TRANSFER AND VIABILITY OF ONE-CELL OVA IN SHEEP*

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Summary. Attempts to obtain fertilization of sheep ova recovered from follicles and deposited in the oviducts of the same ewes were unsuccessful. The recovery and transfer of postovulatory one-cell ova was successful if the paired ewes were closely synchronized as to the end of oestrus. Fertilization apparently was completed soon after the end of oestrus in mated ewes and ova from non-mated ewes were apparently still capable of normal fertilization and development at this time. Ova from non-mated ewes usually do not have corona cells present when recovered from the oviduct. Both fertilized and unfertilized ova appear to be able to withstand stress associated with a single transfer, but a second transfer greatly reduces the chances of a successful pregnancy.

INTRODUCTION

It has been shown by Alliston & Ulberg (1961), using embryo transfer techniques, that embryos produced by ewes maintained at 90°F are irreversibly damaged by 3 days after mating, although the damage is not morphologically apparent. Dutt (1960) placed recently mated ewes in rooms heated to 90°F and found that pregnancy failure, due to high environmental temperatures, steadily decreased as the age of the embryo increased. These observations indicate that the potential young are most sensitive to damage at a very early stage in development, possibly even before fertilization, causing death at a later stage of development.

There are three stages in the reproductive process where this damage could occur: (1) the unfertilized ovum, (2) the spermatozoa after deposition in the ewe but before fertilization, and (3) the ovum after fertilization. The damage could also be cumulative in the three stages. This paper is the result of efforts to develop techniques for the study of unfertilized sheep ova in relation to causes of early embryonic death.

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MATERIALS AND METHODS

Beginning in the late winter of 1960, and continuing through December 1961, approximately 100 grade Western Blackface ewes were used in an attempt to develop a technique for the recovery and transfer of unfertilized ova. Oestrous control in anoestrous and in some of the oestrous ewes was effected by fourteen daily subcutaneous injections of 10 mg progesterone in oil followed by 750 i.u. (anoestrous ewes) or 500 i.u. (oestrous ewes) of pregnant mares’ serum (PMS) 48 hr after the last progesterone injection. Vasectomized or aproned rams were used twice daily for detection of oestrus. Oestrous ewes were isolated without feed or water and recovery of the ova was attempted 12 to 36 hr later.

Ewes were anaesthetized lightly with sodium pentobarbital, a mid-ventral laparotomy was made, and the ova recovered. The ova were examined under a low power stereomicroscope (×10 to 60) and then deposited into the infundibular end of the oviduct or into the uterine lumen by puncture of the uterine wall. Ewes not returning to heat were laparotomized 25 to 35 days later and pregnancy was determined by palpation of the uterus for an amnionic vesicle.

Where ova were recovered from the oviduct, the technique used was similar to that developed by Hunter, Adams & Rowson (1955) and modified by Alliston & Ulberg (1961). Polyethylene tubing was inserted about 1 cm into the infundibular end of the oviduct and held there by a wound clip applicator. The flushing solution was injected into the uterus and forced through the oviduct and tubing into a watchglass (Pl. 1, Fig. 1). Three different flushing solutions were used: (1) modified Krebs’ solution (Lardy & Phillips, 1943) for recovery from follicles, (2) 50 % modified Krebs’ solution and 50 % homologous blood serum for recoveries from the oviduct the 1st year, and (3) 75 % modified Krebs’ solution and 25 % serum from the mated ewe in the 2nd year of the postovulatory transfer.

Preovulatory ova were recovered from follicles (judged to be closest to the time of ovulation) of mated ewes 12 to 24 hr after onset of heat. A sharpened glass tube or 15-gauge hypodermic needle, attached to a short length of polyethylene tubing, was used in recovering the ova. The point of the glass tube was inserted into the side of the follicle and the ovum was then collected by injecting the flushing solution into the follicle from the opposite side with a syringe and collecting the washings in a watchglass. On some occasions ova were recovered from small follicles and deposited in oviducts without other ova. Recovery from these small follicles was made by puncturing the follicle with a sharpened glass capillary tube drawn from 6-mm tubing and allowing capillary action to draw in the ovum. Ova were recovered for re-examination 24 to 48 hr after the original deposition, examined for cleavage, and redeposited in the same oviduct, providing they were not degenerate. Control ewes for measuring inherent fertility were mated, the ova recovered from the oviducts approximately 3 days later and examined before deposition in the opposite uterine horn of the same ewe.

In the recovery and transfer of postovulatory unfertilized ova, the onset of
All phase-contrast photomicrographs of ova are made of unstained ova shortly after recovery and slightly compressed by a coverslip.

Fig. 1. Technique used for flushing ova from oviducts.

Fig. 2. Preovulatory ovum with cumulus cells. Preparation in modified Krebs' solution. 200.

Fig. 3. Unfertilized ovum in modified Krebs' solution. 400.

Fig. 4. Fertilized ovum with pronuclei in modified Krebs' solution. 400.

Fig. 5. Fertilized ovum with two polar bodies in 25% serum, 75% modified Krebs' solution. > 200.

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oestrus was synchronized in pairs of ewes to within 12 hr either by hormonal control or by natural synchronization. One ewe of each pair was mated, usually twice, before a transfer, and the transfer was attempted approximately 24 to 36 hr after onset of oestrus. Initially the time of transfer was arbitrarily selected (24 or 36 hr after initiation of oestrus), while later the time of transfer depended upon synchronous end of heat. In the latter ewes the transfer was performed shortly after they were first found to be out of heat.

Since removal of the fertilized ovum was desired, reciprocal transfers were made, the mated ewe received the unfertilized ovum and the non-mated ewe the ovum from the mated ewe. Initially ova were recovered from the oviducts a second time 24 to 48 hr later, examined for cleavage and redeposited. Later transfers did not include a second recovery. Control animals to measure inherent fertility included ewes in which the ova were transferred between animals approximately 3 days after mating and also ewes in which no transfers were performed.

RESULTS
PREOVULATORY OVA
Thirty-one ova were recovered from follicles and deposited in the oviducts of the ewes from which they originated. The ova were covered with corona radiata cells (Pl. 1, Fig. 2). Eight of these ova were examined 24 to 48 hr later, but none had cleaved. Two of these, however, had two polar bodies 24 hr after being deposited in the oviducts. No pregnancies resulted from transfers of follicular ova whether or not the second recovery was successful. These ewes were in anoestrus; however, three of seven control transfers (4- to 16-cell) resulted in pregnancies and all ova from control ewes were cleaving at the time of transfer.

POSTOVULATORY OVA
Since the transfer of preovulatory ova did not appear feasible, the transfer of postovulatory ova was undertaken next. Ova recovered from non-mated ewes normally were devoid of corona radiata cells and had one polar body present (Pl. 1, Fig. 3). Ova from mated ewes normally had two polar bodies (Pl. 1, Fig. 5), but occasionally only one was observed, indicating fertilization may not have occurred. Also, various rearrangements of cytoplasmic granules were observed in ova from mated ewes, probably indicating various stages of syngamy (Pl. 1, Fig. 4).

When the ova originally recovered from non-mated ewes were examined for cleavage (Table 1), a greater percentage of those recovered 24 hr after the initial detection of oestrus were found to be cleaving (67%) than were those originally recovered 36 hr after the initial detection of oestrus (60%). Conversely, when the ova originally recovered from mated ewes were examined, a greater percentage of those recovered 36 hr after the initial detection of oestrus were cleaving (67%) than were those originally recovered 24 hr after initial detection of oestrus (33%). One degenerating ovum, transferred from an unmated ewe at 36 hr, was found at cleavage check. The only pregnancy
resulting from a transfer of an unfertilized ovum was in the 36-hr group. The only pregnancy resulting from an ovum transferred from a mated ewe was in the 36-hr group; however, the attempt to recover it for re-examination had failed.

Since the pregnancy rate was low, but an acceptable degree of fertilization (as indicated by cleavage) was obtained, a third group of transfers was performed without making the cleavage check. The previous results support the data of Green & Winters (1935) and McKenzie & Terrill (1937), who showed

| Table 1 |
| FATE OF OVA RECIPIROCALLY TRANSFERRED AFTER EXAMINATION DURING TWO STAGES OF DEVELOPMENT |

<table>
<thead>
<tr>
<th>Ova from mated ewes</th>
<th>Ova from non-mated ewes</th>
</tr>
</thead>
<tbody>
<tr>
<td>24 hr*</td>
<td>36 hr*</td>
</tr>
<tr>
<td>Transfers attempted</td>
<td>7</td>
</tr>
<tr>
<td>Ewes ovulated</td>
<td>4†</td>
</tr>
<tr>
<td>Ova recovered</td>
<td>3</td>
</tr>
<tr>
<td>Recovered second</td>
<td>3</td>
</tr>
<tr>
<td>examination†</td>
<td>1</td>
</tr>
<tr>
<td>Ova cleaving</td>
<td>0</td>
</tr>
<tr>
<td>Ewes pregnant</td>
<td>0</td>
</tr>
</tbody>
</table>

* Time of transfer after onset of oestrus.
† Ewes not ovulated at 24 hr are also included in the 36-hr group.
§ Twenty-four to 48 hr after original recovery.
∥ Pregnancy from ovum not recovered the second time.

| Table 2 |
| INITIATION OF PREGNANCY FROM OVA RECIPIROCALLY TRANSFERRED AT END OF OESTRUS |

<table>
<thead>
<tr>
<th>Source of ova</th>
<th>Mated ewes</th>
<th>Non-mated ewes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Transfers attempted</td>
<td>13</td>
<td>13</td>
</tr>
<tr>
<td>Ova recovered</td>
<td>11</td>
<td>10</td>
</tr>
<tr>
<td>Ewes pregnant</td>
<td>4</td>
<td>4*</td>
</tr>
</tbody>
</table>

* Does not include a probable degenerating embryo found at 30 days after mating in one ewe.

that ewes ovulate very close to the end of oestrus, so recoveries were then attempted after the synchronous end of oestrus. As shown in Table 2, 36% of the ova (four out of eleven) transferred from mated ewes resulted in pregnancies and 40% of the ova (four out of ten) transferred from non-mated ewes resulted in pregnancies. A probable degenerating embryo was also found at 30 days in one ewe receiving an ovum from a non-mated ewe.

Two of seven ewes (28.6%) receiving 8- to 16-cell embryos (control transfers) were pregnant and a third had a probable degenerating embryo. Seven of seven non-transferred control ewes were pregnant. No reason can be given
for the poor results with ewes receiving 8- to 16-cell embryos; the anoestrous control ewes of the preovulatory group of transfers had given better results (three out of seven pregnant).

Although the overall pregnancy rate seems low, the data include results obtained while the techniques were being developed. The results with the ova from mated ewes were particularly poor until the flushing medium was changed from 50% homologous serum and 50% modified Krebs’ solution to 25% serum from the mated ewe and 75% modified Krebs’ solution. Four pregnancies resulted from seven such transfers after the change in medium versus none out of four transfers before the change (excluding the data of Table 1). In the last group of transfers performed (included as part of Table 2) three of four ova from mated ewes resulted in pregnancies and two of three ova from non-mated ewes resulted in pregnancies.

**DISCUSSION**

The transfer of one-cell ova, especially unfertilized ova, presents problems not associated with the transfer of 8- to 16-cell embryos. Noyes (1952) obtained up to 40% successful transfer of preovulatory ova in rats if the first maturation division had occurred. Chang (1955) reported that preovulatory rabbit ova would mature in the oviducts or in dilute serum, but only one of thirty-nine such oocytes developed into a normal foetus. It is even more difficult, if not impossible at present, to determine the time of impending ovulation in sheep, with enough accuracy to obtain mature ova. Green & Winters (1935) and McKenzie & Terrill (1937) have shown that most ewes ovulate shortly before or close to the end of oestrus. However, the results obtained in this study were not encouraging for the transfer of preovulatory ova in sheep. Only two of eight ova recovered a second time gave any indication of being fertilized and no pregnancies occurred from a total of thirty-one preovulatory ova transferred. The data from the control ewes indicated that it was possible to obtain a satisfactory fertilization rate and a fair number of pregnancies at that time of year from this group of sheep. This is in accord with the data of Lopyrin, Loginova & Karpov (1950a, b) who indicated that they obtained poor results in transfers of preovulatory ova. Only one of forty-three ewes receiving a preovulatory ovum was reported as becoming pregnant whereas seven of forty-six ewes receiving 1- to 2-day ‘zygotes’ produced lambs.

After ovulation, unfertilized ova of various species are subject to ageing; pigs—Dzuik (1960) and Hancock (1961), ferrets—Chang (1950), Green & Winters (1935) estimated the life of the unfertilized sheep ovum as less than 24 hr. Thibault & Ortavant (1949) have shown that sheep ova may be artificially activated by cold shock. The data presented here indicate that unfertilized ova are probably most viable when recovered at 24 hr after initiation of oestrus, but not all ewes have ovulated at this time and ova from mated ewes may not be fertilized at this time. If transfer is made at 36 hr after initiation of oestrus, ageing of some ova appears to occur. However, if the transfers are made soon after the end of oestrus, without regard to the time, then an acceptable degree of success is obtained.
It may be noted that unfertilized ova were devoid of corona radiata cells, indicating that the presence of spermatozoa was not necessary for the removal of these cells. It may also be noted that the transfer apparently imposes a stress on the reproductive process, since two transfers greatly lowered the chances for a successful pregnancy as compared to a single transfer.

Overall, the recovery rate of postovulatory ova was quite high; forty-two ova were recovered from fifty ewes with fifty-one corpora lutea for 82.4% recovery, which would indicate that ova enter the oviduct very soon after ovulation.

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REFERENCES


