Assessment of spermicides by a stripping technique against human spermatozoa

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Summary. Fifty-two (52) compounds were tested for spermicidal activity by titration against human spermatozoa. The gradual decrease in mean sperm size was measured against increasing concentration of spermicide and the end-point was taken as the point at which all the peripheral cytoplasm had been removed and only the sperm core of nucleus and tail fibres remained. There were 14 compounds that produced this total effect. All were detergents, of various types, and the effect was purely physical. The most potent compounds caused complete stripping at 0·5–50 pmol/cell and most are already used in spermicidal preparations. A further 11 compounds, including sodium hypochlorite and some phenols, caused partial stripping, while 4 compounds caused sperm swelling. The test was not suitable for assessment of metabolic cell poisons.

Introduction

The methods of assessment and the types of potent spermicides have been reviewed by Bernstein (1974). The usual method of measuring the cytotoxic potencies of compounds against specific cell types is to measure the degree of inhibition of growth, i.e. of cell division. This method cannot be used to assess the potency of spermicides and it has been usual to measure some morphological or biochemical sperm characteristic as the end-point, e.g. the decrease in percentage of motile spermatozoa after a set time or the time taken for all the spermatozoa to become immotile. Since the work of Baker (1931) many possible spermicidal agents, as pure compounds (Holzaepfel, Greenlee, Wyant & Ellis, 1959; Harvey & Stuckey, 1962a,b) and as pharmaceutical preparations (Gamble, 1953, 1957; Hartman, 1959; Goldenberg & White, 1975), have been tested in this way. Tests depending on the estimation of the inhibition of metabolic features, e.g. respiration, are relatively few.

A new method has now been developed based on the measurement of sperm size. During measurements of the size of human spermatozoa (Brotherton & Barnard, 1974), it was found that human semen contains so many small fragments and droplets that it was first necessary to dissolve these by treating with Zaponin reagent (Coulter Electronics Ltd, Dunstable, England). This procedure also removed the peripheral cytoplasm from the spermatozoon, leaving the nucleus and the tail fibres to form the 'sperm core', which was the body measured. When it became possible to clean untreated spermatozoa, the loss of total sperm volume due to Zaponin treatment was found to be about 50% (Brotherton, 1975). Application of the technique to the spermatozoa of other species gave a wide range of values. Attempts were then made to separate sperm heads from tails, to measure their respective sizes and study their differing compositions. Although the heads and tails of rodent and rabbit spermatozoa were easily separated from each other, it was found that all procedures led to a significant loss in total sperm volume and that the methods were all unsuccessful for human spermatozoa (Brotherton, 1977a). Ultrasonication caused unspecific sperm fragmentation, rather than cleavage, and a large loss in total sperm volume. This work to find an agent that would cleave human spermatozoa has now been extended and several substances were found to strip spermatozoa in the same manner as Zaponin reagent. It has therefore been possible to assess their potency as spermicides by following the decrease in sperm size as the spermicide concentration is increased.

Materials and Methods

A Coulter Counter, Model Z_B Industrial, was used to count and size human spermatozoa as described previously (Brotherton & Barnard, 1974). Human semen samples were obtained from the sub-fertility clinic and were selected for a count of greater than 40×10^6/ml, normal motility and more than 60 % normal cells. After suitable dilution in Isoton reagent (Coulter Electronics Ltd) the sperm count and size were determined before and after Zaponin treatment. Because of the normal debris in semen the size of the spermatozoa before Zaponin treatment could only be determined approximately. A sample of semen was then diluted with the same amount of Isoton and a known volume of the test substance in Isoton was added. Sperm count and size were measured after exactly 1 min. A fresh portion of semen was taken for each point on the titration curve of a particular substance as it had previously been found that human spermatozoa in Isoton swell over a period of 1–2 h. Similarly, increasing concentrations of test substance could not be added to the same diluted semen sample as in a normal chemical titration as this invalidated the method of calculation. The amount of test substance was related to the number of spermatozoa present in each titration. All titrations were carried out at pH 7·0–7·2 because preliminary investigations showed that sodium hydroxide alone would strip spermatozoa while hydrochloric acid caused swelling. An alkaline pH was found to be extremely detrimental to human spermatozoa.

All chemicals were purchased from Merck, Darmstadt, Germany, unless otherwise stated. The remaining chemicals were purchased from the German agents for the major international chemical firms indicated. Items identified by a trade name were from firms which were not necessarily the primary manufacturers.

Results

Text-figure 1 shows the titration curves obtained for some of the most potent spermicides. The lower horizontal line on each curve is the size of the sperm core after Zaponin treatment for that particular semen sample. It is equivalent to the 'all' effect in a microbiological assay (Brotherton, 1976a). The upper horizontal line is the approximate size of the spermatozoa before Zaponin treatment. It is
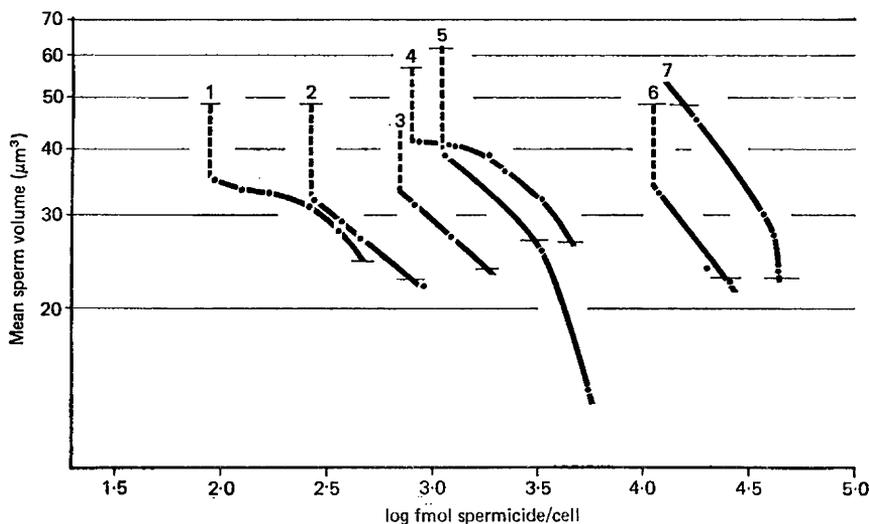
Text-fig. 1. Dose–response curves for the assay of spermicides by the stripping of human spermatozoa; see text for details. 1, Cetylpyridinium chloride; 2, CTAB; 3, Hyamine 10X; 4, benzalkonium chloride; 5, sodium lauryl sulphate; 6, Triton N-101; 7, Triton X-100.

equivalent to the 'nothing' effect in a microbiological assay. In contrast to microbiological assays the whole length of the dose–response curve could not be determined because of the debris in un-treated semen. The debris was only dissolved when the active substance had removed enough sperm cytoplasm to allow the sperm volume to approach that of the core and the size of the treated sperma-tozoa could then be measured accurately. Similarly the ED_{50} value could not be determined as in a microbiological assay. The end-point of the assay was taken as the amount of substance required to just strip the spermatozoa to the core size. This could be measured very accurately from the dose–response curves and the values for the most potent spermicides are shown in Table 1.

Table 1. Minimum concentrations of reagents to produce 100 % stripping of human spermatozoa

	Conc. (pmol/cell)
Triisopropylnaphthalene sulphonic acid sodium salt (Serva)	77·6
Triton X-100 (Merck)	43·7
N-Dodecylpyridinium chloride (Merck)	31·6
Sodium deoxycholate (Merck)	26·9
Triton N-101 (Sigma)	24·0
N-Lauroylsarcosine sodium salt (Sigma)	12·6
Benzethonium chloride (Serva)	6·03
Benzalkonium chloride (Merck)	4·57
Teepol 710 (Serva)	3·09
Sodium lauryl sulphate (Merck)	3·02
Hyamine 10X (Sigma)	1·82
Saponin (Merck; B.D.H.)	0·955
CTAB (Merck)	0·794
Cetylpyridinium chloride (Merck)	0·468

Table 2. Approximate spermicidal potency of compounds that strip human spermatozoa incompletely

	Stripping %	Conc. (pmol/cell)
Sodium hypochlorite	37·8	489,000
	58·3	406,000
	43·3	923,000
	6·0	92,700
	15·5	185,000
	55·2	277,000
Chloramine T (Merck)	18·7	9330
	9·2	2630
	14·0	10,900
	16·5	4580
Brij 35 (Serva)	35·6	16,300
Sodium hydroxide	18·4	60·3
	45·7	151
	51·5	302
Benzyltrimethylammonium chloride (Merck)	17·0	97·1
Ethyl potassium xanthate (Merck)	18·7	155
2-Naphthol (Merck)	38·3	85·6
	37·7	61·4
	31·1	113
Sodium cholate (Merck)	48·5	24·0
4-n-Hexylresorcinol (Merck)	31·4	19·1
2,4,6-Trichlorophenol (Merck)	77·0	11·0
6-Chlorothymol (Merck)	33·1	6·34
	37·9	4·98

Several substances were found that partly stripped human spermatozoa (Table 2). The addition of more substance either failed to bring about further stripping or was not possible for solubility reasons. As the core size was not reached it was not possible to compare their potencies with those of the compounds shown in Table 1 but an approximate idea can be obtained from the degree of stripping obtained relative to the approximate original size of the spermatozoa. Many of these substances appear to be almost as potent as spermicides as those shown in Table 1.

Table 3. Compounds (from Merck) which cause swelling of human spermatozoa

	Approx. increase in sperm vol. (%)	Conc. (pmol/cell)
Hydrochloric acid	18·2	1700
	7·1	439
Acetic acid	3·1	4940
Sodium diethyldithiocarbamate	9·7	97·4
Methylhydroquinone (toluhydroquinone)	8·1	124

Sperm swelling was observed with a few compounds (Table 3) and was also seen with some compounds that at higher concentrations completely stripped the spermatozoa. These cases probably represent interpolation of molecules into the cell membrane before enough becomes present to cause rupture. At this stage the action of the spermicide would be reversible as washing would remove the interpolated molecules. A number of likely spermicides or cell rupturing agents were found to be without effect on sperm size, although some prevented the subsequent stripping by Zaponin reagent (Table 4).

Table 4. Compounds which have no effect on sperm size

No effect	Protection against subsequent stripping with Zaponin
Triton X-114 (Roth, Stuttgart)	Tween 20 (Serva)
Tergitol NP-40 (Sigma)	Tween 80 (Serva)
Polyvinylalcohol, Sigma Type II (Sigma)	Triton X-405 (Serva)
Brij 58 (Serva)	Pluronic F 68 (Serva)
Myrj 59 (Serva)	Monflor 71 (Serva)
Glycerol (Merck)	
Dioctylsulfosuccinic acid sodium salt (Serva)	
n-Dodecylguanidine acetate (Merck)	
2,7-Dimethylbenzene sulphonic acid (Serva)	
Nitrofurantoin + NaOH (Serva)	
Nitrofurazone + NaOH (Serva)	
Spermine tetrahydrochloride (Merck)	
α-Chlorohydrin (Merck)	
Ricinoleic acid, Pract. (Eastman)	
Dodecyltrimethylammonium chloride (Eastman)	
Benzyldimethylammonium chloride (Eastman)	
Trimethylammonium bromide (Eastman)	
Nonyltrimethylammonium bromide (Eastman)	

Discussion

The most potent spermicides belonged to several structural classes (Table 5). The strongest compounds were cationic detergents with a long carbon side chain, i.e. CTAB > Hyamine 10X > benzalkonium chloride > benzethonium chloride. Compounds in which the cationic quaternary nitrogen was part of a 6-membered ring, e.g. cetylpyridinium chloride and *N*-dodecylpyridinium chloride were as potent. Slight variations in the length and structure of the carbon chain brought about large variations in the potency of the compounds: Hyamine 10X contains only an extra ring methyl group compared with benzethonium chloride but was about 3 times more potent, and cetylpyridinium chloride contains 3 more methylene groups in the carbon side chain than does *N*-dodecylpyridinium chloride but is about 68 times more potent. When the carbon chain was reduced to a single methyl group, as in benzyltrimethylammonium chloride, stripping potency was very much reduced and incomplete.

Table 5. Precise nomenclature and structure of the most potent spermicides

Compound	Alternative name(s) with notes on stuctuure
Benzalkonium chloride	Alkylbenzyldimethylammonium chloride with the alkyls varying from C_8H_{17} to $C_{18}H_{37}$; Hyamine 3700 (Merck, Darmstadt: mol. wt approx. 365)
Benzethonium chloride	Benzyldiisobutylphenoxyethoxydimethylammonium chloride; diisobutylphenoxyethoxyethyldimethylbenzylammonium chloride; benzyldimethyl{2-[2-(*p*-1,1,3,3-tetramethylbutylphenoxy)ethoxy]ethyl}-ammonium chloride; Phemerol; Hyamine 1622
Cetylpyridinium chloride	*N*-Cetylpyridinium chloride monohydrate
CTAB	Cetyltrimethylammonium bromide; cetrimonium bromide; citrimide; *N*-cetyl-*N,N,N*-trimethylammonium bromide; hexadecyltrimethylammonium bromide
Hyamine 10X	Methylbenzethonium chloride; benzyldiisobutylcresoxyethoxydimethyl-ammonium chloride
N-Lauroylsarcosine sodium salt	Sodium lauroylsarcosinate; Sarcosyl; Gardol
Sodium lauryl sulphate	Sodium dodecylsulphate; SDS; dodecyl hydrogen sulphate sodium salt
Teepol 710	40% *secondary*-Alkyl sulphate sodium salt in water; Gardinol; Dupanol; Lissapol; Potency was calculated as if it had the same molecular weight as sodium lauryl sulphate
Triton X-100	*p*-Isooctylphenoxypolyethoxyethanol; Antarox CA-630; polyethyleneglycol-mono-*p*-(1,1,3,3-tetramethylbutyl)-phenyl ether; octylphenol polyethyleneglycol ether, $n = 9$–10; polyethyleneglycol *p*-isooctylphenyl ether; *p*-(1,1,3,3-tetramethylbutyl)phenoxypolyethyleneglycol; *o*-[4,(1,1,3,3-tetramethylbutyl)phenyl]deca(oxyethylene); diisobutylphenoxypolyethoxyethanol (= name used by Ortho Pharmaceuticals in U.K.)
Triton N-101	*p*-Nonylphenoxypolyethoxyethanol; Nonoxynol 9; polyoxyethylated nonyl-phenol; nonylphenolpolyethyleneglycol ether; polyethyleneglycol *p*-nonyl-phenol ether
Saponin (Merck, Darmstadt)	An extract from the 'white soap wort', *Gypsophila* sp. (Caryophyllaceae), containing gypsogenin (3β-hydroxy-23-oxoolean-12-en-28-oic acid), a triterpenoid, as the sapogenin, attached to a chain of sugars at the 3β-hydroxy group, consisting mainly of glucose with small amounts of arabinose, rhamnose and galactose. Potency was calculated by assuming that the sugar chain consisted of 9 molecules of glucose, thus giving a mol. wt of 2092. Note that this is not the same saponin defined under that generic name in the Merck Index (Merck, Sharpe & Dohme, U.S.A.)

Structural formulae may be obtained from the Merck Index. Only the most common trade names have been listed and many more are given in the Merck Index and are in use in various parts of the world. Commercially available vaginal contraceptive preparations usually contain one or more of these compounds under a proprietary trade name.

Anionic detergents, such as sodium lauryl sulphate, Teepol 710, *N*-lauroylsarcosine sodium salt and triisopropylnaphthalene sulphonic acid sodium salt, also showed great variations in potency with structure.

The greatest variations were shown by the neutral Triton detergents and the neutral bile salts. The side chain of Triton X-100 is the same as that of benzethonium chloride but it was about 7 times more potent, indicating the contribution of the polyethoxyethylene group. The length of this poly-ethoxyethylene group was important: for the iso-octylphenol compounds, $n = 9$–10 for Triton X-100, while $n = 7$–8 for Triton X-114 which had no effect on sperm size, and $n = 40$ for Triton X-405 which not only had no effect on sperm size but also prevented the subsequent stripping by Zaponin reagent. For the nonylphenol compounds, $n = 9$ for Triton N-101, while $n = 40$ for Tergitol NP-40 which had no effect on sperm size and was incompatible with Zaponin, producing a white precipitate. When the lengths of the molecules were the same, Triton N-101 was about twice as potent as Triton X-100.

In the bile salts, sodium deoxycholate was a potent spermicide but the reaction with the very closely related sodium cholate was very much reduced and did not proceed to complete stripping. Saponin (Merck, Darmstadt; also obtainable from B.D.H.) containing gypsogenin as the sapogenin was confirmed as one of the most potent cell lysing agents. Saponin (Roth, Stuttgart) was about 5 times less potent and did not induce complete stripping. The sapogenin is believed to be quillaic acid which has only one additional hydroxyl group on the triterpenoid molecule when compared with gypso-genin, while the carbohydrate side chain contains significant quantities of glucuronic and galacturonic acids rather than neutral sugars. Both preparations behaved differently from Zaponin reagent, in which the nature of the sapogenin has not been disclosed and which also contains acetic acid. Zaponin reagent produced full stripping in only 1 min, whereas it was necessary to wait for 5 min for the other saponin preparations, the only substances for which such a wait was necessary before measuring the degree of stripping. All the other compounds acted virtually instantaneously. The spermicidal action of other triterpenoid saponin preparations has also been demonstrated (Stolzenberg & Parkhurst, 1974; Setty, Kamboj, Garg & Khanna, 1976).

The steroidal and triterpenoid structures are in the form of plates that can easily intercalate between similar molecules in cell membranes. The structures of the other spermicidal detergents suggest a similar mode of action, which is purely physical in nature, whereby the cell membranes continue to incorporate more substance until they lose their structure. Sperm stripping with these agents was often a continuous process and after the removal of the peripheral cytoplasm, the sperm core may be gradually digested until the spermatozoa are completely dissolved. This process occurred quickly with sodium lauryl sulphate.

Detergents have been widely applied to the dissolution of cell membranes, both for the outer cell membrane and for the membranes around the various subcellular components (Helenius & Simons, 1975). Almost all the surfactants that are effective in solubilizing cell membranes have values for the hydrophilic:lipophilic balance (HLB) of 12·5–14·5; e.g. 13·5 for Triton X-100 and 13·4 for Triton N-101. Compounds with higher HLB values have also been used because they release mainly peri-pheral protein from membranes but do not dissociate the lamellar structure of the membrane. In the present study Brij 58 (HLB = 15·8), Tween 20 (HLB = 16·7), Tween 80 (HLB = 15·0) and Triton X-405 (HLB > 17) were ineffective as sperm stripping agents.

The spermicidal activities of some of the detergents tested have been previously measured as the lowest concentration needed to immobilize human spermatozoa after 30 min when a solution was mixed with an equal volume of semen (0·3 ml) (Harvey & Stuckey, 1962a). Spermicidal concentrations were 0·25–0·125 % for sodium lauryl sulphate, 0·03–0·025 % for benzalkonium chloride, 0·1–0·05 % for CTAB, 0·05–0·025 % for sodium dioctylsulphosuccinate; 0·5 % Tween 80 was non-lethal. It was not possible to confirm in this study the high activity of sodium dioctylsulphosuccinate: at maximum solubility in Isoton there was no significant effect on sperm size. This compound is, however, known to be a potent spermicide and it is possible that it acts thus by a different mechanism. Of the 581 compounds screened by Holzaepfel *et al.* (1959) there were 31 surfactants among the 56 most potent compounds. However, nomenclature is too imprecise to permit identification, although benz-ethonium chloride, benzalkonium chloride and Hyamine 10X were certainly present, probably with

Triton X-100 and Triton N-101. Of the non-detergents recognizable, I could obtain only 2-naphthol, 4-*n*-hexylresorcinol, ethyl potassium xanthate, sodium diethyldithiocarbamate and 2-methylhydroquinone. The first three were confirmed as potent spermicides although they did not strip to the core. Like 2,4,6–trichlorophenol and 6-chlorothymol these compounds are known to be potent bactericides and to digest bacterial membranes (Albert, 1973). Sodium diethyldithiocarbamate and 2-methylhydroquinone caused sperm swelling: the former was the most potent compound tested by Holzaepfel *et al.* (1959) and the dialkyldithiocarbamates in general are known to cause gross morphological changes in human spermatozoa (Rice, 1964).

Some spermicidal detergents have been used for the extraction of human and animal spermatozoa before biochemical investigation of specific components (Gall, Millette & Edelman, 1974; Zaneveld, Wagner, Schlumberger & Schumacher, 1974; Bedford, Bent & Calvin, 1973; Hernandez-Montes, Iglesias & Mujica, 1973) and this is currently an important area of sperm research.

Providing the formulation of the product ensures adequate availability of the active ingredient(s) to the spermatozoa, commercially available spermicides must be extremely effective. The safety of many of the compounds has also been demonstrated toxicologically (Smyth & Calandra, 1969). Triton N-101 is the sole active ingredient in several spermicidal creams, jellies and pessaries, e.g. Patentex Oval (Patentex, Germany), Delfen Cream, Delfen Foam (Ortho Pharmaceuticals, Cilag Chemie) and Duracreme (London Rubber Industries), while Triton X-100 occurs in Ortho-Gynol, Preceptin Gel, Efpa Gel (Ortho). Sodium lauryl sulphate is found with boric acid in Ortho-Creme. Many other commercial preparations contain two or more potent spermicidal detergents, e.g. benzethonium chloride is found with Triton N-101 in Orthoform Pessaries (Ortho) and Emko (Gerhardt, Germany). Many of the preparations are formulated at about pH 4·5 to accord with that of most of the vagina; an acid pH is itself detrimental to human spermatozoa but it is not necessary for the activity of the spermicides. Clinical trials have confirmed the effectiveness of the spermicidal detergent preparations. Tests with Patentex Oval foaming suppositories have shown a Pearl Index of 0·8 pregnancies per 100 women years for 63,759 cycles in 10,017 women (Rammstedt, 1974; Brehm & Haase, 1975). (For definition and discussion of the Pearl Index see Brotherton, 1976b.) Post-coital tests showed that only non-motile spermatozoa were present in the vagina and none was in the cervical canal. Delfen Cream and Preceptin Gel had a Pearl Index of 9·1 in a 6-year study in 980 women (Rovinsky, 1964). A vaginal foam containing 8 % Triton N-101 and 0·2 % benzethonium chloride had a Pearl Index of 3·98 (Bernstein, 1970). A vaginal foam tablet containing at least four potent spermicides has been shown to be effective (Ishihama & Inoue, 1972) and vaginal inserts containing benzalkonium chloride, Triton N-101 and sodium lauryl sulphate inhibit sperm motility very quickly (Chiari & Rappelli, 1975).

Ejaculated rabbit spermatozoa have been counted automatically after treatment with chloramine T (Kihlström, Carlsson & Larsson, 1975). Rabbit semen contains even more debris than that of man and it was not possible to count and size the spermatozoa without prior Zaponin treatment (Brotherton, 1975). In the present study chloramine T dissolved some of the debris in human semen but a loss of 10–20 % of the total sperm volume also occurred. Chloramine T is therefore a weak spermicide with stripping properties. Sodium hypochlorite has similar properties but is even weaker. Normal hygiene involving common soap (sodium lauryl sulphate) and swimming pool water (sodium hypochlorite) are sufficient to kill human spermatozoa.

Sperm size was not affected by α-chlorohydrin although the compound has been shown to inhibit motility and reduce metabolic activity (Hommonnai, Paz, Sofer, Yedwab & Kraicer, 1975; Mohri, Suter, Brown-Woodman, White & Ridley, 1975). Similarly the nitrofurans had no effect on sperm size although they have been shown to inhibit sperm motility *in vitro* (Albert, Mininberg & Davis, 1975; Albert, Salerno, Kapoor & Davis, 1975). It is clear that the stripping test does not apply to metabolic cell poisons, such as quinine and emetine also, whose mode of action is by the inhibition of some central biochemical process after they have entered the intact cell.

Quinine and emetine are currently being investigated by WHO as spermicidal agents for incorporation into intracervical devices. It is therefore pertinent to consider the results from the stripping assay in relation to the potency of these agents in cytocidal assays. In terms of absolute potency, the spermicides shown in Table 1 are about 1000 times less potent as cytocides than the best compounds

which act by inhibiting cell 'growth' or specific metabolic processes. The results from cytocidal assays against yeasts (Brotherton, 1976a) and trichomonads (Brotherton, 1977b) showed that quinine was not among the most effective compounds although emetine was very potent and that the non-specific and entirely physical mechanism of action of the stripping agents is a relatively weak method of killing cells compared with specific metabolic agents but is quicker.

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