr D, Billig H, Jansson T, Manni L & Stener-Victorin E 2016 Maternal testosterone and placental function: effect of electroacupuncture on placental expression of angiogenic markers and fetal growth. *Molecular and Cellular Endocrinology* **433** 1–11. (<https://doi.org/10.1016/j.mce.2016.05.014>)
- Franks S, Gilling-Smith C, Watson H & Willis D 1999 Insulin action in the normal and polycystic ovary. *Endocrinology and Metabolism Clinics of North America* **28** 361–378. ([https://doi.org/10.1016/S0889-8529\(05\)70074-8](https://doi.org/10.1016/S0889-8529(05)70074-8))
- Gant NF, Hutchinson HT, Siiteri PK & MacDonald PC 1971 Study of the metabolic clearance rate of dehydroisoandrosterone sulfate in pregnancy. *American Journal of Obstetrics and Gynecology* **111** 555–563. ([https://doi.org/10.1016/0002-9378\(71\)90472-8](https://doi.org/10.1016/0002-9378(71)90472-8))
- Gerulewicz-Vannini D, Camero Y, Salas J & Hernandez-Andrade E 2006 High plasmatic androgen levels in women affected with pregnancy-induced hypertension. *Revista de Investigacion Clinica* **58** 228–233.
- Ghorashi V & Sheikhatvan M 2008 The relationship between serum concentration of free testosterone and pre-eclampsia. *Endokrynologia Polska* **59** 390–392.
- Gillham JC, Kenny LC & Baker PN 2003 An overview of endothelium-derived hyperpolarising factor (EDHF) in normal and compromised pregnancies. *European Journal of Obstetrics, Gynecology and Reproductive Biology* **109** 2–7. ([https://doi.org/10.1016/S0301-2115\(03\)00044-7](https://doi.org/10.1016/S0301-2115(03)00044-7))
- Gokina NI, Kuzina OY & Vance AM 2010 Augmented EDHF signaling in rat uteroplacental vasculature during late pregnancy. *American Journal of Physiology: Heart and Circulatory Physiology* **299** H1642–H1652. (<https://doi.org/10.1152/ajpheart.00227.2010>)
- Gonzales RJ, Krause DN & Duckles SP 2004 Testosterone suppresses endothelium-dependent dilation of rat middle cerebral arteries. *American Journal of Physiology: Heart and Circulatory Physiology* **286** H552–560. (<https://doi.org/10.1152/ajpheart.00663.2003>)
- Gonzales RJ, Ghaffari AA, Duckles SP & Krause DN 2005 Testosterone treatment increases thromboxane function in rat cerebral arteries. *American Journal of Physiology: Heart and Circulatory Physiology* **289** H578–585. (<https://doi.org/10.1152/ajpheart.00958.2004>)
- Gopalakrishnan K, Mishra JS, Chinnathambi V, Vincent KL, Patrikeev I, Motamedi M, Saade GR, Hankins GD & Sathishkumar K 2016 Elevated testosterone reduces uterine blood flow, spiral artery elongation, and placental oxygenation in pregnant rats. *Hypertension* **67** 630–639. (<https://doi.org/10.1161/HYPERTENSIONAHA.115.06946>)
- Henderson BE, Bernstein L, Ross RK, Depue RH & Judd HL 1988 The early in utero oestrogen and testosterone environment of blacks and whites: potential effects on male offspring. *British Journal of Cancer* **57** 216–218. (<https://doi.org/10.1038/bjc.1988.46>)
- Hertig A, Lier P, Chabbert-Buffet N, Fort J, Pianos A, Eychenne B, Cambourg A, Schumacher M, Berkane N, Lefevre G et al. 2010 Steroid profiling in preeclamptic women: evidence for aromatase deficiency. *American Journal of Obstetrics and Gynecology* **203** 477.e471–477. (<https://doi.org/10.1016/j.ajog.2010.06.011>)
- Horie K, Takakura K, Imai K, Liao S & Mori T 1992 Immunohistochemical localization of androgen receptor in the human endometrium, decidua, placenta and pathological conditions of the endometrium. *Human Reproduction* **7** 1461–1466. (<https://doi.org/10.1093/oxfordjournals.humrep.a137595>)
- Hotchkiss AK & Nelson RJ 2007 An environmental androgen, 17beta-trenbolone, affects delayed-type hypersensitivity and reproductive tissues in male mice. *Journal of Toxicology and Environmental Health: Part A* **70** 138–140. (<https://doi.org/10.1080/15287390600575091>)
- Hsu TY, Lan KC, Tsai CC, Ou CY, Cheng BH, Tsai MY, Kang HY, Tung YH, Wong YH & Huang KE 2009 Expression of androgen receptor in human placentas from normal and preeclamptic pregnancies. *Taiwanese Journal of Obstetrics and Gynecology* **48** 262–267. ([https://doi.org/10.1016/S1028-4559\(09\)60301-6](https://doi.org/10.1016/S1028-4559(09)60301-6))
- Huppertz B & Peeters LL 2005 Vascular biology in implantation and placentation. *Angiogenesis* **8** 157–167. (<https://doi.org/10.1007/s10456-005-9007-8>)
- Iavazzo C & Vitoratos N 2010 Polycystic ovarian syndrome and pregnancy outcome. *Archives of Gynecology and Obstetrics* **282** 235–239. (<https://doi.org/10.1007/s00404-010-1495-0>)
- Ishikuro M, Obara T, Metoki H, Ohkubo T, Yaegashi N, Kuriyama S & Imai Y 2012 Blood pressure changes during pregnancy. *Hypertension Research*. (<https://doi.org/10.1038/hr.2012.33>)

- Jiang B, Kamat A & Mendelson CR 2000 Hypoxia prevents induction of aromatase expression in human trophoblast cells in culture: potential inhibitory role of the hypoxia-inducible transcription factor Mash-2 (mammalian achaete-scute homologous protein-2). *Molecular Endocrinology* **14** 1661–1673. (<https://doi.org/10.1210/mend.14.10.0539>)
- Jobe SO, Fling SN, Ramadoss J & Magness RR 2011 A novel role for an endothelial adrenergic receptor system in mediating catecholestradiol-induced proliferation of uterine artery endothelial cells. *Hypertension* **58** 874–881. (<https://doi.org/10.1161/HYPERTENSIONAHA.111.178046>)
- Jobe SO, Tyler CT & Magness RR 2013 Aberrant synthesis, metabolism, and plasma accumulation of circulating estrogens and estrogen metabolites in preeclampsia implications for vascular dysfunction. *Hypertension* **61** 480–487. (<https://doi.org/10.1161/HYPERTENSIONAHA.111.201624>)
- Kaaja R, Tikkanen MJ, Viinikka L & Ylikorkala O 1995 Serum lipoproteins, insulin, and urinary prostanoid metabolites in normal and hypertensive pregnant women. *Obstetrics and Gynecology* **85** 353–356. ([https://doi.org/10.1016/0029-7844\(94\)00380-V](https://doi.org/10.1016/0029-7844(94)00380-V))
- Kamalanathan S, Sahoo JP & Sathyapalan T 2013 Pregnancy in polycystic ovary syndrome. *Indian Journal of Endocrinology and Metabolism* **17** 37–43. (<https://doi.org/10.4103/2230-8210.107830>)
- Kawano M & Mori N 1983 Prostacyclin producing activity of human umbilical, placental and uterine vessels. *Prostaglandins* **26** 645–662. ([https://doi.org/10.1016/0090-6980\(83\)90201-0](https://doi.org/10.1016/0090-6980(83)90201-0))
- King AJ, Olivier NB, Mohankumar PS, Lee JS, Padmanabhan V & Fink GD 2007 Hypertension caused by prenatal testosterone excess in female sheep. *American Journal of Physiology: Endocrinology and Metabolism* **292** E1837–E1841. (<https://doi.org/10.1152/ajpendo.00668.2006>)
- Kjerluff LE, Sanchez-Ramos L & Duffy D 2011 Pregnancy outcomes in women with polycystic ovary syndrome: a metaanalysis. *American Journal of Obstetrics and Gynecology* **204** 558.e551–558.e556. (<https://doi.org/10.1016/j.ajog.2011.03.021>)
- Kowalczyk TD, Cabaniss ML & Cusmano L 1998 Association of low unconjugated estriol in the second trimester and adverse pregnancy outcome. *Obstetrics and Gynecology* **91** 396–400. ([https://doi.org/10.1016/S0029-7844\(97\)00677-7](https://doi.org/10.1016/S0029-7844(97)00677-7))
- Laivuori H, Kaaja R, Rutanen EM, Viinikka L & Ylikorkala O 1998 Evidence of high circulating testosterone in women with prior preeclampsia. *Journal of Clinical Endocrinology and Metabolism* **83** 344–347. (<https://doi.org/10.1210/jcem.83.2.4543>)
- Landeros RV, Pastore MB & Magness RR 2018 Effects of the catechol and methoxy metabolites of 17beta-estradiol on nitric oxide production by ovine uterine artery endothelial cells. *Reproductive Sciences* Epub. (<https://doi.org/10.1177/1933719118783265>)
- Lau SY, Guild SJ, Barrett CJ, Chen Q, McCowan L, Jordan V & Chamley LW 2013 Tumor necrosis factor- α , interleukin-6, and interleukin-10 levels are altered in preeclampsia: a systematic review and meta-analysis. *American Journal of Reproductive Immunology* **70** 412–427. (<https://doi.org/10.1111/aji.12138>)
- Li X, Zhang W, Lin J, Liu H, Yang Z, Teng Y, Duan S, Lin X, Xie Y, Li Y *et al.* 2018 Risk factors for adverse maternal and perinatal outcomes in women with preeclampsia: analysis of 1396 cases. *Journal of Clinical Hypertension* **20** 1049–1057. (<https://doi.org/10.1111/jch.13302>)
- Lim JH, Kim S, Lee SW, Park SY, Han JY, Chung JH, Kim MY, Yang JH & Ryu HM 2011 Association between genetic polymorphisms in androgen receptor gene and the risk of preeclampsia in Korean women. *Journal of Assisted Reproduction and Genetics* **28** 85–90. (<https://doi.org/10.1007/s10815-010-9485-5>)
- Ling S, Dai A, Williams MR, Myles K, Dilley RJ, Komesaroff PA & Sudhir K 2002 Testosterone (T) enhances apoptosis-related damage in human vascular endothelial cells. *Endocrinology* **143** 1119–1125. (<https://doi.org/10.1210/endo.143.3.8679>)
- Lonnbotn M, Natvig GK, Benediktssdottir B, Burgess JA, Holm M, Jogi R, Lindberg E, Macsali F, Schlunssen V, Skulstad SM *et al.* 2018 Polycystic ovary syndrome, body mass index and hypertensive disorders in pregnancy. *Pregnancy Hypertension* **11** 32–37. (<https://doi.org/10.1016/j.preghy.2017.12.006>)
- Lorentzen B, Birkeland KI, Endresen MJ & Henriksen T 1998 Glucose intolerance in women with preeclampsia. *Acta Obstetrica et Gynecologica Scandinavica* **77** 22–27. (<https://doi.org/10.1080/00016349808565805>)
- Magness RR 1998 Maternal cardiovascular and other physiologic responses to the endocrinology of pregnancy. In *The Endocrinology of Pregnancy* (Contemporary Endocrinology), pp 507–540. Ed FW Bazer. Totowa, NJ: Humana Press Inc. (https://doi.org/10.1007/978-1-4612-1804-3_18)
- Magness RR & Rosenfeld CR 1986 Systemic and uterine responses to alpha-adrenergic stimulation in pregnant and nonpregnant ewes. *American Journal of Obstetrics and Gynecology* **155** 897–904. ([https://doi.org/10.1016/S0002-9378\(86\)80047-3](https://doi.org/10.1016/S0002-9378(86)80047-3))
- Magness RR, Mitchell MD & Rosenfeld CR 1990 Uteroplacental production of eicosanoids in ovine pregnancy. *Prostaglandins* **39** 75–88. ([https://doi.org/10.1016/0090-6980\(90\)90096-E](https://doi.org/10.1016/0090-6980(90)90096-E))
- Magness RR, Rosenfeld CR, Hassan A & Shaul PW 1996 Endothelial vasodilator production by uterine and systemic arteries. I. Effects of ANG II on PGI₂ and NO in pregnancy. *American Journal of Physiology* **270** H1914–H1923. (<https://doi.org/10.1152/ajpheart.1996.270.6.H1914>)
- Magness RR, Shideman CR, Habermehl DA, Sullivan JA & Bird IM 2000 Endothelial vasodilator production by uterine and systemic arteries. V. Effects of ovariectomy, the ovarian cycle, and pregnancy on prostacyclin synthase expression. *Prostaglandins and Other Lipid Mediators* **60** 103–118. ([https://doi.org/10.1016/S0090-6980\(99\)00055-6](https://doi.org/10.1016/S0090-6980(99)00055-6))
- Magness RR, Sullivan JA, Li Y, Phernetton TM & Bird IM 2001 Endothelial vasodilator production by uterine and systemic arteries. VI. Ovarian and pregnancy effects on eNOS and NO(x). *American Journal of Physiology: Heart and Circulatory Physiology* **280** H1692–H1698. (<https://doi.org/10.1152/ajpheart.2001.280.4.H1692>)
- Mains LM, Lathi RB, Burney RO & Dahan MH 2007 Serum total testosterone levels in a patient with late onset 21-hydroxylase deficiency and a twin gestation. *Fertility and Sterility* **87** 1212.e1215–1212.e1218. (<https://doi.org/10.1016/j.fertnstert.2006.07.1545>)
- Makinen JI, Perheentupa A, Irjala K, Pollanen P, Makinen J, Huhtaniemi I & Raitakari OT 2011 Endogenous testosterone and brachial artery endothelial function in middle-aged men with symptoms of late-onset hypogonadism. *Aging Male* **14** 237–242. (<https://doi.org/10.3109/13685538.2011.593655>)
- Manikkam M, Crespi EJ, Doop DD, Herkimer C, Lee JS, Yu S, Brown MB, Foster DL & Padmanabhan V 2004 Fetal programming: prenatal testosterone excess leads to fetal growth retardation and postnatal catch-up growth in sheep. *Endocrinology* **145** 790–798. (<https://doi.org/10.1210/en.2003-0478>)
- Mason JJ, Ushijima K, Doody KM, Nagai K, Naville D, Head JR, Milewich L, Rainey WE & Ralph MM 1993 Regulation of expression of the 3 beta-hydroxysteroid dehydrogenases of human placenta and fetal adrenal. *Journal of Steroid Biochemistry and Molecular Biology* **47** 151–159. ([https://doi.org/10.1016/0960-0760\(93\)90069-9](https://doi.org/10.1016/0960-0760(93)90069-9))
- Mehrabian F & Kelishadi R 2012 Comparison of the metabolic parameters and androgen level of umbilical cord blood in newborns of mothers with polycystic ovary syndrome and controls. *Journal of Research in Medical Sciences* **17** 207–211.
- Metz TD, Allshouse AA, Euser AG & Heyborne KD 2014 Preeclampsia in high risk women is characterized by risk group-specific abnormalities in serum biomarkers. *American Journal of Obstetrics and Gynecology* **211** 512.e511–512.e516. (<https://doi.org/10.1016/j.ajog.2014.04.027>)
- Meulenbreg PM & Hofman JA 1991 Maternal testosterone and fetal sex. *Journal of Steroid Biochemistry and Molecular Biology* **39** 51–54. ([https://doi.org/10.1016/0960-0760\(91\)90012-T](https://doi.org/10.1016/0960-0760(91)90012-T))
- Milczarek R, Sokolowska E, Hallmann A, Kaletha K & Klimek J 2008 NADPH- and iron-dependent lipid peroxidation inhibit aromatase activity in human placental microsomes. *Journal of Steroid Biochemistry and Molecular Biology* **110** 230–235. (<https://doi.org/10.1016/j.jsbmb.2007.11.004>)
- Miller NR, Garry D, Cohen HW & Figueroa R 2003 Serum androgen markers in preeclampsia. *Journal of Reproductive Medicine* **48** 225–229.
- Mitchell F 2012 Reproductive endocrinology: Mass spectrometry 'gold standard' for measuring steroid sex hormones? *Nature Reviews in Endocrinology* **8** 320.
- Mori T, Watanabe K, Iwasaki A, Kimura C, Matsushita H, Shinohara K & Wakatsuki A 2014 Differences in vascular reactivity between pregnant women with chronic hypertension and preeclampsia. *Hypertension Research* **37** 145–150. (<https://doi.org/10.1038/hr.2013.131>)
- Murji A, Proctor LK, Paterson AD, Chitayat D, Weksberg R & Kingdom J 2012 Male sex bias in placental dysfunction. *American Journal of Medical Genetics: Part A* **158A** 779–783. (<https://doi.org/10.1002/ajmg.a.35250>)

- Myatt L 2002 Role of placenta in preeclampsia. *Endocrine* **19** 103–111. (<https://doi.org/10.1385/ENDO:19:1:103>)
- Naden RP & Rosenfeld CR 1981 Effect of angiotensin II on uterine and systemic vasculature in pregnant sheep. *Journal of Clinical Investigation* **68** 468–474. (<https://doi.org/10.1172/JCI110277>)
- Nakao J, Change WC, Murota SI & Orimo H 1981 Testosterone inhibits prostacyclin production by rat aortic smooth muscle cells in culture. *Atherosclerosis* **39** 203–209. ([https://doi.org/10.1016/0021-9150\(81\)90070-8](https://doi.org/10.1016/0021-9150(81)90070-8))
- Nelson SH, Steinsland OS, Wang Y, Yallampalli C, Dong YL & Sanchez JM 2000 Increased nitric oxide synthase activity and expression in the human uterine artery during pregnancy. *Circulation Research* **87** 406–411. (<https://doi.org/10.1161/01.RES.87.5.406>)
- Orlando EF, Kolok AS, Binzick GA, Gates JL, Horton MK, Lambright CS, Gray LE Jr, Soto AM & Guillette LJ Jr 2004 Endocrine-disrupting effects of cattle feedlot effluent on an aquatic sentinel species, the fathead minnow. *Environmental Health Perspectives* **112** 353–358. (<https://doi.org/10.1289/ehp.6591>)
- Osol G & Mandala M 2009 Maternal uterine vascular remodeling during pregnancy. *Physiology* **24** 58–71. (<https://doi.org/10.1152/physiol.00033.2008>)
- Palei AC, Spradley FT, Warrington JP, George EM & Granger JP 2013 Pathophysiology of hypertension in pre-eclampsia: a lesson in integrative physiology. *Acta Physiologica* **208** 224–233. (<https://doi.org/10.1111/apha.12106>)
- Palomba S, Falbo A, Russo T, Battista L, Tolino A, Orio F & Zullo F 2010 Uterine blood flow in pregnant patients with polycystic ovary syndrome: relationships with clinical outcomes. *British Journal of Obstetrics and Gynaecology* **117** 711–721. (<https://doi.org/10.1111/j.1471-0528.2010.02525.x>)
- Palomba S, Russo T, Falbo A, Di CA, Amendola G, Mazza R, Tolino A, Zullo F, Tucci L & La Sala GB 2012 Decidual endovascular trophoblast invasion in women with polycystic ovary syndrome: an experimental case-control study. *Journal of Clinical Endocrinology and Metabolism* **97** 2441–2449. (<https://doi.org/10.1210/jc.2012-1100>)
- Pan T, He G, Chen M, Bao C, Chen Y, Liu G, Zhou M, Li S, Xu W & Liu X 2017 Abnormal CYP11A1 gene expression induces excessive autophagy, contributing to the pathogenesis of preeclampsia. *Oncotarget* **8** 89824–89836. (<https://doi.org/10.18632/oncotarget.21158>)
- Park KM, Kim JI, Ahn Y, Bonventre AJ & Bonventre JV 2004 Testosterone is responsible for enhanced susceptibility of males to ischemic renal injury. *Journal of Biological Chemistry* **279** 52282–52292. (<https://doi.org/10.1074/jbc.M407629200>)
- Parks LG, Lambright CS, Orlando EF, Guillette LJ Jr, Ankley GT & Gray LE Jr 2001 Masculinization of female mosquitofish in Kraft mill effluent-contaminated Fenholloway River water is associated with androgen receptor agonist activity. *Toxicological Sciences* **62** 257–267. (<https://doi.org/10.1093/toxsci/62.2.257>)
- Pasquali R, Gambineri A, Biscotti D, Vicennati V, Gagliardi L, Colitta D, Fiorini S, Cognigni GE, Filicori M & Morselli-Labate AM 2000 Effect of long-term treatment with metformin added to hypocaloric diet on body composition, fat distribution, and androgen and insulin levels in abdominally obese women with and without the polycystic ovary syndrome. *Journal of Clinical Endocrinology and Metabolism* **85** 2767–2774. (<https://doi.org/10.1210/jcem.85.8.6738>)
- Perez-Sepulveda A, Monteiro LJ, Dobierzewska A, Espana-Perrot PP, Venegas-Araneda P, Guzman-Rojas AM, Gonzalez MI, Palominos-Rivera M, Irrazabal CE, Figueroa-Diesel H *et al.* 2015 Placental aromatase is deficient in placental ischemia and preeclampsia. *PLoS ONE* **10** e0139682. (<https://doi.org/10.1371/journal.pone.0139682>)
- Potischman N, Troisi R, Thadhani R, Hoover RN, Dodd K, Davis WW, Sluss PM, Hsieh CC & Ballard-Barbash R 2005 Pregnancy hormone concentrations across ethnic groups: implications for later cancer risk. *Cancer Epidemiology, Biomarkers and Prevention* **14** 1514–1520. (<https://doi.org/10.1158/1055-9965.EPI-04-0869>)
- Powe CE, Levine RJ & Karumanchi SA 2011 Preeclampsia, a disease of the maternal endothelium: the role of antiangiogenic factors and implications for later cardiovascular disease. *Circulation* **123** 2856–2869. (<https://doi.org/10.1161/CIRCULATIONAHA.109.853127>)
- Recabarren SE, Rojas-Garcia PP, Recabarren MP, Alfaro VH, Smith R, Padmanabhan V & Sir-Petermann T 2008 Prenatal testosterone excess reduces sperm count and motility. *Endocrinology* **149** 6444–6448. (<https://doi.org/10.1210/en.2008-0785>)
- Reckelhoff JF, Zhang H & Granger JP 1998 Testosterone exacerbates hypertension and reduces pressure-natriuresis in male spontaneously hypertensive rats. *Hypertension* **31** 435–439. (<https://doi.org/10.1161/01.HYP.31.1.435>)
- Roberts JM & Redman CW 1993 Pre-eclampsia: more than pregnancy-induced hypertension. *Lancet* **341** 1447–1451. ([https://doi.org/10.1016/0140-6736\(93\)90889-O](https://doi.org/10.1016/0140-6736(93)90889-O))
- Roberts JM, Taylor RN & Goldfien A 1991 Clinical and biochemical evidence of endothelial cell dysfunction in the pregnancy syndrome preeclampsia. *American Journal of Hypertension* **4** 700–708. (<https://doi.org/10.1093/ajh/4.8.700>)
- Rohrmann S, Sutcliffe CG, Bienstock JL, Monseque D, Akereyeni F, Bradwin G, Rifai N, Pollak MN, Agurs-Collins T & Platz EA 2009 Racial variation in sex steroid hormones and the insulin-like growth factor axis in umbilical cord blood of male neonates. *Cancer Epidemiology, Biomarkers and Prevention* **18** 1484–1491. (<https://doi.org/10.1158/1055-9965.EPI-08-0817>)
- Rosenfeld CR, Morriss FH Jr, Makowski EL, Meschia G & Battaglia FC 1974 Circulatory changes in the reproductive tissues of ewes during pregnancy. *Gynecologic Investigation* **5** 252–268. (<https://doi.org/10.1159/000301658>)
- Rosing U & Carlstrom K 1984 Serum levels of unconjugated and total oestrogens and dehydroepiandrosterone, progesterone and urinary oestriol excretion in pre-eclampsia. *Gynecologic and Obstetric Investigation* **18** 199–205. (<https://doi.org/10.1159/000299081>)
- Salam RA, Das JK, Ali A, Bhaumik S & Lassi ZS 2015 Diagnosis and management of preeclampsia in community settings in low and middle-income countries. *Journal of Family Medicine and Primary Care* **4** 501–506. (<https://doi.org/10.4103/2249-4863.174265>)
- Salamalekis E, Bakas P, Vitoratos N, Eleftheriadis M & Creatsas G 2006 Androgen levels in the third trimester of pregnancy in patients with preeclampsia. *European Journal of Obstetrics, Gynecology and Reproductive Biology* **126** 16–19. (<https://doi.org/10.1016/j.ejogrb.2005.07.007>)
- Samadi AR, Mayberry RM & Reed JW 2001 Preeclampsia associated with chronic hypertension among African-American and White women. *Ethnicity and Disease* **11** 192–200.
- Sarkar P, Bergman K, Fisk NM, O'Connor TG & Glover V 2007 Amniotic fluid testosterone: relationship with cortisol and gestational age. *Clinical Endocrinology* **67** 743–747. (<https://doi.org/10.1111/j.1365-2265.2007.02955.x>)
- Sarkar P, Bergman K, O'Connor TG & Glover V 2008 Maternal antenatal anxiety and amniotic fluid cortisol and testosterone: possible implications for foetal programming. *Journal of Neuroendocrinology* **20** 489–496. (<https://doi.org/10.1111/j.1365-2826.2008.01659.x>)
- Sathishkumar K, Balakrishnan M, Gao HJ & Yallampalli C 2011a Elevated testosterone levels during pregnancy induce preeclampsia-like syndrome in pregnant rats. *Reproductive Sciences* **18** 359a.
- Sathishkumar K, Elkins R, Chinnathambi V, Gao H, Hankins GD & Yallampalli C 2011b Prenatal testosterone-induced fetal growth restriction is associated with down-regulation of rat placental amino acid transport. *Reproductive Biology and Endocrinology* **9** 110. (<https://doi.org/10.1186/1477-7827-9-110>)
- Sathishkumar K, Elkins R, Yallampalli U, Balakrishnan M & Yallampalli C 2011c Fetal programming of adult hypertension in female rat offspring exposed to androgens in utero. *Early Human Development* **87** 407–414. (<https://doi.org/10.1016/j.earlhumdev.2011.03.001>)
- Sathishkumar K, Balakrishnan M, Chinnathambi V, Chauhan M, Hankins GD & Yallampalli C 2012 Fetal sex-related dysregulation in testosterone production and their receptor expression in the human placenta with preeclampsia. *Journal of Perinatology* **32** 328–335. (<https://doi.org/10.1038/jp.2011.101>)
- Schiffer B, Daxenberger A, Meyer K & Meyer HH 2001 The fate of trenbolone acetate and melengestrol acetate after application as growth promoters in cattle: environmental studies. *Environmental Health Perspectives* **109** 1145–1151. (<https://doi.org/10.1289/ehp.011091145>)
- Seely EW & Solomon CG 2003 Insulin resistance and its potential role in pregnancy-induced hypertension. *Journal of Clinical Endocrinology and Metabolism* **88** 2393–2398. (<https://doi.org/10.1210/jc.2003-030241>)

- Serin IS, Kula M, Basbug M, Unluhizarci K, Gucer S & Tayyar M 2001 Androgen levels of preeclamptic patients in the third trimester of pregnancy and six weeks after delivery. *Acta Obstetrica et Gynecologica Scandinavica* **80** 1009–1013. (<https://doi.org/10.1034/j.1600-0412.2001.801107.x>)
- Shao X, Liu Y, Liu M, Wang Y, Yan L, Wang H, Ma L, Li YX, Zhao Y & Wang YL 2017 Testosterone represses estrogen signaling by upregulating miR-22: a mechanism for imbalanced steroid hormone production in preeclampsia. *Hypertension* **69** 721–730. (<https://doi.org/10.1161/HYPERTENSIONAHA.116.08468>)
- Sharifzadeh F, Kashanian M & Fatemi F 2012 A comparison of serum androgens in pre-eclamptic and normotensive pregnant women during the third trimester of pregnancy. *Gynecological Endocrinology* **28** 834–836.
- Sir-Petermann T, Maliqueo M, Angel B, Lara HE, Perez-Bravo F & Recabarren SE 2002 Maternal serum androgens in pregnant women with polycystic ovarian syndrome: possible implications in prenatal androgenization. *Human Reproduction* **17** 2573–2579. (<https://doi.org/10.1093/humrep/17.10.2573>)
- Sir-Petermann T, Hirtsfeld C, Maliqueo M, Codner E, Echiburru B, Gazitua R, Recabarren S & Cassola F 2005 Birth weight in offspring of mothers with polycystic ovarian syndrome. *Human Reproduction* **20** 2122–2126. (<https://doi.org/10.1093/humrep/dei009>)
- Sladek SM, Magness RR & Conrad KP 1997 Nitric oxide and pregnancy. *American Journal of Physiology* **272** R441–R463. (<https://doi.org/10.1152/ajpregu.1997.272.2.R441>)
- Smith AS, Birnie AK & French JA 2010 Maternal androgen levels during pregnancy are associated with early-life growth in Geoffroy's marmosets, *Callithrix geoffroyi*. *General and Comparative Endocrinology* **166** 307–313. (<https://doi.org/10.1016/j.ygcen.2009.10.008>)
- Stanheewicz AE, Jandu S, Santhanam L & Alexander LM 2017 Alterations in endothelin type B receptor contribute to microvascular dysfunction in women who have had preeclampsia. *Clinical Science* **131** 2777–2789. (<https://doi.org/10.1042/CS20171292>)
- Stark MJ, Dierckx L, Clifton VL & Wright IM 2006 Alterations in the maternal peripheral microvascular response in pregnancies complicated by preeclampsia and the impact of fetal sex. *Journal of the Society for Gynecologic Investigation* **13** 573–578. (<https://doi.org/10.1016/j.jsig.2006.06.006>)
- Steckler T, Wang J, Bartol FF, Roy SK & Padmanabhan V 2005 Fetal programming: prenatal testosterone treatment causes intrauterine growth retardation, reduces ovarian reserve and increases ovarian follicular recruitment. *Endocrinology* **146** 3185–3193. (<https://doi.org/10.1210/en.2004-1444>)
- Steier JA, Ulstein M & Myking OL 2002 Human chorionic gonadotropin and testosterone in normal and preeclamptic pregnancies in relation to fetal sex. *Obstetrics and Gynecology* **100** 552–556. (<https://doi.org/10.1097/00006250-200209000-00024>)
- Sun M, Maliqueo M, Benrick A, Johansson J, Shao R, Hou L, Jansson T, Wu X & Stener-Victorin E 2012 Maternal androgen excess reduces placental and fetal weights, increases placental steroidogenesis, and leads to long-term health effects in their female offspring. *American Journal of Physiology: Endocrinology and Metabolism* **303** E1373–E1385. (<https://doi.org/10.1152/ajpendo.00421.2012>)
- Sutton-Tyrrell K, Zhao X, Santoro N, Lasley B, Sowers M, Johnston J, Mackey R & Matthews K 2010 Reproductive hormones and obesity: 9 years of observation from the Study of Women's Health across the Nation. *American Journal of Epidemiology* **171** 1203–1213. (<https://doi.org/10.1093/aje/kwq049>)
- Svarthberg J, Midtby M, Bona KH, Sundsfjord J, Joakimsen RM & Jorde R 2003 The associations of age, lifestyle factors and chronic disease with testosterone in men: the Tromsø Study. *European Journal of Endocrinology* **149** 145–152. (<https://doi.org/10.1530/eje.0.1490145>)
- Sykes SD, Pringle KG, Zhou A, Dekker GA, Roberts CT, Lumbers ER & SCOPE Consortium 2014 Fetal sex and the circulating renin-angiotensin system during early gestation in women who later develop preeclampsia or gestational hypertension. *Journal of Human Hypertension* **28** 133–139. (<https://doi.org/10.1038/jhh.2013.51>)
- Takeyama J, Sasano H, Suzuki T, Iinuma K, Nagura H & Andersson S 1998 17 β -hydroxysteroid dehydrogenase types 1 and 2 in human placenta: an immunohistochemical study with correlation to placental development. *Journal of Clinical Endocrinology and Metabolism* **83** 3710–3715. (<https://doi.org/10.1210/jcem.83.10.5212>)
- Tamimi R, Lagiou P, Vatten LJ, Mucci L, Trichopoulos D, Hellerstein S, Ekblom A, Adami HO & Hsieh CC 2003 Pregnancy hormones, preeclampsia, and implications for breast cancer risk in the offspring. *Cancer Epidemiology, Biomarkers and Prevention* **12** 647–650.
- Thornburg KL, Jacobson SL, Giraud GD & Morton MJ 2000 Hemodynamic changes in pregnancy. *Seminars in Perinatology* **24** 11–14. ([https://doi.org/10.1016/S0146-0005\(00\)80047-6](https://doi.org/10.1016/S0146-0005(00)80047-6))
- Troisi R, Potischman N, Roberts JM, Ness R, Crombleholme W, Lykins D, Siiteri P & Hoover RN 2003 Maternal serum oestrogen and androgen concentrations in preeclamptic and uncomplicated pregnancies. *International Journal of Epidemiology* **32** 455–460. (<https://doi.org/10.1093/ije/dyg094>)
- Turnipseed MR, Bentley K & Reynolds JW 1976 Serum dehydroepiandrosterone sulfate in premature infants and infants with intrauterine growth retardation. *Journal of Clinical Endocrinology and Metabolism* **43** 1219–1225. (<https://doi.org/10.1210/jcem-43-6-1219>)
- Veiga-Lopez A, Steckler TL, Abbott DH, Welch KB, MohanKumar PS, Phillips DJ, Refsal K & Padmanabhan V 2011 Developmental programming: impact of excess prenatal testosterone on intrauterine fetal endocrine milieu and growth in sheep. *Biology of Reproduction* **84** 87–96. (<https://doi.org/10.1095/biolreprod.110.086686>)
- Vyas AK, Hoang V, Padmanabhan V, Gilbreath E & Mielteka KA 2016 Prenatal programming: adverse cardiac programming by gestational testosterone excess. *Scientific Reports* **6** 28335. (<https://doi.org/10.1038/srep28335>)
- Wan J, Hu Z, Zeng K, Yin Y, Zhao M, Chen M & Chen Q 2018 The reduction in circulating levels of estrogen and progesterone in women with preeclampsia. *Pregnancy Hypertension* **11** 18–25. (<https://doi.org/10.1016/j.preghy.2017.12.003>)
- Wang YC, Su HY, Liu JY, Chang FW & Chen CH 2005 Maternal and female fetal virilization caused by pregnancy luteomas. *Fertility and Sterility* **84** 509. (<https://doi.org/10.1016/j.fertnstert.2005.02.029>)
- Warmann S, Roth C, Gluer S & Fuchs J 2000 Congenital adrenal hyperplasia associated with maternal pregnancy luteoma and the Antley-Bixler syndrome. *Journal of Pediatric Surgery* **35** 528–530. ([https://doi.org/10.1016/S0022-3468\(00\)90232-X](https://doi.org/10.1016/S0022-3468(00)90232-X))
- Williams DJ, Vallance PJ, Neild GH, Spencer JA & Imms FJ 1997 Nitric oxide-mediated vasodilation in human pregnancy. *American Journal of Physiology* **272** H748–H752. (<https://doi.org/10.1152/ajpheart.1997.272.2.H748>)
- Yanes LL, Sartori-Valinotti JC, Iliescu R, Romero DG, Racusen LC, Zhang H & Reckelhoff JF 2009 Testosterone-dependent hypertension and upregulation of intrarenal angiotensinogen in Dahl salt-sensitive rats. *American Journal of Physiology: Renal Physiology* **296** F771–F779. (<https://doi.org/10.1152/ajprenal.90389.2008>)
- Yu RM, Chaturvedi G, Tong SK, Nusrin S, Giesy JP, Wu RS & Kong RY 2015 Evidence for microRNA-mediated regulation of steroidogenesis by hypoxia. *Environmental Science and Technology* **49** 1138–1147. (<https://doi.org/10.1021/es504676s>)

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