STUDIES ON THE MODE OF ACTION OF ORAL CONTRACEPTIVES: EFFECT OF CHLORMADINONE ON PITUITARY FSH AND LH CONTENTS OF THE FEMALE RAT

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Summary. Chlormadinone, a very potent synthetic progestagen, was administered subcutaneously for 10 days (400 μg/day) to intact, unilaterally and bilaterally spayed rats, either adult or immature, and pituitary levels of FSH (hCG augmentation method of Steelman & Pohley, 1953) and LH (oAAD method of Parlow, 1958) were determined. The chlormadinone treatment induced a significant atrophy of the ovaries as well as ovarian interstitial tissue and a total blockade of ovarian compensatory hypertrophy in the unilaterally spayed rats. The pituitary concentration (μg/mg) and the total content (μg/gland) of both LH and FSH were significantly increased by chlormadinone treatment of the intact and unilaterally spayed rats despite a significant atrophy of the pituitary gland due to treatment. In the bilaterally spayed adult and immature rats, chlormadinone did not alter the ovariectomy-induced rise in LH and FSH stores of the pituitary gland.

These results suggest that chlormadinone, when administered for 10 days in the dosages employed, induces storage of FSH and LH in the pituitary gland primarily by impairing the release of these hormones, without exerting any detectable inhibitory effects on their synthesis.

INTRODUCTION

Despite their widespread use in fertility control and gynaecological disorders, the exact mode of action of synthetic progestagens remains obscure. Although it is likely that the total effectiveness of these agents probably results from their action at multiple sites in the body (such as the ovary, uterine tubes, uterus, cervix, pituitary and hypothalamus), their primary action is believed to be exerted at the pituitary-hypothalamic level, inhibiting the output of luteinizing hormone (LH) and consequently ovulation (Vorys, Ullery & Stevens, 1965; Flowers, Vorys, Stevens, Miller & Jensen, 1966).

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Previously it has been reported that prolonged administration of a progesterone derivative, Provera, results in depletion of pituitary LH stores (Labhsetwar, 1966b), while the administration of a 19-nor steroid derivative, norethynodrel, for a period of 10 or less days was found to result in a significant increase in the pituitary LH content of the female (Labhsetwar, 1967b) as well as the male rat (Brown, Wells & Youngson, 1965).

In the present study one of the most potent derivatives of progesterone, chlormadinone (6-chloro-A^6-17a-acetoxyprogesterone), was employed. It is estimated to be 3500 times more active than norethisterone, both in the Clauberg assay and anti-ovulatory test in the rabbit (Rudel & Kincl, 1966). As little as 0.5 mg of the compound taken orally without oestrogen has been reported to exert a contraceptive effect in women (Martinez-Manautou, Cortez, Giner, Aznar, Casasola & Rudel, 1966). It exhibits a very mild anti-gonadotrophic activity in parabiotic rats (Brennan & Kray, 1963) and has been found virtually free from virilizing (Kraay & Brennan, 1963) and androgenic properties (see Rudel & Kincl, 1966). Additionally it is a true progestagen in the sense that it can maintain pregnancy in spayed rats (Brennan & Kraay, 1963). It is of considerable interest to study the mode of action of such a compound in influencing the secretion of gonadotrophins (FSH and LH).

**MATERIALS AND METHODS**

Female rats of the Holtzman strain were caged in a temperature- and light- (14 hr of artificial illumination and 10 hr of darkness) controlled room and allowed free access to Purina chow and water.

Five experiments, as outlined in Table 1, were performed, employing five different types of rats. Experiments 1 and 2 were repeated at a later date (IB and 2B, Table 1). In any given experiment the control and the treated animals were of the same age, were run simultaneously and also killed on the same day, except in Exp. 1B (see Table 1).

Chlormadinone was prepared in corn or sesame oil and injected once a day subcutaneously for 10 days. The daily dose was 400 μg/rat (approximately 1.50 mg/kg in adult and 4 mg/kg in immature rats).

One day after the last injection the rats were killed with ether. The anterior pituitary glands were removed, blotted to remove excess blood, and immediately weighed. The glands within each experimental group were pooled and kept frozen (−20°C) for subsequent bio-assays. The ovaries (when present), adrenal glands and uteri were then also weighed. All autopsies were performed between 08.00 and 11.30 hours.

*Output of gonadotrophins*

This was evaluated in Exps. 2A, 2B and 3 by studying the ovarian compensatory hypertrophy in unilaterally spayed rats as described elsewhere (Labhsetwar, 1967c).

*Ovarian sensitivity*

To test the ovarian sensitivity, immature rats (26 days old) were primed with PMSG and HCG as described under LH assay. Beginning 48 hr after HCG injection,
when all ovulations have presumably occurred, the animals received daily subcutaneous injections of 400 µg (or about 4 mg/kg) of chlormadinone in oil for 8 days. A day after the last injection, the left ovary was removed as control and 1 µg of LH (NIH-LH-s-11) was injected into a tail vein. The remaining ovary was removed 4 hr±10 min later and the ascorbic acid (AA) content was determined. The percentage depletion of AA was calculated by comparing the 'treated' ovary with the control removed from the same animal.

Bio-assay for LH

The LH level was determined by a slight modification (Labhsetwar, 1967c) of the OAAD method of Parlow (1958), employing one ovary and a 4-hr interval. With the exception of experiment 1A, where a two plus one assay design was used, a symmetrical four-point design with two doses of standard LH (0.8 µg and 4 µg of NIH-LH-s-11) and two doses of pituitary tissue (0.4 mg and 2 mg) was employed. In the bilaterally spayed rats the doses of the pituitary glands were reduced by half. Initially one ovary was used and the second ovary was used 48 hr later. Four or five assay rats/dose were employed. The assay statistics (relative potency, confidence limits, index of precision, etc.) were calculated according to the method described by Gaddum (1953) for parallel line assays. The mean index of precision (λ) for ten determinations was 0.251±0.05 (s.e.).

Bio-assay for FSH

A slight modification (Labhsetwar, 1967c) of the method of Steelman &
### Table 2

**Effects of Chlormadinone on Various Organ Weights of Rats (Mean ± S.E.)**

<table>
<thead>
<tr>
<th>Expt. no.</th>
<th>Treatment</th>
<th>No. rats</th>
<th>Final B.W. (g)</th>
<th>Daily B.W. gain†</th>
<th>Organ weights (mg/100 g B.W.)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Pituitary</td>
</tr>
<tr>
<td>1A</td>
<td>None</td>
<td>7</td>
<td>214 ± 4</td>
<td>1.4</td>
<td>4.13 ± 0.08</td>
</tr>
<tr>
<td></td>
<td>Chlormadinone</td>
<td>5</td>
<td>227 ± 3*</td>
<td>2.7</td>
<td>3.46 ± 0.06*</td>
</tr>
<tr>
<td>1B</td>
<td>None</td>
<td>10</td>
<td>236 ± 3†</td>
<td>1.5</td>
<td>4.00 ± 0.11*</td>
</tr>
<tr>
<td></td>
<td>Chlormadinone</td>
<td>8</td>
<td>246 ± 3*</td>
<td>2.6</td>
<td>3.09 ± 0.13***</td>
</tr>
<tr>
<td>2B</td>
<td>None</td>
<td>8</td>
<td>245 ± 2</td>
<td>1.1</td>
<td>4.24 ± 0.14*</td>
</tr>
<tr>
<td></td>
<td>Chlormadinone</td>
<td>8</td>
<td>265 ± 2*</td>
<td>2.8</td>
<td>3.39 ± 0.03*</td>
</tr>
<tr>
<td>3</td>
<td>None</td>
<td>3</td>
<td>147 ± 2</td>
<td>3.7</td>
<td>3.80 ± 0.35</td>
</tr>
<tr>
<td></td>
<td>Chlormadinone</td>
<td>4</td>
<td>151 ± 3</td>
<td>4.3</td>
<td>3.23 ± 0.05</td>
</tr>
<tr>
<td>4</td>
<td>None</td>
<td>6</td>
<td>276 ± 4</td>
<td>3.5</td>
<td>3.49 ± 0.20</td>
</tr>
<tr>
<td></td>
<td>Chlormadinone</td>
<td>6</td>
<td>273 ± 4</td>
<td>2.9</td>
<td>3.39 ± 0.05</td>
</tr>
<tr>
<td>5</td>
<td>None</td>
<td>9</td>
<td>174 ± 4</td>
<td>5.2</td>
<td>3.58 ± 0.07</td>
</tr>
<tr>
<td></td>
<td>Chlormadinone</td>
<td>9</td>
<td>173 ± 4</td>
<td>5.1</td>
<td>3.41 ± 0.09</td>
</tr>
</tbody>
</table>

* P < 0.05; **P < 0.01; *** P < 0.005.

† Mean initial B.W. — Mean final B.W.

†† Body weights taken on the day when treated animals were killed; however, organ weights (mg/100 g B.W.) calculated from body weights taken at autopsy on the day of first dioestrous smear.
Pohley (1953) was used. The assays were conducted on two occasions. Each assay included two doses of reference standard (NIH-FSH-s-3) and one dose of unknown from each treatment group, several unknowns being tested on each occasion. The augmenting dose of HCG was 40 i.u./rat, and three or four assay rats/dose were used. The index of precision calculated from pooled sum of squares and combined slope was found to be 0.123, which is well within acceptable range.

RESULTS

Body weight
Administration of chlormadinone to rats with one or both ovaries resulted in a significant increase in body weight \( (P<0.05, \text{Table 2}) \). The mean daily body weight gain in three groups of adult treated rats (two intact and one unilaterally spayed) was significantly higher than a similar mean for three corresponding control groups \( (2.70\pm0.03 \text{ g versus } 1.33\pm0.18 \text{ g}, P<0.01) \). Bilateral ovariectomy itself significantly increased the body-weight gain; chlormadinone failed to augment this weight gain, either in adult or immature rats (Table 2).

Organ weights
The chlormadinone administration resulted in a significant atrophy of the pituitary glands, ovaries and adrenals in the intact and unilaterally spayed rats, both on a relative \( (\text{mg/100 g body wt, Table 2}) \) and an absolute \( (\text{mg/rat, Tables 3 and 4}) \) body weight basis. In the adult rats, bilateral ovariectomy itself reduced the pituitary and adrenal weights significantly when compared with the intact controls (Table 2); chlormadinone in these rats further reduced the adrenal but not the pituitary weights \( (P<0.01, \text{Table 2}) \).

In the intact and unilaterally spayed rats treated with chlormadinone, the relative uterine weights were consistently and sometimes significantly \( (P<0.05) \) lower than those of the respective controls. In contrast, in the bilaterally spayed adult as well as immature rats treated with progestagen, uterine weights were significantly higher than those of the respective controls \( (P<0.05, \text{Table 2}) \), confirming the previous report (Harper, 1964). At autopsy none of the treated rats with one or both ovaries revealed ballooned uteri, while in the control groups (except in Exp. 3) some of the animals showed this condition.

Ovarian histology
Paraffin sections \( (7 \mu \) were stained either with haematoxylin–eosin or a tetrachrome method, which selectively stains collagen. The corpora lutea in the treated animals, although fewer in number, appeared healthy and contained fewer connective tissue elements than the controls. Vesicular follicles of varying sizes, up to but not including pre-ovulatory size, were present in the ovaries of the treated animals, but most of them appeared atretic. The interstitial tissue in the ovaries of the treated animals also appeared atrophic.

Ovarian compensatory hypertrophy
Progestagen treatment resulted in inhibition of ovarian compensatory hyper trophy in all experiments (Table 3), and prevented the compensatory increase
in the number of corpora lutea usually seen in the unilaterally spayed rats (Table 3).

**Ovarian sensitivity**

Chlormadinone treatment did not significantly affect the ovarian weights (225±18 mg in control, versus 200±13 mg in treated, $P>0.05$) or initial AA concentration (48.8 in control, versus 51.1 µg/100 mg of ovary in the treated). Furthermore, exogenous LH caused the same degree of AA depletion in the treated as in the control animals (17.3±2.3, versus 20.7±5.6%, $P>0.05$).

### Table 3

**Influence of Chlormadinone on Ovarian Compensatory Hypertrophy and Luteal Count in Unilaterally Spayed Rats (Mean±S.E.)**

<table>
<thead>
<tr>
<th>Experiment no.</th>
<th>Treatment</th>
<th>No. rats</th>
<th>Left ov. wt. (mg)*</th>
<th>Right ov. wt. (mg)*</th>
<th>Ovarian hypertrophy (%)</th>
<th>No. CL in rt ov.</th>
</tr>
</thead>
<tbody>
<tr>
<td>2A</td>
<td>None</td>
<td>4</td>
<td>38.0±2.7</td>
<td>56.5±1.2***</td>
<td>49</td>
<td>9.7±0.5</td>
</tr>
<tr>
<td></td>
<td>Chlormadinone</td>
<td>5</td>
<td>35.0±2.5</td>
<td>33.0±1.0</td>
<td>—</td>
<td>5.0±0.7***</td>
</tr>
<tr>
<td>2B</td>
<td>None</td>
<td>8</td>
<td>38.2±1.5</td>
<td>52.2±2.3***</td>
<td>37</td>
<td>11.9±0.4</td>
</tr>
<tr>
<td></td>
<td>Chlormadinone</td>
<td>8</td>
<td>40.7±3.3</td>
<td>34.5±1.9</td>
<td>—</td>
<td>6.2±2.5**</td>
</tr>
<tr>
<td>3c</td>
<td>None</td>
<td>3</td>
<td>17.0±2.7</td>
<td>39.0±1.1***</td>
<td>129</td>
<td>10.3±1.2</td>
</tr>
<tr>
<td></td>
<td>Chlormadinone</td>
<td>4</td>
<td>15.4±1.2</td>
<td>23.0±2.2**</td>
<td>49</td>
<td>3.5±1.2**</td>
</tr>
</tbody>
</table>

* $P<0.05$; ** $P<0.01$; *** $P<0.005$.
* Weighed 10 days after removal of left ovary.
* *Significantly different from the respective controls.
* c Significantly different from left ovarian weight.
* d Vaginae of all animals except one were closed at the time of first injection on 36th day of life.
* e mg/rat.

**Pituitary LH and FSH**

In the control animals unilateral ovariectomy did not significantly modify the pituitary levels of either LH or FSH (Table 4), confirming the previous reports (Edgren, Parlow, Peterson & Jones, 1965; Labhsetwar, 1967c). In contrast, the removal of both ovaries resulted in a several-fold increase in the pituitary stores of both gonadotrophins (Table 4).

Administration of chlormadinone to intact and unilaterally spayed rats resulted in marked increases (>100%) in the concentration (µg/mg) and the total content (µg/pituitary) of both LH and FSH (Table 3). The gonadotrophin content (µg/gland) increased, despite a significant atrophy of the pituitary gland both on a relative (Table 2) and an absolute basis (Table 4) due to treatment.

In the bilaterally spayed adult and immature rats, progestagen treatment did not alter the ovariectomy-induced rise in the pituitary stores of either LH or FSH (Table 4).

**FSH/LH ratio**

The ratio of FSH concentration (µg/mg) to that of LH was always in favour of FSH in both the control and the treated animals. The removal of one or both ovaries or administration of chlormadinone did not materially affect the ratio (Table 4).
<table>
<thead>
<tr>
<th>Expt. no.</th>
<th>Treatment</th>
<th>Pit. wt (mg)</th>
<th>LH* conc. (µg/mg)</th>
<th>95% confidence limits</th>
<th>LH content (µg/gland)</th>
<th>% change</th>
<th>FSH* conc. (µg/mg)</th>
<th>95% confidence limits</th>
<th>FSH content (µg/gland)</th>
<th>% change</th>
<th>Ratio* FSH/LH</th>
</tr>
</thead>
<tbody>
<tr>
<td>1A</td>
<td>None</td>
<td>9.27</td>
<td>0.50*</td>
<td>0.02-1.15</td>
<td>4.63</td>
<td>+123</td>
<td>1.0</td>
<td>0.7-1.3</td>
<td>9.3</td>
<td>+414</td>
<td>1.5</td>
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<td></td>
<td>Chlormadinone</td>
<td>7.86*</td>
<td>1.30*</td>
<td>0.71-2.35</td>
<td>10.35</td>
<td></td>
<td>6.0</td>
<td>2.4-9.6</td>
<td>47.8</td>
<td></td>
<td>4.0</td>
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<tr>
<td>1B</td>
<td>None</td>
<td>9.37</td>
<td>0.39</td>
<td>0.19-0.79</td>
<td>3.65</td>
<td>+160</td>
<td>1.3</td>
<td>1.1-1.5</td>
<td>12.3</td>
<td>+158</td>
<td>3.4</td>
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<td></td>
<td>Chlormadinone</td>
<td>7.61*</td>
<td>1.25</td>
<td>0.81-1.91</td>
<td>9.51</td>
<td></td>
<td>4.2</td>
<td>2.9-5.5</td>
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<td></td>
<td>3.3</td>
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<td>10.40</td>
<td>0.73</td>
<td>0.45-1.17</td>
<td>7.59</td>
<td>+137</td>
<td>1.0f</td>
<td>—</td>
<td>10.4</td>
<td>+385</td>
<td>1.4</td>
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<td></td>
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<td>2.00</td>
<td>1.14-3.50</td>
<td>18.00</td>
<td></td>
<td>5.6</td>
<td>3.6-7.6</td>
<td>50.4</td>
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<td>2.8</td>
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<td>9.70</td>
<td>7.33</td>
<td>4.52-11.89</td>
<td>71.10</td>
<td>-24</td>
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<td>14.1-23.5</td>
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<td>28.4</td>
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<td>261</td>
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<td>4.3</td>
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<td>6.22</td>
<td>6.65</td>
<td>3.24-13.64</td>
<td>41.36</td>
<td>-29</td>
<td>20.1</td>
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<td>125</td>
<td>+9</td>
<td>2.9</td>
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<td>4.97</td>
<td>2.72-9.27</td>
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<td></td>
<td>23.2</td>
<td>22.7-23.6</td>
<td>136</td>
<td></td>
<td>4.1</td>
</tr>
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</table>

* P < 0.05.
*µg equivalent of NIH-LH-s-11/mg wet pituitary.
*µg equivalent of NIH-FSH-s-3/mg wet pituitary.
* Ratio of FSH concentration (µg/mg) to that of LH.
*Animals in this group killed when in dioestrus.
* A 2 + 1 assay design; in all others 2 + 2 assay design.
* Value based on two assay rats only.
*mg/rat.
DISCUSSION

The data suggest that chlormadinone administration results in increased pituitary stores of FSH and LH. A significant atrophy of the ovaries in the intact treated rats (Table 2), together with an inhibition of ovarian compensatory hyper-trophy (Table 3), a lack of compensatory increase in the number of corpora lutea in the unilaterally spayed rats (Table 3) and the atrophy of the interstitial tissue of the ovary indicate either a decline in the output of FSH and LH, or a decrease in the ovarian sensitivity to these hormones. However, ovaries of immature pseudopregnant rats treated with this progestagen were as sensitive to the ovarian-ascorbic-acid-depleting activity of exogenous LH as those of the controls. Furthermore, ovaries in rats (Harper, 1964) and rabbits (Rudel & Kincl, 1966) treated with this compound remain capable of responding to gonadotrophins. These observations, taken together, preclude a significant peripheral action of the agent. Therefore our observations can be most readily explained by assuming a decline in the discharge of both FSH and LH from the pituitary gland of the treated rats. The decreased output of FSH and LH associated with their increased pituitary stores suggests that chlormadinone primarily impaired the release of these hormones, without significantly interfering with their synthesis in the pituitary gland. This is further corroborated by the failure of this compound to reduce the ovariectomy-induced rise in the pituitary levels of FSH and LH (Table 4).

Harper (1965) postulated, from the augmented uterine response to placental gonadotrophins of immature rats fed chlormadinone, that this progestagen causes storage of gonadotrophins in the pituitary gland. The present study involving adult rats provides experimental evidence in support of this hypothesis, and further demonstrates that the storage involves both FSH and LH.

The storage of gonadotrophins in the pituitary may result from the action of the compound at the pituitary and/or hypothalamic level, or it may be secondary to its ability to induce pseudopregnancy. The latter condition is associated with increases in the pituitary stores of LH and FSH (Van Rees & de Groot, 1965; Labhsetwar, 1967a; Schwartz & Rothchild, 1964), while progesterone (Everett, 1964) as well as synthetic progestagen-norethynodrel (Labhsetwar, 1966a) have been found capable of inducing pseudopregnancy in the rat.

It seems that inhibition of LH release by chlormadinone is partial, involving only the ovulatory surge of the hormone. The presence of vesicular follicles of varying sizes in the ovaries of the treated animals indicates that gonadotrophins (particularly FSH) adequate to support follicular growth were being secreted in the presence of chlormadinone. These follicles probably secrete oestrogens, since significant uterine atrophy did not always result after administration of chlormadinone to animals with one or both ovaries. Alternatively, the uterotrophic action of the agent may have overcome the oestrogen deficiency resulting from reduction in LH output. The uterotrophic property of this agent was evident in the absence of both ovaries (Table 2).

Two important properties of this compound observed in this study, are its ability to (i) increase body weight, and (ii) cause an atrophy of the pituitary gland, both on an absolute and a relative basis, in intact and unilaterally
spayed rats. Since both of these effects were absent in the bilaterally spayed rats, they appear to be mediated through the ovary and most probably result from neutralization of effects of endogenous oestrogens by chlormadinone. The anti-oestrogenic property of this compound is well documented (Kraay & Brennan, 1963). In contrast, the adrenolytic effect of chlormadinone appears to be independent of the ovaries (Table 2).

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The author wishes to thank Dr A. C. Enders for his interest and support throughout the course of the work and for advice on histological procedures employed in this study.

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