

1 **Supplementary materials and methods**

2 ***Animals***

3 *R26-mTmG* female mice (double-fluorescent Cre reporter mice that express
4 membrane-targeted tandem dimmer TOMATO prior to Cre-mediated excision and
5 membrane-targeted green fluorescent protein (GFP) after excision (Muzumdar *et al.*
6 2007)) were crossed with *Vasa-Cre* male mice. Male offspring from this cross that
7 inherited the *Vasa-Cre* allele were collected at newborn stage and immunostained as
8 described below in order to verify the Cre recombinase activity.

9 The animal experiments were carried out in accordance with the Guidelines for Animal
10 Use and Experimentation of the University of Tokyo (approval IDs: P13-764, P14-876,
11 PM15-085, and PH16-084) and the Tokyo Medical and Dental University (approval
12 IDs: 0140007A, 0150259C2, 0160024C2, and 0170248C2).

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14 ***Whole-mount immunostaining***

15 Testes and epididymides of newborn mice were fixed in 4% paraformaldehyde. After
16 washing in phosphate-buffered saline, the samples were treated with CUBIC solution 1
17 (Susaki *et al.* 2014) for 1 week at 4°C to facilitate the visualization of immunoreactive
18 signals, and then blocked with Tris-buffered blocking solution. Subsequently, the
19 samples were incubated for 12 hours at 4°C with anti-VASA (1:1,000 dilution; Abcam,
20 Cambridge, United Kingdom) antibody. The immunoreactive signals were visualized by
21 using Alexa-680 conjugated secondary antibody (Molecular Probes) in combination
22 with the nuclear counterstaining of 4',6-diamidino-2-phenylindole (DAPI). The stained
23 samples were then transferred to CUBIC solution 2 (Susaki *et al.* 2014) and analyzed
24 using Leica TCS SP8 (Leica Microsystems GmbH) confocal laser microscope as

25 described previously (Aiyama *et al.* 2015).

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28 **Supplementary references**

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31 the Terminal Segments of the Seminiferous Tubules in Hamster Testes. *Stem cells* **33**

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33 **Muzumdar MD, Tasic B, Miyamichi K, Li L & Luo L** 2007 A global double-

34 fluorescent Cre reporter mouse. *Genesis* **45** 593-605.

35 **Susaki EA, Tainaka K, Perrin D, Kishino F, Tawara T, Watanabe TM, Yokoyama**

36 **C, Onoe H, Eguchi M, Yamaguchi S, Abe T, Kiyonari H, Shimizu Y, Miyawaki A,**

37 **Yokota H & Ueda HR** 2014 Whole-brain imaging with single-cell resolution using

38 chemical cocktails and computational analysis. *Cell* **157** 726-739.

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