Combating ovarian aging depends on the use of existing ovarian follicles, not on putative oogonial stem cells

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Abstract

Ovarian aging is characterized by both a reduction in egg quality and a drastic reduction in the number of ovarian follicles. It has been generally accepted for 60 years that a fixed population of primordial follicles is established in the ovaries during early life, and in most mammalian species, oocytes cannot renew themselves in postnatal or adult life. This dogma, however, has been challenged over the past decade. In this review, we summarize the recent studies on primordial follicles and putative oogonial stem cells and discuss what resources in the ovary might be more reliable and promising source tools for combating ovarian aging.

Introduction

Ovarian aging is characterized by both a reduction in egg quality and a drastic reduction in the total number of ovarian follicles (Broekmans et al. 2007). In mammals, a fixed population of primordial follicles is established in the ovaries during early life and serves as the source of developing follicles and oocytes during the entire reproductive life of the organism (Zuckerman 1951). Regardless of whether new follicles are formed during adult life or not, it is firmly established that ovarian aging occurs inevitably over the lifespan of the organism. The reduction of oocyte quality with aging is believed to be mainly due to an increase in meiotic nondisjunction that leads to an increased rate of aneuploidy in the early embryos (Nagaoka et al. 2012). As a woman ages, her pool of primordial follicles also gradually but steadily shrinks and menopause occurs when the number of primordial follicles falls to below about 1000 (Faddy et al. 1992). Despite the gradual loss of primordial follicles over time, their relative abundance makes them an attractive source of material for fighting ovarian aging.

The issue of ovarian aging and its associated effects on fertility has received greater attention as an increasing number of women in modern society choose to postpone the age at which they bear children. In addition, cancer-related mortality rates are decreasing due to advances in more effective treatments, but this has led to an increase in non-physiological ovarian aging because these treatments can adversely affect germ cell survival and cause premature ovarian failure (POF) and infertility (Jeruss & Woodruff 2009).

Primordial follicles: the natural source for fighting ovarian aging

Despite the primary importance of primordial follicles in determining the reproductive lifespan and the rate of ovarian aging, very little is known about the molecular mechanisms regulating their development. Very recent studies using genetically modified mouse models, however, have made significant contributions to our understanding of how the development of primordial follicles is regulated. For example, studies from Dr Diego Castrillon's laboratory at the University of Texas Southwestern Medical Center and Dr Kui Liu's laboratory at the University of Gothenburg in Sweden have independently demonstrated that phosphatase and tensin homolog deleted on chromosome ten (Pten), a negative regulator of the phosphatidylinositol 3 kinase (PI3K) signaling pathway, serves as an essential inhibitory molecule that suppresses the activation of primordial follicles (John et al. 2008, Reddy et al. 2008). Deletion of the Pten gene that inactivates the protein leads to accelerated follicular activation and a rapid loss of the entire primordial follicle pool that renders the Pten mutant females infertile at young adulthood. Further studies by Kui Liu's group with genetically modified mouse models revealed a series of factors in the PI3K and mTOR signaling pathways (e.g., Pdk1, Tsc1, and Tsc2) that are actively involved in the regulation of primordial follicle activation and survival (for reviews, see Adhikari & Liu 2009, 2010, Reddy et al. 2010 and Zheng et al. 2012). These findings clearly show that the majority of primordial follicles must be kept in a quiescent state for...
later use in order to maintain the normal length of the female reproductive lifespan and to prevent accelerated ovarian aging. These results also suggest that the activation of primordial follicles might be enhanced if the inhibition of follicular activation can be effectively removed (Fig. 1A).

Ovarian tissue is rich in primordial follicles, which make up the majority of the total follicular population in the human ovary. Pioneering studies from Dr John Eppig’s group in the Jackson Laboratory showed that live mice could be generated by in vitro culture of mouse neonatal ovaries that are rich in primordial follicles (Eppig & O’Brien 1996, O’Brien et al. 2003). These experiments suggest the possibility of utilizing primordial follicles for fertility treatment.

In prepubescent girls who suffer from POI, ovarian tissue cryopreservation is the only fertility-preserving option because neither ovarian stimulation and collection of mature oocytes nor obtaining fertilized embryos is feasible (Hovatta 2004). With the aim of fighting ovarian aging and preserving fertility, there has been an increasing trend recently to cryopreserve ovarian tissues from young women (West et al. 2009). One technique that might have potential for fertility preservation, therefore, is to obtain mature eggs from the cryopreserved primordial follicles.

Based on the earlier findings that Pten plays a crucial role in primordial follicle growth (John et al. 2008, Reddy et al. 2008), two recent studies have independently reported the first attempts to activate primordial follicles in vitro with a synthetic small molecule that suppresses the activities of Pten (Li et al. 2010, Adhikari et al. 2012). These studies showed that primordial follicles from postnatal mouse ovaries can be successfully activated by administration of the Pten inhibitor and that these activated follicles can develop into mature and fertilizable oocytes when transplanted under the kidney capsules of recipient mice. These oocytes can give rise to healthy progeny mice, suggesting that it is safe to use the Pten inhibitor for the activation of primordial follicles (Li et al. 2010, Adhikari et al. 2012). In vitro culture of human cortical tissues in the presence of the Pten inhibitor also led to the activation of primordial follicles that yielded mature oocytes upon prolonged transplantation under the kidney capsule of recipient mice (Li et al. 2010). These experiments have significant clinical implications for dealing with ovarian aging using pre-existing primordial follicles as a source for obtaining fertilizable oocytes (Fig. 1A).

‘Oogonial stem cells’ and postnatal folliculogenesis: more research is needed

It has been generally accepted for more than 60 years that in most mammalian species oocytes cannot renew themselves in postnatal life and that the number of primordial follicles in the ovary is fixed at birth (Zuckerman 1951). This dogma, however, has been challenged over the past decade. Beginning in 2004, a series of studies from Dr Jonathan Tilly’s laboratory at Harvard Medical School suggested that so-called ‘oogonial stem cells (OSCs)’ exist in postnatal ovaries. In 2004, Tilly’s group took their first shot against Zuckerman’s dogma (Johnson et al. 2004). By counting

![Figure 1](image-url)
follies, the authors proposed that primordial follicles in mice could be renewed at a rate of 77 follicles per day in each ovary. Moreover, they purported that a group of OSCs, which had originated from the epithelium of the ovarian surface, served as the source of the oocytes that fueled the follicular replenishment. Notably, these ‘OSCs’ were found to be mitotically active and were positive for the germline cell marker dead box polypeptide-4 (Ddx4). However, because many of the results in this study were confusing and could lead to divergent conclusions, critical comments were raised by other researchers immediately after the publication of this work (Albertini 2004, Gosden 2004, Telfer 2004). Several laboratories tried to repeat this work but they all failed to reproduce the same results (Bristol-Gould et al. 2006, Begum et al. 2008, Kerr et al. 2012).

One year later, in response to criticism from the field (Telfer et al. 2005), Tilly’s group amended their previous report and claimed that the OSCs had actually originated from the bone marrow and peripheral blood (Johnson et al. 2005). In this study, Tilly’s group reported that the cells from transplanted bone marrow or peripheral blood renewed the ovarian follicles and rescued the fertility of chemotherapy drug-sterilized female mice. Interestingly, the same germline cell marker, Ddx4, was employed in this work to identify the ‘germline potent’ bone marrow cells. Nevertheless, Johnson et al.’s postulation was soon refuted by another simple and straightforward study. Eggan et al. (2006) surgically joined the circulatory systems of a wild-type mouse and a mouse ubiquitously expressing green fluorescent protein (GFP) and found that no GFP-positive eggs were produced in the wild-type mouse (Eggan et al. 2006). Moreover, unpublished results from Kui Liu’s laboratory using tracing technology after bone marrow transplantation from fluorescent reporter mice also showed that there was no oocyte renewal from the transplanted bone marrow, which confirmed Eggan et al.’s results (Eggan et al. 2006).

A major study was published in 2009 by Dr Ji Wu’s laboratory at Shanghai Jiaotong University in China. Wu’s group claimed that they had isolated a population of OSCs (referred to as female germine stem cells in their study) from postnatal mouse ovaries by magnetic-activated cell sorting that was based on a polyclonal anti-human DDX4 antibody (Zou et al. 2009). These cells were 12–20 μm in diameter and were round in shape. They were positive for several germine cell markers and could be stably passed more than 68 times when cultured in vitro, features that are found in spermatogonial stem cells. Notably, Wu’s laboratory claimed that live mice were born from chemotherapy drug-sterilized female mice after these putative OSCs were transplanted into the ovaries of these mice (Zou et al. 2009).

No other groups were able to successfully reproduce Wu’s results and obtain functional OSCs from postnatal mouse ovaries until 2012 when Dr Tilly’s group published their isolation of mouse and human OSCs using the same DDX4 antibody-based cell-sorting system (White et al. 2012). Interestingly, the morphology of the OSCs obtained by Tilly seems to be distinct from the cells obtained by Wu. The isolated OSCs from Dr Tilly’s group were only 5–8 μm in diameter and were much smaller than embryonic primordial germ cells, male germline stem cells (10–20 μm in diameter) (Spiegelman & Bennett 1973), and the cells obtained by Dr Wu’s group (12–20 μm in diameter) (Zou et al. 2009).

Tilly’s group also claimed that they had obtained human follicles by xenotransplantation of human ovarian cortical tissues, which had been injected with human OSCs into immunodeficient female mice. They also claimed that mouse blastocysts had been obtained after transplantation of mouse OSCs into mouse ovaries. However, no functional human eggs or live mouse pups were generated in this work (White et al. 2012). Based on this finding, Dr Tilly claimed that ‘cells with cell surface expression of DDX4 that are present in the ovaries of reproductive age women represent adult human OSCs’ and that these ‘might be useful for new fertility applications’ (White et al. 2012).

This study soon attracted the attention of the mass media and patients, and it was believed that Tilly’s study would open new doors for treating female infertility and postponing menopause. In contrast to this over-optimism, however, other researchers voiced their concerns (Oatley & Hunt 2012) (http://www.bionews.org.uk/page_133180.asp). The major issue was the use of the human DDX4 antibody as the main tool to purify these purported OSCs. Ddx4/DDX4 is a cytoplasmic protein and is not known to be expressed on the cell surface (Tanaka et al. 2000). It is hard to understand, therefore, how the DDX4 antibody can bind to intracellular Ddx4/DDX4 protein and fish out the OSCs.

More recently, Dr Kui Liu’s laboratory has traced the development of Ddx4-positive cells in mouse ovaries using Rainbow mice, a fluorescent reporter mouse model (Zhang et al. 2012). This tracing method is different from antibody-based cell sorting. It does not purify the cells but instead looks directly at the endogenous promoter activity of the Ddx4 gene without any manipulation of the cells. Live-cell imaging showed that Ddx4-expressing cells in postnatal mouse ovaries did not proliferate when cultured in vitro, which was in direct contrast to the previously published studies (Zou et al. 2009, White et al. 2012). Recently, two new studies have confirmed the findings of Dr Liu. Using an inducible cell-lineage tracing method, Lei and Spradling from the Carnegie Institution for Science showed that no active ‘OSCs’ could be detected in adult mouse ovaries, and that the primordial follicle pool in a 4-week-old mouse ovary was sufficient to support fertility for the entire reproductive lifespan of the mouse (Lei & Spradling 2013). In a non-human primate species, Yuan et al. from the Nankai University in China detected the mitotic and germline cell markers in monkey ovaries at different age by the
immunostaining method, and found no proliferated germline stem cells in postnatal ovaries of rhesus monkeys (Yuan et al. 2013). These lines of evidence further support the notion that no OSCs exist in postnatal mammalian ovaries (Fig. 1B).

Two other studies have also reported the isolation of OSCs from postnatal mouse ovaries (Pacchiarotti et al. 2010, Hu et al. 2012). However, these putative OSCs could only form morphologically oocyte-like cells in vitro and no meiotically competent functional oocytes could be generated. Nevertheless, the capability to form such oocyte-like cells is not unique to the putative OSCs. Embryonic stem cells (Hubner et al. 2003) and somatic stem cells from other organs (Dyce et al. 2006, Danner et al. 2007, Linher et al. 2009) have also been reported to form non-functional oocyte-like cells or blastocyst-like structures during in vitro culture (Table 1). The nature of these purported OSCs, therefore, is highly questionable.

Based on the work to date, there is still a lack of comprehensive validation for the existence of OSCs in postnatal ovaries. Even if the OSCs do exist in postnatal mammalian ovaries, there is still a long way to go before female infertility could be treated by stem cell therapy. Thus, the resource we already have in the ovary – the existing follicles – is still the only hope that we can currently rely on for treating female infertility and fighting ovarian aging (Fig. 1).

### Perspectives

The molecular mechanisms controlling the activation of primordial follicles have started to be revealed, mostly using genetically modified mouse models. However, very little is known about the developmental dynamics of primordial follicles under physiological conditions. Pregranulosa cells provide a unique environment in which the oocytes of primordial follicles can remain dormant for long periods of time in mammalian ovaries, but the molecular mechanisms regulating the differentiation of pregranulosa cells into granulosa cells are largely unknown. The role of pregranulosa cells in the activation of primordial follicles is also yet to be identified, and in-depth studies of these issues will be of great value to our understanding of the process of ovarian aging.

Despite the shortcomings with primordial follicles, even more is unknown about the putative OSCs and there is a lack of direct evidence regarding the nature of these cells. The location of OSCs in the adult mammalian ovary, the mechanisms regulating the proliferation and differentiation of the OSCs in vivo, and the physiological function of the OSCs in female reproduction are all mysteries (Fig. 1B). There is still a long way to go before the existence of OSCs can be fully accepted by researchers in the field and before such cells can be safely used in clinical practice.

### Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the review.

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### References


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